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Request for grant of a patent

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1. Your reference — RVCW / P26781GB

2. Patent application number *(The Patent Office will fill in this part)* — 0217434.0

UL02 E736668-7 002866

3. Full name, address and postcode of the or of each applicant *(underline all surnames)*

Royal Veterinary College
University of London
Royal College Street
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United Kingdom

Patents ADP number *(if you know it)* 7974983002

If the applicant is a corporate body, give the country/state of its incorporation — United Kingdom

4. Title of the invention

BIOLOGICAL MATERIAL

5. Name of your agent *(if you have one)*

"Address for service" in the United Kingdom to which all correspondence should be sent *(including the postcode)*

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Patents ADP number *(if you know it)* **1305010**

6. If you are declaring priority from one or more earlier patent applications, give the country and the date of filing of the or of each of these earlier applications and *(if you know it)* the or each application number

Country	Priority application number *(if you know it)*	Date of filing *(day / month / year)*

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Number of earlier application	Date of filing *(day / month / year)*

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Abstract	1
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Statement of inventorship and right to grant of a patent *(Patents Form 7/77)*	NO
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11. I/We request the grant of a patent on the basis of this application.

Signature E Potter Clarkson — Date 26 July 2002

ERIC POTTER CLARKSON

12. Name and daytime telephone number of person to contact in the United Kingdom — **0115 9552211**

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BIOLOGICAL MATERIAL

The present invention relates to biological material, and in particular to a canine respiratory coronavirus that is present in dogs having canine infectious respiratory disease.

Canine infectious respiratory disease (CIRD) is a highly contagious disease of dogs housed in crowded conditions such as re-homing centres and boarding or training kennels. Many dogs suffer only from a mild cough and recover after a short time, however in some cases a severe bronchopneumonia can develop (Appel and Binn, 1987).

The pathogenesis of CIRD is considered to be multifactorial, involving several viruses and bacteria. The infectious agents considered to be the major causative pathogens of CIRD are canine parainfluenzavirus (CPIV) (Binn *et al.*, 1967), canine adenovirus type 2 (CAV-2) (Ditchfield *et al.*, 1962) and the bacterium *Bordetella bronchiseptica* (Bemis *et al.*, 1977, Keil *et al.*, 1998). Also, canine herpesvirus, human reovirus and mycoplasma species have been isolated from dogs with symptoms of CIRD (Karpas *et al.*, 1968, Lou and Wenner 1963, Randolph *et al.*, 1993) Additional factors like stress may also be important.

CIRD is rarely fatal but it delays re-homing of dogs at rescue centres and it causes disruption of schedules in training kennels as well as considerable treatment costs.

Vaccines are available against some of the infectious agents associated with this disease, namely *Bordetella bronchiseptica* as well as CPIV and CAV-2. However, despite the use of these vaccines, CIRD is still prevalent in kennels world-wide, which is possibly due to the vaccines not providing protection against all the infectious agents involved in CIRD.

We have discovered a novel coronavirus, which we have called canine respiratory coronavirus (CRCV), in a large kennelled dog population with a history of endemic respiratory disease, and we have shown that this virus is associated with CIRD.

Some members of the family *coronaviridae* are known to cause respiratory disease in humans, cattle, swine and poultry (Mäkelä *et al.*, 1998, Pensaert *et al.*, 1986, Ignjatovic and Sapats 2000). For example, bovine respiratory coronavirus is associated with shipping fever in cattle which is a multifactorial respiratory disease (Storz *et al.*, 2000).

However, coronaviruses were not suspected to have a role in the pathogenesis of CIRD. Indeed, with only a single exception, canine coronaviruses have been reported to be enteric viruses and to cause acute diarrhoea mainly in young dogs (for example, Tennant *et al.*, 1993). In a large study of viruses involved in canine respiratory diseases, Binn *et al.* (1979) reported the detection of a canine coronavirus in the lung of a single dog that was also infected with SV5 and canine adenovirus II, two other viruses that are associated with canine respiratory disease.

There are 30-40 dog vaccines commercially available in the UK for use against a number of pathogens that can cause a range of diseases, such as neurological, enteric, hepatic and respiratory diseases. Most of these vaccines contain microbial agents such as Distemper virus, Canine Adenovirus-2, Canine parvovirus, canine influenza virus and *Leptospira canicola* and *L. icterohaemorrhagiae.* None of these vaccines contain canine coronaviruses.

The only dog vaccine solely for use against respiratory diseases of which we are aware is Intrac, which is marketed as a vaccine for "kennel-cough".

Intrac is a freeze-dried live vaccine containing modified *Bordetella bronchisepticum*, which is a bacterium associated with "kennel cough".

Coyne M.J. & May S.W., (1995) in their article entitled "Considerations in using a canine coronavirus vaccine" (published as a Pfizer Technical Bulletin on the Internet at http://www.pfizer.com/ah/vet/tref/trbull/ccv.html), lists over 20 commercially available vaccines against either canine coronaviruses alone or against canine coronaviruses together with other organisms. Each of these vaccines is for canine enteric disease, and there is no suggestion that a canine coronavirus may be associated with respiratory disease.

US Patents Nos. 6,057,436 and 6,372,224, both to Miller *et al* and assigned to Pfizer, Inc., describe the spike gene of the enteric canine coronavirus and uses therefor, including its use as a vaccine against gastroenteritis. Neither of these two patents suggest that a canine coronavirus may be involved in CIRD.

Members of the family *coronaviridae* are enveloped viruses, 80-160nm in diameter, containing a linear positive-stranded RNA genome. The structural proteins of coronaviruses are the spike glycoprotein (S), the membrane glycoprotein (M) and the nucleocapsid protein. The hemagglutinating esterase glycoprotein (HE) is found only on the surface of group II coronaviruses (e.g. bovine coronavirus and murine hepatitis virus) (Spaan *et al,* 1988). Further details of the structure of coronoviruses may be found in the chapter by Cavanagh *et al* entitled "Coronviridae" p407-411, in "Virus Taxonomy, 6th Report of the International Committee on Taxonomy of Viruses", pub. Springer-Verlag Wein, New York, Eds. Murphy *et al*, which is incorporated herein by reference.

The canine respiratory coronavirus (CRCV) of the invention may be characterised as a coronavirus present in the respiratory tracts of dogs with infectious respiratory disease. To further characterise CRCV, we have determined the sequence of 250 nucleotide residues of the CRCV polymerase (pol) cDNA (Figure 1) which corresponds to an 83 amino acid partial sequence of the pol protein (Figure 2). We have also cloned and determined the sequence of the 4092 nucleotide residues of the CRCV spike (S) cDNA (Figure 3), corresponding to 1363 amino acids (Figure 4). We have identified that CRCV has a surprisingly low homology to the enteric canine coronavirus (CCV) while it has an unexpectedly high level of homology to bovine coronavirus (strain LY138 or Quebec) and human coronavirus (strain OC43).

A culture of "Spike D-1 CRCV", which is XL1-Blue *E. coli* (Stratagene) containing a pT7Blue2 plasmid (Novagen) whose insert contains a portion of the CRCV spike cDNA, has been deposited under the Budapest Treaty at NCIMB Ltd under Accession number NCIMB 41146 on 25 July 2002. The depositor of NCIMB 41146 is the Royal Veterinary College, Royal College Street, London NW1 OTU, UK. The address of NCIMB Ltd is 23 St. Machar Drive, Aberdeen, Scotland, AB24 3RY, UK.

The phylogenetic relationship of CRCV to eleven known coronaviruses was determined based upon a comparison of the 250 nucleotide sequence from the CRCV pol gene and the corresponding regions of the other viruses (Figure 5). The bovine coronavirus (BCV), human coronavirus (HCV) strain OC43 and hemagglutinating encephalomyelitis virus (HEV) were found to be most closely related to CRCV, while the enteric CCV was found to be only distantly related to CRCV.

Over the 250 sequenced residues of the pol cDNA, corresponding to 83 amino acids, CRCV has only 68.5% and 75.9% sequence identity at the

nucleotide and amino acid levels, respectively, with the equivalent region of the enteric CCV (strain 1-71) pol gene (Genbank Accession No. AF124986), as shown in Figure 6 and 7.

Except as described below, the percentage identity between two nucleotide or two amino acid sequences was determined using FASTA version 34 (Pearson WR. (1990) "Rapid and sensitive sequence comparison with FASTP and FASTA". *Methods Enzymol.*;183:63-98). FASTA settings were Gap open penalty -16 and Gap extension penalty –4.

The percentage identity between the CRCV and enteric CCV spike sequences was determined using GCG version 10 (Genetics Computer Group, (1991), Program Manual for the GCG Package, Version 7, April 1991, 575 Science Drive, Madison, Wisconsin, USA 53711). The GCG parameters used were: Gap creation penalty 50, gap extension penalty 3 for DNA, and Gap creation penalty 8 and Gap extension penalty 2 for Protein.

Sequence alignments were performed using ClustalX (Thompson *et al.*, 1997).

Over the 4092 sequenced nucleotide residues of the CRCV S gene, corresponding to 1363 amino acids, CRCV has 45% and 21.2% sequence identity at the nucleotide (Figure 8) and amino acid levels, respectively, with the equivalent region of the enteric CCV (strain 1-71) S gene.

By contrast, over the 250 sequenced residues of the pol cDNA, CRCV has 98.8% sequence identity with the equivalent region of the BCV strain Quebec pol gene (Genbank Accession No. AF220295), 98.4% sequence identity with the BCV strain LY138 pol gene (Genbank Accession No. AF124985) and 98.4% sequence identity with the HCV OC43 pol gene (Genbank Accession No. AF124989).

There was only a single amino acid difference between the CRCV pol protein over the 83 sequenced amino acids and the BCV, HCV and HEV pol proteins which is that CRCV has E (Glu) as opposed to D (Asp) at the position corresponding to position 4975 in the BCV genome (Accession No. SWALL: Q91A29). Thus the CRCV pol protein is 99% identical to the BCV, HCV and HEV pol proteins over this region.

The one and three letter amino acid codes of the IUPAC-IUB Biochemical Nomenclature Commission are used herein.

Over the 4092 sequenced nucleotide residues, corresponding to 1363 amino acids, of the CRCV S gene, CRCV has 97.3% and 96% identity with the equivalent region of BCV strain LY138 (Genbank Accession No. AF058942) at the nucleotide and amino acid levels respectively. CRCV has 96.9% (nucleotide) and 95.2% (amino acid) identity with HCV strain OC43 (Genbank Accession No. Z32768), and 83.8% (nucleotide) and 80.395% (amino acid) identity with HEV (Genbank Accession Nos. AF481863 (cDNA) and AAM 77000 (protein)) as shown in Figures 9 and 10.

The amino acids that are different between the CRCV S protein and each of the BCV, HCV and HEV S proteins, within the 1363 amino acids of the CRCV S protein, are listed in Table 1 below. Thus the amino acids listed in Table 1 could be said to be CRCV S protein-specific amino acids. The amino acids are numbered from the initial M residue at the start of the CRCV protein, as shown in Figure 4.

Table 1: List of 39 amino acids specific to the CRCV S protein that are not present in the BCV, HCV and HEV S proteins.

Position	Amino acid
103	V
118	V
166	D
171	M
179	K
192	P
210	S
235	H
267	F
388	F
407	M
436	S
440	I
447	I
501	F
525	Y
528	N
540	L
582	K
608	G

Position	Amino acid
692	G
695	S
757	W
758	G
763	Q
769	T
786	P
792	H
818	R
827	P
828	V
887	F
933	D
977	F
1011	T
1018	S
1063	K
1256	L
1257	M

A first aspect of the invention provides a coronavirus S protein, or fragment thereof, having at least 75% amino acid sequence identity with the CRCV S protein whose amino acid sequence is listed in Figure 4, and having at least one of V at position 103; V at position 118; D at position 166; M at position 171; K at position 179; P at position 192; S at position 210; H at position 235; F at position 267; F at position 388; M at position 407; S at position 436; I at position 440; I at position 447; F at position 501; Y at position 525; N at position 528; L at position 540; K at position 582; G at position 608; G at position 692; S at position 695; W at position 757; G at position 758; Q at position 763; T at position 769; P at position 786; H at position 792; R at position 818; P at position 827; V at position 828; F at position 887; D at position 933; F at position 977; T at position 1011; S at position 1018; K at position 1063; L at position 1256; and M at position 1257. The amino acids are numbered from the initial M at the start of the CRCV S protein, as listed in Figure 4.

It is appreciated that the partial nucleotide sequence of CRCV S can be readily determined by a person or ordinary skill in the art by sequencing the insert of the plasmid contained in *E. coli* strain D-1 CRCV, that has been deposited under the Budapest Treaty at NCIMB Ltd. under Accession number NCIMB 41146 on 25 July 2002. Furthermore, this DNA can be used as a hybridisation probe, or as the basis for the design of probes, in the isolation of CRCV nucleic acid in dogs.

For the avoidance of doubt, the invention includes a coronavirus S protein, or fragment thereof, having at least 75% amino acid sequence identity with the CRCV S protein, and comprising at least one of the amino acids specific for the CRCV S protein at the position listed in Table 1.

By "protein" we also include the meaning glycoprotein. The amino acid sequence of a glycoprotein refers to the amino acid sequence of the

polypeptide backbone of the glycoprotein, irrespective of the type, number, sequence and position of the sugars attached thereto.

Typically, the invention includes an isolated or recombinant protein, and not an unmodified CRCV protein present as a CRCV component.

The invention includes a coronavirus S protein, or fragment thereof, having at least 76% amino acid sequence identity with the CRCV S protein, or at least 77%, or at least 78%, or at least 79%, or at least 80%, or at least 81%, or at least 82%, or at least 83%, or at least 84%, or at least 85%, or at least 86%, or at least 87%, or at least 88%, or at least 89%, or at least 90%, or at least 91%, or at least 92%, or at least 93%, or at least 94%, or at least 95%, or at least 96%, or at least 97%, or at least 98%, or at least 99% amino acid sequence identity with the CRCV S protein, and comprising at least one of the amino acids specific for the CRCV S protein at the position listed in Table 1.

The invention also includes a coronavirus S protein, or fragment thereof, having at least 75%, or at least 80%, or at least 85% or at least 90% or at least 95% amino acid sequence identity with the CRCV S protein, and comprising at least 2, or at least 3, or at least 4, or at least 5, or at least 6, or at least 7, or at least 8, or at least 9, or at least 10, or at least 11, or at least 12, or at least 13, or at least 14, or at least 15, or at least 16, or at least 17, or at least 18, or at least 19, or at least 20, or at least 21, or at least 22, or at least 23, or at least 24, or at least 25, or at least 26, or at least 27, or at least 28, or at least 29, or at least 30, or at least 31, or at least 32, or at least 33, or at least 34, or at least 35, or at least 36, or at least 37, or at least 38 of the amino acids specific for CRCV S protein at the positions listed in Table 1.

Preferably, the coronavirus S protein, or fragment thereof comprises all 39 of the amino acid residues specific for CRCV S protein at the positions listed in Table 1.

Thus the invention includes a BCV, HCV or HEV S protein or fragment thereof, that has been modified at at least one position listed in Table 1 to resemble the CRCV S protein.

Preferably, the coronavirus S protein of the invention is a CRCV S protein that comprises or consists of the sequence listed in Figure 4, or a variant thereof with at least 97% identity with the sequence listed in Figure 4. Preferably, the variant has at least 98%, or at least 99% amino acid sequence identity with the sequence listed in Figure 4. More preferably the variant has at least 99.1%, or at least 99.2%, or at least 99.3%, or at least 99.4%, or at least 99.5%, or at least 99.6%, or at least 99.7%, or at least 99.8%, or at least 99.9% amino acid sequence identity with the sequence listed in Figure 4.

Thus the variant of the coronavirus S protein of the invention includes a protein that comprises or consists of the sequence listed in Figure 4 but has between 1 and 40 amino acid differences from the sequence listed in Figure 4. Preferably, the variant has less than 40 amino acid differences from the sequence listed in Figure 4. More preferably the variant has less than 35, less than 30, or less than 25, or less than 20, or less than 15, or 10 or 9 or 8 or 7 or 6 or 5 or 4 or 3 or 2 amino acid differences, or a single amino acid difference, from the sequence listed in Figure 4.

The invention also includes a CRCV S protein fragment comprising a fragment of the sequence listed in Figure 4 which comprises at least one of the amino acids specific for CRCV S protein at the position listed in Table 1.

The invention includes a coronavirus S protein, or fragment thereof, having at least 75% amino acid sequence identity with BCV strain LY138 S protein (Genbank Accession No. AF058942), and comprising at least one of V at position 103; V at position 118; D at position 166; M at position 171; K at position 179; P at position 192; S at position 210; H at position 235; F at position 267; F at position 388; M at position 407; S at position 436; I at position 440; I at position 447; ; F at position 501; Y at position 525; N at position 528; L at position 540; K at position 582; G at position 608; G at position 692; S at position 695; W at position 757; G at position 758; Q at position 763; T at position 769; P at position 786; H at position 792; R at position 818; P at position 827; V at position 828; F at position 887; D at position 933; F at position 977; T at position 1011; S at position 1018; K at position 1063; L at position 1256 and M at position 1257.

For the avoidance of doubt, the invention includes a coronavirus S protein, or fragment thereof, having at least 75% amino acid sequence identity with BCV strain LY138 S protein, and comprising at least one of the amino acids specific for CRCV S protein at the position listed in Table 1.

The invention includes a coronavirus S protein, or fragment thereof, having at least 76% amino acid sequence identity with BCV strain LY138 S protein, or at least 77%, or at least 78%, or at least 79%, or at least 80%, or at least 81%, or at least 82%, or at least 83%, or at least 84%, or at least 85%, or at least 86%, or at least 87%, or at least 88%, or at least 89%, or at least 90%, or at least 91%, or at least 92%, or at least 93%, or at least 94%, or at least 95%, or at least 96%, or at least 97%, or at least 98%, or at least 99% amino acid sequence identity with BCV strain LY138 S protein, and having at least one of the amino acids specific for CRCV S protein at the position listed in Table 1.

The invention also includes a coronavirus S protein, or fragment thereof, having at least 75%, or at least 80%, or at least 85%, or at least 90%, or at least 95% amino acid sequence identity with BCV strain LY138 S protein, and comprising at least 2, or at least 3, or at least 4, or at least 5, or at least 6, or at least 7, or at least 8, or at least 9, or at least 10, or at least 11, or at least 12, or at least 13, or at least 14, or at least 15, or at least 16, or at least 17, or at least 18, or at least 19, or at least 20, or at least 21, or at least 22, or at least 23, or at least 24, or at least 25, or at least 26, or at least 27, or at least 28, or at least 29, or at least 30, or at least 31, or at least 32, or at least 33, or at least 34, or at least 35, or at least 36, or at least 37, or at least 38 of the amino acids specific for CRCV S protein at the positions listed in Table 1.

Preferably, the coronavirus S protein, or fragment thereof comprises all 39 of the amino acid residues specific for CRCV S protein at the positions listed in Table 1.

A second aspect of the invention provides a coronavirus pol protein, or fragment thereof, having at least 90% amino acid sequence identity with the BCV pol protein and comprising the amino acid E at the position corresponding to position 4975 in the BCV genome (Accession No. SWALL: Q91A29).

The invention includes a coronavirus pol protein, or fragment thereof, having at least 91% amino acid sequence identity with BCV strain LY138 pol protein, or at least 92%, or at least 93%, or at least 94%, or at least 95%, or at least 96%, or at least 97%, or at least 98%, or at least 99% amino acid sequence identity with BCV strain LY138 pol protein, and having the amino acid E at the position corresponding to position 4975 in the BCV genome (Accession No. SWALL: Q91A29).

Preferably, the coronavirus pol protein, or fragment thereof is a CRCV pol protein or fragment that thereof comprises or consists of the amino acid sequence listed in Figure 2.

Thus the invention includes a BCV, HCV or HEV pol protein or fragment thereof, that has been modified at the amino acid corresponding to position 4975 in the BCV genome, to resemble the CRCV pol protein.

The invention also includes a CRCV pol protein fragment comprising a fragment of the sequence listed in Figure 2 and having the amino acid E at the position corresponding to position 4975 in the BCV genome.

The coronavirus S or pol proteins as defined above in the first and second aspect of the invention may be termed herein "CRCV" or "CRCV-like" proteins.

A "CRCV S protein" is an S protein or fragment thereof that has the native CRCV S amino acid sequence as listed in Figure 4, or a fragment thereof which comprises at least one of the amino acids specific for a CRCV S protein at the positions listed in Table 1.

A "CRCV pol protein" is a pol protein or fragment thereof that has the native CRCV pol amino acid sequence as listed in Figure 2, or a fragment thereof which comprises the amino acid E at the position corresponding to position 4975 in the BCV genome.

A "CRCV-like S protein" is an S protein or fragment thereof that does not have an amino acid sequence identical to the native CRCV S amino acid sequence (Figure 4), but has at least 75% sequence identity with the corresponding region of the CRCV or BCV strain LY138 S protein, and has at least one of the amino acids specific for a CRCV S protein at the positions listed in Table 1.

A "CRCV-like S protein" also includes an S protein that does not have an amino acid sequence identical to the native CRCV S amino acid sequence (Figure 4), but that comprises or consists of a variant of the sequence listed in Figure 4 with at least 97% identity with the sequence listed in Figure 4. Preferably, the variant has at least 98%, or at least 99% amino acid sequence identity with the sequence listed in Figure 4. More preferably the variant has at least 99.1%, or at least 99.2%, or at least 99.3%, or at least 99.4%, or at least 99.5%, or at least 99.6%, or at least 99.7%, or at least 99.8%, or at least 99.9% amino acid sequence identity with the sequence listed in Figure 4.

A "CRCV-like pol protein" is a pol protein or fragment thereof that does not have an amino acid sequence identical to the native CRCV pol amino acid sequence, but has at least 90% sequence identity with the corresponding BCV strain LY138 pol protein, and which has an E at the position corresponding to position 4975 in the BCV genome.

Preferably, the CRCV or CRCV-like protein, or fragment thereof, is at least 10 amino acids in length. More preferably, the CRCV or CRCV-like protein, or fragment thereof, is at least 20, or at least 30, or at least 40, or at least 50, or at least 100, or at least 200, or at least 300, or at least 400, or at least 500, or at least 600, or at least 700, or at least 800, or at least 900, or at least 1,000, or at least 1,100, or at least 1,200 amino acids in length.

Preferably, the CRCV or CRCV-like protein, or fragment thereof, is less than about 1,300 amino acids in length. More preferably, the CRCV or CRCV-like protein, or fragment thereof, is less than about 1,200, or less than about 1,100, or less than about 1,000, or less than about 900, or less than about 800, or less than about 700, or less than about 600, or less than about 500, or less than about 400, or less than about 300, or less than about 200, or less than about 100, or less than about 50 amino acids in length.

CRCV proteins may be isolated from CRCV, or may be made using protein chemistry techniques for example using partial proteolysis of isolated proteins (either exolytically or endolytically), or by *de novo* synthesis. Alternatively, the CRCV proteins, as well as CRCV-like proteins, may be made by recombinant DNA technology. Suitable techniques for cloning, manipulation, modification and expression of nucleic acids, and purification of expressed proteins, are well known in the art and are described for example in Sambrook *et al* (2001) "*Molecular Cloning, a Laboratory Manual*", 3rd edition, Sambrook *et al* (eds), Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, USA, incorporated herein by reference.

Shorter fragments of CRCV and CRCV-like proteins, *ie* peptides, may be synthesised using standard techniques. Peptides may be synthesised by the Fmoc-polyamide mode of solid-phase peptide synthesis as disclosed by Lu *et al* (1981) *J. Org. Chem.* **46**, 3433 and references therein. Temporary N-amino group protection is afforded by the 9-fluorenylmethyloxycarbonyl (Fmoc) group. Repetitive cleavage of this highly base-labile protecting group is effected using 20% piperidine in N,N-dimethylformamide. Side-chain functionalities may be protected as their butyl ethers (in the case of serine threonine and tyrosine), butyl esters (in the case of glutamic acid and aspartic acid), butyloxycarbonyl derivative (in the case of lysine and histidine), trityl derivative (in the case of cysteine) and 4-methoxy-2,3,6-trimethylbenzenesulphonyl derivative (in the case of arginine). Where glutamine or asparagine are C-terminal residues, use is made of the 4,4'-dimethoxybenzhydryl group for protection of the side chain amido functionalities. The solid-phase support is based on a polydimethyl-acrylamide polymer constituted from the three monomers dimethylacrylamide (backbone-monomer), bisacryloylethylene diamine (cross linker) and acryloylsarcosine methyl ester (functionalising agent). The peptide-to-resin

cleavable linked agent used is the acid-labile 4-hydroxymethyl-phenoxyacetic acid derivative. All amino acid derivatives are added as their preformed symmetrical anhydride derivatives with the exception of asparagine and glutamine, which are added using a reversed N,N-dicyclohexyl-carbodiimide/1-hydroxybenzotriazole mediated coupling procedure. All coupling and deprotection reactions are monitored using ninhydrin, trinitrobenzene sulphonic acid or isotin test procedures. Upon completion of synthesis, peptides are cleaved from the resin support with concomitant removal of side-chain protecting groups by treatment with 95% trifluoroacetic acid containing a 50% scavenger mix. Scavengers commonly used are ethanedithiol, phenol, anisole and water, the exact choice depending on the constituent amino acids of the peptide being synthesised. Trifluoroacetic acid is removed by evaporation *in vacuo*, with subsequent trituration with diethyl ether affording the crude peptide. Any scavengers present are removed by a simple extraction procedure which on lyophilisation of the aqueous phase affords the crude peptide free of scavengers. Reagents for peptide synthesis are generally available from Calbiochem-Novabiochem (UK) Ltd, Nottingham NG7 2QJ, UK. Purification may be effected by any one, or a combination of, techniques such as size exclusion chromatography, ion-exchange chromatography and (principally) reverse-phase high performance liquid chromatography. Analysis of peptides may be carried out using thin layer chromatography, reverse-phase high performance liquid chromatography, amino-acid analysis after acid hydrolysis and by fast atom bombardment (FAB) mass spectrometric analysis.

A third aspect of the invention provides a polynucleotide that encodes a CRCV or CRCV-like S or pol protein according to the first and second aspects of the invention, or the complement thereof.

Preferably, the polynucleotide encodes a CRCV S protein according to the first aspect of the invention, or the complement thereof.

More preferably, the polynucleotide encoding the CRCV S protein comprises or consists of the sequence listed in Figure 3.

It is appreciated that the sequence listed in Figure 3 contains a Y at position 3531, which refers to either C or T. In both cases the corresponding amino acid is Ile. Thus the invention includes a polynucleotide encoding a CRCV S protein which comprises or consists of the sequence listed in Figure 3, and having C at position 3531. The invention also includes a polynucleotide encoding a CRCV S protein which comprises or consists of the sequence listed in Figure 3, and having T at position 3531.

The invention also includes a CRCV S polynucleotide comprising a fragment of the sequence listed in Figure 3, that encodes a protein having at least one of the amino acids specific for CRCV S protein at the position listed in Table 1, or the complement thereof.

Preferably, the polynucleotide encoding the pol protein comprises or consists of the sequence listed in Figure 1, or the complement thereof.

The invention also includes a CRCV pol polynucleotide comprising a fragment of the sequence listed in Figure 1 that encodes a protein having E at the position corresponding to position 4975 in the BCV genome, or the complement thereof.

The polynucleotides as defined above are referred to herein as CRCV or CRCV-like polynucleotides of the invention.

A "CRCV-like polynucleotide" is a polynucleotide that does not have a base sequence identical to all or a fragment of the native CRCV cDNA

sequence as listed in Figures 1 and 3, but that encodes a CRCV or CRCV-like S or pol protein as defined above, or the complement thereof.

The CRCV is a positive strand RNA virus. The polynucleotide of the invention may be DNA or RNA. The RNA may be positive or negative strand RNA. The DNA may be single or double stranded DNA.

Suitable techniques for cloning and sequencing a cDNA from a positive strand RNA virus such as CRCV are well known in the art and are described for example in Sambrook *et al* 2001, incorporated herein by reference.

The CRCV or CRCV-like polynucleotides of the invention may be any suitable size. However, for certain purposes, such as probing or amplifying, it is preferred if the nucleic acid has fewer than 3,000, more preferably fewer than 1000, more preferably still from 10 to 100, and in further preference from 15 to 30 base pairs (if the nucleic acid is double-stranded) or bases (if the nucleic acid is single stranded). As is described more fully below, single-stranded DNA oligonucleotides, suitable for use as hybridisation probes or as primers in a polymerase chain reaction, are particularly preferred.

Oligonucleotides that can specifically amplify, or hybridise to CRCV S or pol polynucleotides, as opposed to BCV, HCV or HEV S or pol polynucleotides, are particularly preferred. Suitable oligonucleotides can be determined by a person of skill in the art by reference to the nucleotide sequence comparisons in Figures 6, 8 and 9.

It is appreciated that the CRCV or CRCV-like oligonucleotides may, even under highly stringent conditions, hybridise to nucleic acid, whether RNA or DNA, from HCV, BCV, and HEV as well as from CRCV. However, it is

preferred if the CRCV or CRCV-like oligonucleotides hybridise to nucleic acid from CRCV under more stringent conditions than to nucleic acid from HCV, BCV or HEV. This can either be determined experimentally or by a comparison of the oligonucleotide sequence with the respective CRCV, HCV, BCV and HEV sequences, as is well known to one of skill in the art (Sambrook *et al* 2001).

Conveniently, the CRCV or CRCV-like polynucleotides or oligonucleotides further comprise a detectable label.

By "detectable label" is included any convenient radioactive label such as ^{32}P, ^{33}P or ^{35}S which can readily be incorporated into a nucleic acid molecule using well known methods; any convenient fluorescent or chemiluminescent label which can readily be incorporated into a nucleic acid is also included. In addition the term "detectable label" also includes a moiety which can be detected by virtue of binding to another moiety (such as biotin which can be detected by binding to streptavidin); and a moiety, such as an enzyme, which can be detected by virtue of its ability to convert a colourless compound into a coloured compound, or *vice versa* (for example, alkaline phosphatase can convert colourless *o*-nitrophenylphosphate into coloured *o*-nitrophenol). Conveniently, the nucleic acid probe may occupy a certain position in a fixed array and whether a nucleic acid hybridises to it can be determined by reference to the position of hybridisation in the fixed array.

Labelling with [^{32}P]dCTP may be carried out using a Rediprime® random primer labelling kit supplied by Amersham.

Primers which are suitable for use in a polymerase chain reaction (PCR; Saiki *et al* (1988) *Science* 239, 487-491) are preferred. Suitable PCR primers may have the following properties:

It is well known that the sequence at the 5′ end of the oligonucleotide need not match the target sequence to be amplified.

It is usual that the PCR primers do not contain any complementary structures with each other longer than 2 bases, especially at their 3′ ends, as this feature may promote the formation of an artefactual product called "primer dimer". When the 3′ ends of the two primers hybridise, they form a "primed template" complex, and primer extension results in a short duplex product called "primer dimer".

Internal secondary structure should be avoided in primers. For symmetric PCR, a 40-60% G+C content is often recommended for both primers, with no long stretches of any one base. The classical melting temperature calculations used in conjunction with DNA probe hybridization studies often predict that a given primer should anneal at a specific temperature or that the 72°C extension temperature will dissociate the primer/template hybrid prematurely. In practice, the hybrids are more effective in the PCR process than generally predicted by simple T_m calculations.

Optimum annealing temperatures may be determined empirically and may be higher than predicted. *Taq* DNA polymerase does have activity in the 37-55°C region, so primer extension will occur during the annealing step and the hybrid will be stabilised. The concentrations of the primers are equal in conventional (symmetric) PCR and, typically, within 0.1- to 1μM range.

It will further be appreciated that if a control amplification reaction is to be carried out, for example using primers complementary to an ubiquitously expressed protein, that it may be beneficial for the products of the control and CRCV or CRCV-like products to be of different sizes, such that the two products may be distinguished by the detection means employed, for

example by mobility on agarose gel electrophoresis. However, it may be desirable for the two products to be of similar size, for example both between 100 and 1000, or between 100 and 600 nucleotides long. This may aid simultaneous analysis of the products, for example by gel electrophoresis, and may also mean that the control and CRCV or CRCV-like amplification reactions may have similar performance characteristics, in terms, for example, of relative rates of accumulation of product at different stages during the reaction.

Any of the nucleic acid amplification protocols can be used in the method of the invention including the polymerase chain reaction, QB replicase and ligase chain reaction. Also, NASBA (nucleic acid sequence based amplification), also called 3SR, can be used as described in Compton (1991) *Nature* 350, 91-92 and *AIDS* (1993), Vol 7 (Suppl 2), S108 or SDA (strand displacement amplification) can be used as described in Walker *et al* (1992) *Nucl. Acids Res.* 20, 1691-1696. The polymerase chain reaction is particularly preferred because of its simplicity.

When a pair of suitable nucleic acids of the invention are used in a PCR it is convenient to detect the product by gel electrophoresis and ethidium bromide staining. As an alternative, it is convenient to use a labelled oligonucleotide capable of hybridising to the amplified DNA as a probe. When the amplification is by PCR the oligonucleotide probe hybridises to the interprimer sequence as defined by the two primers. The oligonucleotide probe is preferably between 10 and 50 nucleotides long, more preferably between 15 and 30 nucleotides long. It may be longer than the amplified DNA or include one or both of the primers, but in this case, the hybridisation conditions should be such that the probe should not hybridise to the primers alone, but only to an amplified product that also contains interprimer sequence that is capable of hybridising to the probe.

The probe may be labelled with a radionuclide such as ^{32}P, ^{33}P and ^{35}S using standard techniques, or may be labelled with a fluorescent dye. When the oligonucleotide probe is fluorescently labelled, the amplified DNA product may be detected in solution (see for example Balaguer *et al* (1991) "Quantification of DNA sequences obtained by polymerase chain reaction using a bioluminescence adsorbent" *Anal. Biochem.* 195, 105-110 and Dilesare *et al* (1993) "A high-sensitivity electrochemiluminescence-based detection system for automated PCR product quantitation" *BioTechniques* 15, 152-157.

PCR products can also be detected using a probe which may have a fluorophore-quencher pair or may be attached to a solid support or may have a biotin tag or they may be detected using a combination of a capture probe and a detector probe.

Fluorophore-quencher pairs are particularly suited to quantitative measurements of PCR reactions (eg RT-PCR). Fluorescence polarisation using a suitable probe may also be used to detect PCR products.

The invention also includes a vector comprising the CRCV or CRCV-like polynucleotide of the third aspect of the invention.

Typical prokaryotic vector plasmids are: pUC18, pUC19, pBR322 and pBR329 available from Biorad Laboratories (Richmond, CA, USA); p*Trc*99A, pKK223-3, pKK233-3, pDR540 and pRIT5 available from Pharmacia (Piscataway, NJ, USA); pBS vectors, Phagescript vectors, Bluescript vectors, pNH8A, pNH16A, pNH18A, pNH46A available from Stratagene Cloning Systems (La Jolla, CA 92037, USA).

A typical mammalian cell vector plasmid is pSVL available from Pharmacia (Piscataway, NJ, USA). This vector uses the SV40 late promoter to drive

expression of cloned genes, the highest level of expression being found in T antigen-producing cells, such as COS-1 cells. An example of an inducible mammalian expression vector is pMSG, also available from Pharmacia (Piscataway, NJ, USA). This vector uses the glucocorticoid-inducible promoter of the mouse mammary tumour virus long terminal repeat to drive expression of the cloned gene.

Useful yeast plasmid vectors are pRS403-406 and pRS413-416 and are generally available from Stratagene Cloning Systems (La Jolla, CA 92037, USA). Plasmids pRS403, pRS404, pRS405 and pRS406 are Yeast Integrating plasmids (YIps) and incorporate the yeast selectable markers *HIS3, TRP1, LEU2* and *URA3*. Plasmids pRS413-416 are Yeast Centromere plasmids (YCps).

Generally, the CRCV or CRCV-like polynucleotide of the invention is inserted into an expression vector, such as a plasmid, in proper orientation and correct reading frame for expression. It may be linked to the appropriate transcriptional and translational regulatory control nucleotide sequences recognised by the desired host prior to insertion into the vector, although such controls are generally available in the expression vector. Thus, the polynucleotide of the invention insert may be operatively linked to an appropriate promoter. Eukaryotic promoters include the CMV immediate early promoter, the HSV thymidine kinase promoter, the early and late SV40 promoters and the promoters of retroviral LTRs. Other suitable promoters will be known to the skilled artisan. The expression constructs desirably also contain sites for transcription initiation and termination, and in the transcribed region, a ribosome binding site for translation. (Hastings *et al*, International Patent No. WO 98/16643, published 23 April 1998)

Methods well known to those skilled in the art can be used to construct expression vectors containing the coding sequence and, for example

appropriate transcriptional or translational controls. One such method involves ligation via homopolymer tails. Homopolymer polydA (or polydC) tails are added to exposed 3' OH groups on the DNA fragment to be cloned by terminal deoxynucleotidyl transferases. The fragment is then capable of annealing to the polydT (or polydG) tails added to the ends of a linearised plasmid vector. Gaps left following annealing can be filled by DNA polymerase and the free ends joined by DNA ligase.

Another method involves ligation via cohesive ends. Compatible cohesive ends can be generated on the DNA fragment and vector by the action of suitable restriction enzymes. These ends will rapidly anneal through complementary base pairing and remaining nicks can be closed by the action of DNA ligase.

A further method uses synthetic molecules called linkers and adaptors. DNA fragments with blunt ends are generated by bacteriophage T4 DNA polymerase or *E.coli* DNA polymerase I which remove protruding 3' termini and fill in recessed 3' ends. Synthetic linkers, pieces of blunt-ended double-stranded DNA which contain recognition sequences for defined restriction enzymes, can be ligated to blunt-ended DNA fragments by T4 DNA ligase. They are subsequently digested with appropriate restriction enzymes to create cohesive ends and ligated to an expression vector with compatible termini. Adaptors are also chemically synthesised DNA fragments which contain one blunt end used for ligation but which also possess one preformed cohesive end.

Synthetic linkers containing a variety of restriction endonuclease sites are commercially available from a number of sources including International Biotechnologies Inc, New Haven, CN, USA.

A desirable way to modify the polynucleotide of the invention is to use the polymerase chain reaction as disclosed by Saiki *et al* (1988) *Science* **239**, 487-491. In this method the DNA to be enzymatically amplified is flanked by two specific oligonucleotide primers which themselves become incorporated into the amplified DNA. The specific primers may contain restriction endonuclease recognition sites which can be used for cloning into expression vectors using methods known in the art.

The invention also includes a host cell transformed with the vector comprising the CRCV or CRCV-like polynucleotide. The host cell can be either prokaryotic or eukaryotic. If the CRCV or CRCV-like polynucleotide, in the vector, is to be expressed as a glycoprotein, the host cell is a eukaryotic host cell, and preferably a mammalian host cell.

Bacterial cells are preferred prokaryotic host cells and typically are a strain of *E. coli* such as, for example, the *E. coli* strains DH5 available from Bethesda Research Laboratories Inc., Bethesda, MD, USA, and RR1 available from the American Type Culture Collection (ATCC) of Rockville, MD, USA (No ATCC 31343). Preferred eukaryotic host cells include yeast and mammalian cells, preferably vertebrate cells such as those from a mouse, rat, monkey or human fibroblastic cell line. Yeast host cells include YPH499, YPH500 and YPH501 which are generally available from Stratagene Cloning Systems, La Jolla, CA 92037, USA. Preferred mammalian host cells include Chinese hamster ovary (CHO) cells available from the ATCC as CCL61, NIH Swiss mouse embryo cells NIH/3T3 available from the ATCC as CRL 1658, and monkey kidney-derived COS-1 cells available from the ATCC as CRL 1650. Preferred insect cells are Sf9 cells which can be transfected with baculovirus expression vectors.

Transformation of appropriate cell hosts with a vector is accomplished by well known methods that typically depend on the type of vector used. With

regard to transformation of prokaryotic host cells, see, for example, Cohen *et al* (1972) *Proc. Natl. Acad. Sci. USA* **69**, 2110 and Sambrook *et al* (2001) *Molecular Cloning, A Laboratory Manual*, 3rd Ed. Cold Spring Harbor Laboratory, Cold Spring Harbor, NY. Transformation of yeast cells is described in Sherman *et al* (1986) *Methods In Yeast Genetics, A Laboratory Manual*, Cold Spring Harbor, NY. The method of Beggs (1978) *Nature* **275**, 104-109 is also useful. With regard to vertebrate cells, reagents useful in transfecting such cells, for example calcium phosphate and DEAE-dextran or liposome formulations, are available from Stratagene Cloning Systems, or Life Technologies Inc., Gaithersburg, MD 20877, USA.

Electroporation is also useful for transforming cells and is well known in the art for transforming yeast cell, bacterial cells and vertebrate cells.

For example, many bacterial species may be transformed by the methods described in Luchansky *et al* (1988) *Mol. Microbiol.* **2**, 637-646 incorporated herein by reference. The greatest number of transformants is consistently recovered following electroporation of the DNA-cell mixture suspended in 2.5x PEB using 6250V per cm at 25μFD.

Methods for transformation of yeast by electroporation are disclosed in Becker & Guarente (1990) *Methods Enzymol.* **194**, 182.

Physical methods may be used for introducing DNA into animal and plant cells. For example, microinjection uses a very fine pipette to inject DNA molecules directly into the nucleus of the cells to be transformed. Another example involves bombardment of the cells with high-velocity microprojectiles, usually particles of gold or tungsten that have been coated with DNA.

Successfully transformed cells, ie cells that contain a CRCV or CRCV-like DNA construct, can be identified by well known techniques. For example, one selection technique involves incorporating into the expression vector a DNA sequence (marker) that codes for a selectable trait in the transformed cell. These markers include dihydrofolate reductase, G418 or neomycin resistance for eukaryotic cell culture, and tetracyclin, kanamycin or ampicillin resistance genes for culturing in *E.coli* and other bacteria. Alternatively, the gene for such selectable trait can be on another vector, which is used to co-transform the desired host cell.

The marker gene can be used to identify transformants but it is desirable to determine which of the cells contain recombinant DNA molecules and which contain self-ligated vector molecules. This can be achieved by using a cloning vector where insertion of a DNA fragment destroys the integrity of one of the genes present on the molecule. Recombinants can therefore be identified because of loss of function of that gene.

Another method of identifying successfully transformed cells involves growing the cells resulting from the introduction of an expression construct of the present invention to produce the CRCV or CRCV-like S or pol protein. Cells can be harvested and lysed and their DNA content examined for the presence of the DNA using a method such as that described by Southern (1975) *J. Mol. Biol.* **98**, 503 or Berent *et al* (1985) *Biotech.* 3, 208. Alternatively, the presence of the protein in the supernatant can be detected using antibodies as described below.

In addition to directly assaying for the presence of recombinant DNA, successful transformation can be confirmed by well known immunological methods when the recombinant DNA is capable of directing the expression of the protein. For example, cells successfully transformed with an expression vector produce proteins displaying appropriate antigenicity. Samples of cells

suspected of being transformed are harvested and assayed for the protein using suitable antibodies.

Thus, in addition to the transformed host cells themselves, the present invention also contemplates a culture of those cells, preferably a monoclonal (clonally homogeneous) culture, or a culture derived from a monoclonal culture, in a nutrient medium.

Host cells that have been transformed by the recombinant CRCV or CRCV-like polynucleotide, typically in a vector as described above, are then cultured for a sufficient time and under appropriate conditions known to those skilled in the art in view of the teachings disclosed herein to permit the expression of the CRCV or CRCV-like protein encoded by the CRCV or CRCV-like polynucleotide, which can then be recovered.

The CRCV or CRCV-like protein can be recovered and purified from recombinant cell cultures by well-known methods including ammonium sulphate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite chromatography and lectin chromatography. Most preferably, high performance liquid chromatography ("HPLC") is employed for purification.

For example, for expression in a baculovirus system, recombinant DNA encoding the CRCV spike gene may be cloned into a suitable transfer vector such as pMelBac (Invitrogen). Co-transfection with baculovirus DNA (eg Bac-N-Blue/Invitrogen) results in a recombinant baculovirus encoding the spike gene. Infection of a suitable insect cell line (e.g. Sf9, Sf21, High Five/Invitrogen) at an appropriate multiplicity of infection leads to expression of the recombinant spike protein. Protein expression is

confirmed by western blotting or ELISA using appropriate reagents (e.g. convalescent canine serum or other virus specific antiserum).

The invention thus includes a method of obtaining a CRCV or CRCV-like protein encoded by the CRCV or CRCV-like polynucleotide of the present invention. The method comprises culturing the host cell comprising the CRCV or CRCV-like polynucleotide, typically in a vector; expressing the protein in the host cell, and purifying the protein. The invention further includes the protein obtainable by this method.

The invention thus also includes a method of obtaining a glycosylated CRCV or CRCV-like protein, typically an S protein, encoded by the CRCV or CRCV-like polynucleotide of the present invention. The method comprises culturing a eukaryotic, or more preferably mammalian, host cell comprising the CRCV or CRCV-like polynucleotide, typically in a vector; expressing the protein in the host cell; and purifying the glycosylated protein. The invention further includes the glycosylated protein obtainable by this method.

In a fourth aspect, the invention provides a method of making an anti-CRCV antibody comprising raising an immune response to a CRCV or CRCV-like S protein of the invention as described above in the first aspect of the invention in an animal, and preparing an antibody from the animal or from an immortal cell derived therefrom. Alternatively, the method may comprise selecting an antibody from an antibody-display library using a CRCV or CRCV-like S protein of the invention as described above in the first aspect of the invention.

Methods and techniques for producing a monoclonal antibody are well known to a person of skill in the art, for example those disclosed in *"Monoclonal Antibodies: A manual of techniques"*, H Zola (CRC Press,

1988) and in *"Monoclonal Hybridoma Antibodies: Techniques and Applications"*, J G R Hurrell (CRC Press, 1982), incorporated herein by reference.

Optionally, the method further comprises determining whether the antibody thus obtained has greater affinity for the CRCV S protein than for the BCV S protein, and preferably also whether the antibody has a greater affinity for the CRCV S protein than for the HCV and HEV S proteins. Methods for determining the relative affinity of antibodies for antigens are known in the art.

The invention also includes an anti-CRCV antibody obtainable by the method of the fourth aspect of the invention, that has greater affinity for the CRCV S protein than for the BCV S protein. Preferably, the antibody also has a greater affinity for the CRCV S protein than for the HCV and HEV S proteins. Preferably, the antibody is a monoclonal antibody. However, the invention includes a monospecific anti-CRCV antibody. The antibody may be an antibody fragment, as described below.

The monoclonal or monospecific antibody may be a chimaeric antibody, as discussed by Neuberger *et al* (1988, *8th International Biotechnology Symposium* Part 2, 792-799). The monoclonal or monospecific antibody may also be a "caninised" antibody, for example by inserting the CDR regions of mouse antibodies into the framework of canine antibodies.

The invention also includes anti-CRCV antibody fragments. The variable heavy (V_H) and variable light (V_L) domains of antibodies are involved in antigen recognition, a fact first recognised by early protease digestion experiments. Further confirmation was found by "humanisation" of rodent antibodies, in which variable domains of rodent origin may be fused to constant domains of human origin such that the resultant antibody retains the

antigenic specificity of the rodent parented antibody (Morrison *et al* (1984) *Proc. Natl. Acad. Sci. USA* **81**, 6851-6855).

That antigenic specificity is conferred by variable domains and is independent of the constant domains is known from experiments involving the bacterial expression of antibody fragments, all containing one or more variable domains.

These molecules include Fab-like molecules (Better *et al* (1988) *Science* **240**, 1041); Fv molecules (Skerra *et al* (1988) *Science* **240**, 1038); single-chain Fv (ScFv) molecules where the V_H and V_L partner domains are linked via a flexible oligopeptide (Bird *et al* (1988) *Science* **242**, 423; Huston *et al* (1988) *Proc. Natl. Acad. Sci. USA* **85**, 5879) and single domain antibodies (dAbs) comprising isolated V domains (Ward *et al* (1989) *Nature* **341**, 544). A general review of the techniques involved in the synthesis of antibody fragments which retain their specific binding sites is to be found in Winter & Milstein (1991) *Nature* **349**, 293-299.

By "ScFv molecules" we mean molecules wherein the V_H and V_L partner domains are linked via a flexible oligopeptide.

The advantages of antibody fragments, rather than whole antibodies, are several-fold. The smaller size of the fragments may lead to improved pharmacological properties, such as better penetration of solid tissue. Effector functions of whole antibodies, such as complement binding, are removed. Fab, Fv, ScFv and dAb antibody fragments can all be expressed in and secreted from *E. coli*, thus allowing the facile production of large amounts of the fragments.

Whole antibodies, and $F(ab')_2$ fragments are "bivalent". By "bivalent" we mean that the antibodies and $F(ab')_2$ fragments have two antigen combining

sites. In contrast, Fab, Fv, ScFv and dAb fragments are monovalent, having only one antigen combining sites.

In a fifth aspect, the invention provides a method of determining whether a dog has been exposed to CRCV. The method comprises obtaining a suitable sample from the dog, and identifying CRCV or an anti-CRCV antibody in the sample.

The invention includes a method of detecting, in a sample obtained from a dog, past exposure of the dog to CRCV, the method comprising obtaining a suitable sample from the dog, and identifying anti-CRCV antibodies in the sample.

In one preferred embodiment, the suitable sample can be any antibody containing sample such as serum, saliva, tracheal wash or bronchiolar lavage.

Preferably, the anti-CRCV antibody can be detected using a BCV, HCV, HEV or CRCV antigen, more preferably, using a BCV or CRCV antigen.

More preferably, identifying an anti-CRCV antibody in the sample comprises identifying an antibody that selectively binds to an S protein whose amino acid sequence is at least 75% identical with the amino acid sequence of the CRCV S protein (Figure 4); an S protein whose amino acid sequence is at least 75% identical with the amino acid sequence of the BCV S protein (Genbank Accession No. AF058942); HCV S protein (Genbank Accession No. L14643); to a coronavirus having an S protein at least 75% identical with BCV S protein (Genbank Accession No. AF058942), or a fragment thereof; or to a coronavirus having an S protein at least 75% identical with the CRCV S protein, or a fragment thereof.

More preferably, identifying an antibody that selectively binds to an S protein whose amino acid sequence is at least 75% identical with the amino acid sequence of the BCV S protein, comprises identifying an antibody that selectively binds to an S protein whose amino acid sequence is at least 80% identical, or at least 85% identical, or at least 90% identical, or at least 95% identical with the amino acid sequence of the BCV S protein (Genbank Accession No. AF058942) or a fragment thereof.

More preferably, identifying an anti-CRCV antibody in the sample comprises identifying an antibody that selectively binds to the BCV S protein (Genbank Accession No. AF058942).

Even more preferably, identifying an antibody that selectively binds to an S protein whose amino acid sequence is at least 75% identical with the amino acid sequence of the CRCV S protein, comprises identifying an antibody that selectively binds to an S protein whose amino acid sequence is at least 80% identical, or at least 85% identical, or at least 90% identical, or at least 95% identical with the amino acid sequence of the CRCV S protein (Figure 4) or a fragment thereof.

Yet more preferably, identifying an anti-CRCV antibody in the sample comprises identifying an antibody that selectively binds to a CRCV or CRCV-like S protein as defined in the first aspect of the invention.

Most preferably, identifying an anti-CRCV antibody in the sample comprises identifying an antibody that selectively binds to the CRCV S protein as listed in Figure 4, or a fragment thereof.

The invention includes a method of detecting CRCV in a sample obtained from a dog, the method comprising obtaining a suitable sample from the dog, and identifying CRCV in the sample.

It is appreciated that there may be some naturally occurring sequence variation between different isolates of CRCV. The invention thus includes identifying CRCV isolates whose S and pol genes and proteins have some sequence variation from the sequences provided in Figures 1 to 4. It is appreciated, however, that the same methods will be used to detect the variant isolates of CRCV as well as the isolate characterised by the sequences listed in Figures 1 to 4.

In a preferred embodiment, the suitable sample can be a lung wash, tracheal wash, tonsillar swab or a biopsy or post-mortem sample from the respiratory tract of the dog.

Preferably, in this embodiment, identifying CRCV comprises identifying a nucleic acid component of CRCV.

Typically, this will be performed by extracting RNA from the sample, and obtaining cDNA therefrom, for example as is described in Example 1. Thereafter, a CRCV nucleic acid component is identified in the cDNA, for example using techniques involving high stringency hybridisation, specific amplification, and nucleotide sequencing, as are well known to a person of skill in the art (Sambrook *et al* (2001) *supra*).

Preferably, identifying CRCV comprises identifying a polynucleotide that hybridises at high stringency to the BCV genome, such as the LY138 strain genome (Genbank Accession No. AF058942) or a portion thereof.

Further preferably, identifying CRCV comprises identifying a polynucleotide that hybridises at high stringency to the CRCV S or pol polynucleotides (Figures 1 and 3) or a portion thereof.

By "hybridising at high stringency" is meant that the polynucleotide and the nucleic acid to which it hybridises have sufficient nucleotide sequence

similarity that they can hybridise under highly stringent conditions. As is well known in the art, the stringency of nucleic acid hybridisation depends on factors such as length of nucleic acid over which hybridisation occurs, degree of identity of the hybridising sequences and on factors such as temperature, ionic strength and CG or AT content of the sequence.

Nucleic acids which can hybridise at high stringency to the CRCV cDNA molecule include nucleic acids which have >90% sequence identity, preferably those with >95% or >96% or >97% or >98, more preferably those with >99% sequence identity, over at least a portion of the CRCV cDNA.

Typical highly stringent hybridisation conditions which lead to selective hybridisation are known in the art, for example those described in Sambrook *et al* 2001 (*supra*), incorporated herein by reference.

An example of a typical hybridisation solution when a nucleic acid is immobilised on a nylon membrane and the probe nucleic acid is≥ 500 bases is:

6 x SSC (saline sodium citrate)

0.5% sodium dodecyl sulphate (SDS)

100 μg/ml denatured, fragmented salmon sperm DNA

The hybridisation is performed at 68°C. The nylon membrane, with the nucleic acid immobilised, may be washed at 68°C in 0.1 x SSC.

20 x SSC may be prepared in the following way. Dissolve 175.3 g of NaCl and 88.2 g of sodium citrate in 800 ml of H_2O. Adjust the pH to 7.0 with a few drops of a 10 N solution of NaOH. Adjust the volume to 1 litre with H_2O. Dispense into aliquots. Sterilise by autoclaving.

An example of a typical hybridisation solution when a nucleic acid is immobilised on a nylon membrane and the probe is an oligonucleotide of between 15 and 50 bases is:

3.0 M trimethylammonium chloride (TMACl)

0.01 M sodium phosphate (pH 6.8)

1 mm EDTA (pH 7.6)

0.5% SDS

100 µg/ml denatured, fragmented salmon sperm DNA

0.1% non-fat dried milk

The optimal temperature for hybridisation is usually chosen to be 5°C below the T_i for the given chain length. T_i is the irreversible melting temperature of the hybrid formed between the probe and its target sequence. Jacobs *et al* (1988) *Nucl. Acids Res.* **16**, 4637 discusses the determination of T_is. The recommended hybridization temperature for 17-mers in 3M TMACl is 48-50°C; for 19-mers, it is 55-57°C; and for 20-mers, it is 58-66°C.

Preferably, identifying CRCV comprises using a polynucleotide having at least 80%, or at least 85%, or at least 90%, or at least 95% identity with a portion of the BCV genome (Genbank Accession No. AF058942).

More preferably, identifying CRCV comprises using a polynucleotide having at least 80%, or at least 85%, or at least 90%, or at least 95% identity with a portion of the CRCV S polynucleotide (Figure 3), or having at least 90%, or at least 95% identity with a portion of the CRCV pol polynucleotide (Figure 1).

More preferably, identifying CRCV comprises identifying a CRCV polynucleotide as defined above with respect to the third aspect of the invention.

Most preferably, identifying CRCV comprises identifying a CRCV polynucleotide comprising or consisting of a sequence listed in Figure 1 or Figure 3, or a fragment thereof.

In another preferred embodiment, identifying CRCV comprises identifying a protein component of CRCV.

Preferably, identifying a protein component of CRCV comprises identifying a CRCV protein as defined above in the first or second aspects of the invention.

Most preferably, identifying a protein component of CRCV comprises identifying a CRCV protein comprising or consisting of the amino acid sequence listed in Figure 2 or Figure 4, or a fragment thereof.

Assaying a protein component of CRCV in a biological sample can occur using any art-known method. Preferred for assaying CRCV protein levels in a biological sample are antibody-based techniques.

Preferably, identifying a protein component of CRCV comprises using an antibody reactive with CRCV.

More preferably, the antibody reactive with CRCV is an anti-BCV antibody, an anti-HCV antibody, an anti-HEV antibody, or an anti-CRCV antibody obtainable by the methods of the fourth aspect of the invention.

For example, CRCV protein expression can be studied with classical immunohistological methods. In these, the specific recognition is provided

by the primary antibody (polyclonal or monoclonal) but the secondary detection system can utilise fluorescent, enzyme, or other conjugated secondary antibodies. As a result, an immunohistological staining of tissue section for pathological examination is obtained. Tissues can also be extracted, e.g., with urea and neutral detergent, for the liberation of CRCV protein for Western-blot or dot/slot assay (Jalkanen, M., *et al, J. Cell. Biol.* 101:976-985 (1985); Jalkanen, M., *et al, J. Cell. Biol.* 105:3087-3096 (1987)). In this technique, which is based on the use of cationic solid phases, quantitation of CRCV protein can be accomplished using isolated CRCV protein as a standard. This technique can also be applied to body fluid samples.

Other antibody-based methods useful for detecting CRCV protein expression include immunoassays, such as the enzyme linked immunosorbent assay (ELISA) and the radioimmunoassay (RIA). For example, a CRCV reactive monoclonal antibody can be used both as an immunoadsorbent and as an enzyme-labeled probe to detect and quantify the CRCV protein. The amount of CRCV protein present in the sample can be calculated by reference to the amount present in a standard preparation using a linear regression computer algorithm. Such an ELISA for detecting a tumour antigen is described in Iacobelli *et al, Breast Cancer Research and Treatment* 11: 19-30 (1988). In another ELISA assay, two distinct specific monoclonal antibodies can be used to detect CRCV protein in a body fluid. In this assay, one of the antibodies is used as the immunoadsorbent and the other as the enzyme-labeled probe.

The above techniques may be conducted essentially as a "one-step" or "two-step" assay. The "one-step" assay involves contacting CRCV protein with immobilized antibody and, without washing, contacting the mixture with the labeled antibody. The "two-step" assay involves washing before

contacting the mixture with the labeled antibody. Other conventional methods may also be employed as suitable. It is usually desirable to immobilize one component of the assay system on a support, thereby allowing other components of the system to be brought into contact with the component and readily removed from the sample.

Suitable enzyme labels include, for example, those from the oxidase group, which catalyze the production of hydrogen peroxide by reacting with substrate. Glucose oxidase is particularly preferred as it has good stability and its substrate (glucose) is readily available. Activity of an oxidase label may be assayed by measuring the concentration of hydrogen peroxide formed by the enzyme-labeled antibody/substrate reaction. Besides enzymes, other suitable labels include radioisotopes, such as iodine (125I, 121I), carbon (14C), sulfur 35S), tritium (3H), indium (112In), and technetium (99mTc), and fluorescent labels, such as fluorescein and rhodamine, and biotin.

In a sixth aspect, the invention provides an immunosorbent assay for detecting anti-CRCV S antibodies. The assay comprises a solid phase coated with a CRCV or CRCV-like S protein as defined in the first aspect of the invention, or obtainable using the methods of the third aspect of the invention, or an antigenic fragment thereof, wherein anti-CRCV S antibodies in a sample exposed to the solid phase will bind to the protein; and a detectable label conjugate which will bind to the anti-CRCV antibodies bound to the solid phase.

It is appreciated that an antigenic fragment of the CRCV or CRCV-like S protein that coats the solid phase is of sufficient size to be bound by an anti-CRCV S antibody, and which comprises at least one of the amino acids specific for CRCV S protein as listed in Table 1.

Preferably, the CRCV or CRCV-like S protein, or antigenic fragment thereof, that coats the solid phase is at least 10 amino acids in length. More preferably, the CRCV or CRCV-like S protein, or antigenic fragment thereof, is at least 20, or at least 30, or at least 40, or at least 50, or at least 100, or at least 200, or at least 300, or at least 400, or at least 500, or at least 600, or at least 700, or at least 800, or at least 900, or at least 1,000 amino acids in length.

Preferably, the CRCV or CRCV-like S protein, or antigenic fragment thereof, that coats the solid phase is less than about 1200 amino acids in length. More preferably, the CRCV or CRCV-like S protein, or antigenic fragment thereof, is less than about 1,100, or less than about 1,000, or less than about 900, or less than about 800, or less than about 700, or less than about 600, or less than about 500, or less than about 400, or less than about 300, or less than about 200, or less than about 100, or less than about 50 amino acids in length.

Preferably, the solid phase is a microtitre well.

Further preferably, the conjugate comprises anti-dog antibody.

Preferably, the conjugate comprises an enzyme, for example horseradish peroxidase. Further preferably, the immunosorbent assay also comprises a substrate for the enzyme.

Further details of suitable immunosorbent assays and ELISAs are provided above.

The invention includes a kit of parts which include the components of the immunosorbent assay. The kit of parts may thus include a solid phase such as a microtitre plate, CRCV or CRCV-like S protein for coating the solid phase, a detectable label conjugate, such as an anti-dog antibody, which will

bind to anti-CRCV antibodies bound to the solid phase. If the detectable label conjugate is an enzyme, the kit of parts may also include a substrate for the enzyme. The kit may also include a positive control sample that contains an anti-CRCV S protein antibody, such as those described with reference to the fourth aspect of the invention, and a negative control sample.

The invention thus includes a solid substrate with a CRCV or CRCV-like S protein as defined in the first aspect of the invention, or obtainable using the methods of the third aspect of the invention, or an antigenic fragment thereof, attached thereto, for capturing anti-CRCV S antibodies from a liquid sample, wherein anti-CRCV S antibodies in a sample exposed to the solid substrate will bind to the S protein.

Typically, protein is coated on microtitre plates overnight at 4°C to 37°C, depending on the stability of the antigen. Unbound protein is washed off with a wash buffer such as phosphate buffered saline or Tris buffered saline. Serum or other samples are incubated on the plate, typically at 37°C for between 1 and several hours. Unbound material is washed off, the plates are incubated with enzyme-labelled (e.g. horseradish peroxidase) antibody, such as anti-canine IgG or IgM for serum samples, or anti-canine IgA for lung washes, for 1 to several hours at 37°C. Unbound antibody is washed off and plates are incubated with a substrate such as OPD for about 10 min, and the optical density measured in a photometer.

Preferably, the solid substrate is a microtitre well.

In a seventh aspect, the invention provides a vaccine composition for vaccinating dogs comprising (i) a coronavirus having an S protein with at least 75% amino acid identity with CRCV S protein, or (ii) a coronavirus having an S protein with at least 75% amino acid identity with BCV S

protein, or (iii) a coronavirus protein having at least 75% amino acid identity with a CRCV protein or an immunogenic fragment thereof, or (iv) a coronavirus protein having at least 75% amino acid identity with a BCV protein or an immunogenic fragment thereof, or (v) a nucleic acid encoding said coronaviral protein or immunogenic fraction thereof.

Preferably, the vaccine is packaged and presented for use in dogs.

When the vaccine comprises a coronavirus protein, or an immunogenic fragment thereof, the protein preferably has at least 80%, or at least 85%, or at least 90%, or at least 95% amino acid identity with the corresponding portion of a BCV or CRCV protein.

Preferably, the coronavirus protein is a BCV, HCV, HEV or CRCV protein, or a modification thereof.

Typical protein modifications include amino acid substitutions to improve the antigenticity of the vaccine. BCV, HCV and HEV proteins may be modified to be more like a CRCV protein. For example, the spike protein of BCV, HCV or HEV may be modified to include a CRCV amino acid at any of differences shown in the comparison in Figure 10, or listed in Table 1.

Proteins in which one or more of the amino acid residues are chemically modified, may be used providing that the function of the protein, namely the production of specific antibodies *in vivo*, remains substantially unchanged. It is appreciated that synthesised proteins may be suitably modified before or after their synthesised. Such modifications include forming salts with acids or bases, especially physiologically acceptable organic or inorganic acids and bases, forming an ester or amide of a terminal carboxyl group, and attaching

amino acid protecting groups such as N-t-butoxycarbonyl. Such modifications may protect the peptide from *in vivo* metabolism.

The protein may be present as single copies or as multiples, for example tandem repeats. Such tandem or multiple repeats may be sufficiently antigenic themselves to obviate the use of a carrier. It may be advantageous for the protein to be formed as a loop, with the N-terminal and C-terminal ends joined together, or to add one or more Cys residues to an end to increase antigenicity and/or to allow disulphide bonds to be formed. If the protein is covalently linked to a carrier, preferably a polypeptide, then the arrangement is preferably such that the protein of the invention forms a loop.

According to current immunological theories, a carrier function should be present in any immunogenic formulation in order to stimulate, or enhance stimulation of, the immune system. It is thought that the best carriers embody (or, together with the antigen, create) a T-cell epitope. The peptides may be associated, for example by cross-linking, with a separate carrier, such as serum albumins, myoglobins, bacterial toxoids and keyhole limpet haemocyanin. More recently developed carriers which induce T-cell help in the immune response include the hepatitis-B core antigen (also called the nucleocapsid protein), presumed T-cell epitopes such as Thr-Ala-Ser-Gly-Val-Ala-Glu-Thr-Thr-Asn-Cys, beta-galactosidase and the 163-171 peptide of interleukin-1. The latter compound may variously be regarded as a carrier or as an adjuvant or as both. Alternatively, several copies of the same or different proteins of the invention may be cross-linked to one another; in this situation there is no separate carrier as such, but a carrier function may be provided by such cross-linking. Suitable cross-linking agents include those listed as such in the Sigma and Pierce catalogues, for example glutaraldehyde, carbodiimide and succinimidyl 4-(N-maleimidomethyl)cyclohexane-1-

carboxylate, the latter agent exploiting the -SH group on the C-terminal cysteine residue (if present).

If the protein is prepared by expression of a suitable nucleotide sequence in a suitable host, then it may be advantageous to express it as a fusion product with a peptide sequence which acts as a carrier. Kabigen's "Ecosec" system is an example of such an arrangement.

It is appreciated that the coronavirus component of the vaccine may be linked to other antigens to provide a dual effect.

Preferably, the coronavirus protein in the vaccine composition is an S protein. More preferably, the S protein is a CRCV or CRCV-like S protein as defined above in the first aspect of the invention or obtainable by the methods of the third aspect of the invention, a BCV S protein, an HCV S protein, an HEV S protein, or an immunogenic fragment thereof.

Most preferably, the vaccine composition contains a CRCV S protein that comprises or consists of the amino acid sequence listed in Figure 4, or an immunogenic fragment thereof having at least 97% identity with the sequence listed in Figure 4. Preferably, the variant has at least at least 98%, or at least 99% amino acid sequence identity with the sequence listed in Figure 4. More preferably the variant has at least 99.1%, or at least 99.2%, or at least 99.3%, or at least 99.4%, or at least 99.5%, or at least 99.6%, or at least 99.7%, or at least 99.8%, or at least 99.9% amino acid sequence identity with the sequence listed in Figure 4.

Additionally or alternatively, the vaccine composition may comprise coronavirus proteins such as a hemagglutinin-esterase protein (HE) or an integral membrane protein (M), or the small membrane protein (E) (Lai

MMC & Cavanagh D, (1997) "The molecular biology of coronaviruses" *Adv.Vir.Res*, 48: 1-100).

In one embodiment, the HE, E or M proteins are BCV, HCV of HEV proteins. In another embodiment, the HE, E or M proteins are CRCV proteins.

When the vaccine comprises a coronavirus, preferably the coronavirus comprises an S protein with at least 80%, or at least 85%, or at least 90%, or at least 95% amino acid identity with the BCV S protein. More preferably, the coronavirus comprises an S protein with at least 80%, or at least 85%, or at least 90%, or at least 95% amino acid identity with the CRCV S protein.

In another preferred embodiment, the vaccine composition comprises a virus selected from BCV, HCV, HEV and CRCV, or a modification thereof.

It is appreciated that dog vaccines effective against a canine virus may be derived from a non-canine virus. For example US Patent No. 5,750,112 to Gill, and assigned to Solvay Animal Health Inc, discloses a vaccine against enteric canine coronavirus containing inactivated feline enteric coronavirus. The disclosure of US 5,750,112 is incorporated herein by reference.

In one preferred embodiment, the virus is an inactivated virus. Methods for inactivating viruses for use in vaccines are well known in the art. Suitable methods include chemical methods, such as the use of beta proprio-lactone (BPL). Suitable inactivated bovine coronavirus vaccines may include inactivated BCV which is a component of bovine vaccines such as "Rotovec Corona" from Schering-Plough (http://www.ukvet.co.uk/rotovec/scour.htm); "Lactovac" by Hoechst Roussel Vet Ltd, (Veterinary Formulary 5th Edition of the Veterinary Data

Sheet Compendium); "First Defense" by Immuncell Corp, USA; "Scour Bos 4" by Grand Laboraotries and "Scour Guard 3K" by Pfizer.

In an alternative embodiment, the virus is an attenuated virus. Methods for attenuating viruses for use in vaccines are well known in the art.

Preferably, the vaccine composition also comprises a pharmaceutically acceptable adjuvant.

Preferably, when the vaccine comprises a nucleic acid, the nucleic acid encoding the coronaviral protein or immunogenic fraction thereof, for use as a vaccine is a CRCV or CRCV-like S polynucleotide. More preferably, the nucleic acid comprises or consists of the nucleotide sequence listed in Figure 3, or a fraction thereof.

For vaccine use the CRCV or CRCV-like S nucleic acid can be delivered in various replicating (e.g. recombinant adenovirus vaccine) or non-replicating (DNA vaccine) vectors.

In a preferred embodiment, the vaccine may contain recombinant CRCV or CRCV-like S protein, as well as other immunogenic coronavirus proteins.

In another preferred embodiment, the vaccine may contain recombinant CRCV or CRCV-like S protein, as well as other pathogenic organisms involved in respiratory disease of dogs such as canine parainfluenzavirus, canine adenovirus type 2, the bacterium *Bordetella bronchiseptica*, canine herpesvirus, human reovirus and mycoplasma species, or immunogenic proteins therefrom.

Vaccination would be useful especially but not exclusively for dogs prior to entry into a boarding kennel or for the vaccination of dogs in breeding facilities.

A typical dose of a vaccine comprised of recombinant protein is about 5-10 μg. A typical dose of a vaccine comprised of inactivated virus is about 1-10 mg.

In an eighth aspect, the invention provides the use of (i) a coronavirus having an S protein with at least 75% amino acid identity with CRCV S protein, or (ii) a coronavirus having an S protein with at least 75% amino acid identity with BCV S protein, or (iii) a coronavirus protein having at least 75% amino acid identity with a CRCV protein or an immunogenic fragment thereof, or (iv) a coronaviral protein having at least 75% amino acid identity with a BCV protein, or an immunogenic fragment thereof, or (v) a nucleic acid encoding said coronaviral protein or immunogenic fraction thereof, in the preparation of a medicament for stimulating an immune response against CRCV in a dog.

The invention includes the use of (i) a coronavirus having an S protein with at least 75% amino acid identity with CRCV S protein, or (ii) a coronavirus having an S protein with at least 75% amino acid identity with BCV S protein, or (iii) a coronavirus protein having at least 75% amino acid identity with a CRCV protein or an immunogenic fragment thereof, or (iv) a coronaviral protein having at least 75% amino acid identity with a BCV protein, or an immunogenic fragment thereof, or (v) a nucleic acid encoding said coronaviral protein or immunogenic fraction thereof, in the preparation of a medicament for prophylaxis of respiratory disease in a dog, typically CIRD.

When a coronavirus protein, or an immunogenic fragment thereof, is used in the preparation of the medicament, the protein preferably has at least 80%, or at least 85%, or at least 90%, or at least 95% amino acid identity with the corresponding portion of a BCV protein. Preferably the protein has

at least 80%, or at least 85%, or at least 90%, or at least 95% amino acid identity with the corresponding portion of a CRCV protein.

Preferably, the coronaviral protein used in the preparation of the medicament is a BCV, HCV, HEV or CRCV protein, or a modification thereof, as described above with reference to the seventh aspect of the invention.

More preferably, the coronaviral protein used in the preparation of the medicament is an S protein. Yet more preferably, the S protein comprises an CRCV or CRCV-like S protein as defined above in the first aspect of the invention or obtainable by the methods of the third aspect of the invention, a BCV S protein, an HCV S protein, or an immunogenic fragment thereof.

Most preferably, the coronaviral protein used in the preparation of the medicament comprises or consists of the amino acid sequence listed in Figure 4, or an immunogenic fragment thereof having at least 97% identity with the sequence listed in Figure 4. Preferably, the variant has at least least 98%, or at least 99% amino acid sequence identity with the sequence listed in Figure 4. More preferably the variant has at least 99.1%, or at least 99.2%, or at least 99.3%, or at least 99.4%, or at least 99.5%, or at least 99.6%, or at least 99.7%, or at least 99.8%, or at least 99.9% amino acid sequence identity with the sequence listed in Figure 4.

Additionally or alternatively, the coronaviral protein used in the preparation of the medicament may comprise HE, E or M coronavirus proteins. In one embodiment, the HE, E or M proteins are BCV, HCV or HEV proteins. In another embodiment, the HE, E or M proteins are CRCV proteins.

When a coronavirus is used in the preparation of the medicament, the coronavirus preferably comprises an S protein with at least 80%, or at least

85%, or at least 90%, or at least 95% amino acid identity with the BCV S protein. More preferably the coronavirus comprises an S protein with at least 80%, or at least 85%, or at least 90%, or at least 95% amino acid identity with the CRCV S protein.

In an ninth aspect, the invention provides a CRCV or CRCV-like S protein as defined above in the first aspect of the invention or obtainable by the methods of the third aspect of the invention, for use in medicine. Typically, the S protein will be used in veterinary medicine.

In a tenth aspect, the invention provides a method of vaccinating a dog against CRCV, the method comprising administering to the dog a vaccine composition as described above in the eighth aspect of the invention.

Typically, the vaccine will be administered *via* the intramuscular, subcutaneous or intranasal routes

In another embodiment, a dog can passively acquire immunity against CRCV by being administered an antibody that reacts with CRCV. The antibody that reacts with CRCV may be an anti-BCV, anti-HCV antibody, but is preferably an anti-CRCV antibody. Preferably, the antibody that reacts with CRCV is an anti-S protein antibody. Most preferably, the antibody that reacts with CRCV is an anti-CRCV S protein antibody as described in the fourth aspect of the invention.

In an eleventh aspect, the invention provides a method for combating the spread of CRCV between dogs comprising determining whether a dog is infected with CRCV according to the methods as described above in the fifth aspect of the invention, or using the immunosorbent assay or solid substrate as described above in the sixth aspect of the invention, and, if the dog is infected with CRCV, quarantining the dog.

By "quarantining" a dog we include the meaning of keeping the dog separate from all other dogs. We also include the meaning of keeping the dog separate from dogs that have not been vaccinated against CRCV, which can be performed as described above. We also include the meaning of keeping the dog separate from dogs that have not been infected by CRCV, which can be determined as described above.

In a twelfth aspect, the invention provides a method for combating the spread of CRCV between dogs comprising determining whether a dog is infected with CRCV according to the methods described above in the fifth aspect of the invention, or using the immunosorbent assay or solid substrate as described above in the sixth aspect of the invention, and, if the dog is infected with CRCV, vaccinating other dogs that have been, are, or are likely to be in contact with the dog.

A thirteenth aspect of the invention provides a method for identifying a test vaccine capable of preventing or reducing the incidence of canine infectious respiratory disease (CIRD) in dogs. The method comprises (a) determining whether a dog has been exposed to CRCV, typically according to the methods described above in the fifth aspect of the invention or using the immunosorbent assay or solid substrate as described above in the sixth aspect of the invention, (b) if the dog has not been exposed to CRCV, administering the test vaccine to the dog, (c) inoculating the dog with CRCV, and (d) determining whether the dog develops CIRD. The absence of CIRD in step (d) indicates that the test vaccine is capable of preventing CIRD.

Typically, this method is performed on a set of dogs.

Preferably, the method involves the use of a set of control dog which are not administered the test vaccine in step (b). The significantly lower incidence

of CIRD in the set of dogs that has been administered the test vaccine than in the control set indicates that the test vaccine is capable of preventing or reducing the incidence of CIRD.

The invention also includes a vaccine identified by this method.

All of the documents referred to herein are incorporated herein, in their entirety, by reference.

The invention will now be described in more detail with the aid of the following Figures and Examples.

Figure 1

Partial nucleotide sequence (250 residues) of the CRCV polymerase (pol) cDNA.

Figure 2

Partial amino acid sequence (83 residues) of the CRCV pol protein derived from the nucleotide sequence of Figure 1.

Figure 3

Nucleotide sequence (4092 residues) of the CRCV Spike (S) cDNA. The Y at position 3531 refers to either C or T.

Figure 4

Amino acid sequence (1363 residues) of the CRCV S protein derived from the nucleotide sequence of Figure 3.

Figure 5

Consensus tree for cDNA sequences from a 250 nucleotide region of the polymerase gene of 12 coronaviruses. The sequence obtained from the canine respiratory coronavirus is designated T101. The numbers indicate bootstrap values obtained by analysis of 100 data sets.

BCV: bovine coronavirus, CCV: canine coronavirus, FIPV: feline infectious peritonitis virus, HEV: hemagglutinating encephalomyelitis virus, IBV: infectious bronchitis virus, MHV: mouse hepatitis virus, OC43: human coronavirus strain OC43, SDAV: sialodacryoadenitis virus, TCV: turkey coronavirus, TGEV: transmissible gastroenteritis virus, 229E: human coronavirus strain 229E, T101: canine respiratory coronavirus (PCR product from tracheal sample T101)

Figure 6

CLUSTAL X (1.8) multiple sequence alignment of the 250 nucleotide partial sequence of the pol cDNA of CRCV (T101), BCV, HCV (OC43), HEV and CCV (enteric CCV).

Figure 7

CLUSTAL X (1.8) multiple sequence alignment of the 83 amino acid partial sequence of the pol protein of CRCV (protCRCVpol) with HCV (protHCVpoly), HEV (protHEVpoly), BCV (protBCVpoly) and CECV (enteric CCV) (protCECVpol).

Figure 8

CLUSTAL X (1.8) sequence alignment of the nucleotide sequence of the CRCV spike cDNA (CRCVspike) and enteric CCV spike cDNA (CECVspike).

Figure 9

CLUSTAL X (1.8) multiple sequence alignment of the 4092 nucleotides of the CRCV spike cDNA (CRCVspike) sequence with BCV (BCVspike), HCV (HCVspike) and HEV (HEVspike) spike cDNAs. The Y at position 3531 in the CRCV sequence refers to either C or T.

Figure 10

CLUSTAL X (1.8) multiple sequence alignment of the 1363 amino acid sequence of the CRCV spike protein (CRCVspikepr) with BCV (BCVspikepro), HCV (HCVspikepro), HEV (HEVspikepro) and enteric CCV (CECVspikepr) spike proteins.

Figure 11

RT-PCR using nested set of primers (Spike 1 and 2 followed by Spike 3 and 4). BCV: Bovine coronavirus positive control sample; A72: Coronavirus negative A72 cells; H_2O: PCR mix without DNA; T5 – T21: Tracheal samples of study dogs. The agarose gel electrophoresis shows PCR products of the expected size of 442bp for the positive control (BCV) and samples T12 and T21.

Figure 12

Comparison of the prevalence of respiratory disease in two groups of dogs. Dogs in group 1 were positive for serum antibodies to respiratory coronavirus on day of entry into the kennel, dogs in group 2 were negative. The graph shows the percentage of dogs developing respiratory disease in group 1 compared to group2 ($p<0.001$). n is the total number of dogs in each group.

Example 1: Detection of a novel coronavirus associated with canine infectious respiratory disease

Summary

An investigation into the causes of canine infectious respiratory disease (CIRD) was carried out in a large re-homing kennel. Tissue samples taken from the respiratory tract of diseased dogs were tested for the presence of coronaviruses using RT-PCR with conserved primers for the polymerase gene. Sequence analysis of four positive samples showed the presence of a novel coronavirus with high similarity to both bovine and human coronavirus (strain OC43) in their polymerase and spike genes whereas there was a low similarity to comparable genes in the enteric canine coronavirus. This canine respiratory coronavirus (CRCV) was detected by RT-PCR in 32/119 tracheal and 20/119 lung samples with the highest prevalence being detected in dogs with mild clinical symptoms. Serological analysis showed that the presence of antibodies against CRCV on the day of entry into the kennel decreased the risk of developing respiratory disease.

Materials and Methods

Study population

Dogs from a well-established re-homing kennel with a history of endemic respiratory disease were monitored for this study. On entry into the kennel, all dogs were vaccinated with KAVAK DA_2 PiP69 (Fort Dodge) a live attenuated vaccine for distemper virus, canine adenovirus type 2, canine parainfluenzavirus and canine parvovirus. Also, a killed leptospirosis vaccine was used (Fort Dodge). The health status of each dog was assessed twice a day by a veterinary clinician and the respiratory symptoms were

graded as follows: 1: no respiratory signs, 2: mild cough, 3: cough and nasal discharge, 4: cough, nasal discharge and inappetence, 5: bronchopneumonia. The overall health status of the dogs was graded as follows: 1: good health, 2: poor health, 3: very poor health. The age, breed and sex of the dogs were recorded.

For 119 dogs a full post mortem examination was performed. The tissue samples were stored at –70°C until further use.

Serum samples were collected from 112 dogs on day of entry into the re-homing kennel. For 81 dogs a follow-up serum was available on day 7 and for 112 dogs a serum was available on day 21 after entry.

Of the 112 dogs, 31 remained healthy during the 21 days between the first and the last serum sample whereas 81 dogs developed respiratory disease.

Sera from 35 dogs housed elsewhere were obtained from the diagnostic service of the Royal Veterinary College. These sera had been submitted for biochemical analysis for various reasons. Five of these sera were from 18-month-old beagles with no history of respiratory disease. Sera were routinely stored at –20°C.

RNA extraction and RT-PCR

RNA was extracted from tracheal and lung tissue of 119 dogs using TriReagent (Sigma). Approximately 25-50 mg of homogenised tissue was used and RNA was extracted as recommended by the manufacturer.

Synthesis of cDNA was performed using Random Hexamers (Roche) and ImPromII reverse transcriptase (Promega).

The polymerase gene of coronaviruses is known to be highly conserved, and has previously been used for phylogenetic analysis of this virus family (Stephensen *et al.,* 1999). For the detection of coronaviruses a modification of the primers 2Bp and 4Bm directed against the polymerase gene as described by Stephensen *et al.* (1999) were used (Conscoro5: 5' -ACT-CAR-ATG-AAT-TTG-AAA-TAT-GC; and Conscoro6: 5' -TCA-CAC-TTA-GGA-TAR-TCC-CA).

PCR was performed using Taq polymerase (Promega) in the provided reaction buffer containing a final concentration of 2.5 mM $MgCl_2$ and 0.5µM of primers. For PCR with the primers Conscoro5 and Conscoro6 the following temperature profile was used: After denaturation at 95°C for 5 min, 10 cycles were carried out at 95°C for 1 min, annealing at 37°C for 1 min and extension at 72°C for 1min. This was followed by 10 cycles using an annealing temperature of 45°C, 10 cycles at an annealing temperature of 50°C and 10 cycles at an annealing temperature of 53°C followed by a final extension at 72°C for 10 min.

A 20µl fraction of the PCR product was analysed on a 1.5% agarose gel and blotted onto a nylon membrane (Roche) after electrophoresis. The nylon membrane was hybridised with an oligonucleotide probe specific for the PCR product at 37°C overnight (Probe Conscoro: AAG-TTT-TAT-GGY-GGY-TGG-GA). The probe was 3'A-tailed with Digoxigenin-dUTP and was detected using anti-Digoxigenin conjugate and CSPD chemoluminescent substrate (Roche).

Primer sequences specific for the spike gene were derived from an alignment of the spike region of bovine coronavirus strain LY-1 (AF058942) and human coronavirus strain OC43 (L14643).

A PCR was performed with the primers Spike 1 and Spike 2, followed by a nested PCR using the primers Spike 3 and Spike 4 and 2µl of the product of the first amplification.

The numbers in brackets refer to the nucleotide position in the bovine coronavirus genome.

Spike 1: 5'-CTT-ATA-AGT-GCC-CCC-AAA-CTA-AAT (25291-25314)

Spike 2: 5' -CCT-ACT-GTG-AGA-TCA-CAT-GTT-TG (25912-25890)

Spike 3: 5' -GTT-GGC-ATA-GGT-GAG-CAC-CTG (25320-25339)

Spike 4: 5' -GCA-ATG-CTG-GTT-CGG-AAG-AG (25762-25742)

The temperature profile used was denaturation at 95°C for 5 min, followed by 35 cycles of denaturation at 95°C for 1 min, annealing at 55°C for 40 sec and elongation at 72°C for 1 min. The final extension was performed at 72°C for 10 min. The nested PCR produced a 442bp fragment.

PCR products were cloned into the pGEM-T-easy vector (Promega) and sequenced using the Thermo sequenase fluorescent labelled primer cycle sequencing kit with 7-deaza-dGTP (Amersham Pharmacia) using Cy5 labelled primers.

Phylogenetic analysis

An alignment of the 250 bp cDNA sequence from the polymerase gene to the corresponding sequences of 11 coronaviruses was performed using ClustalX (Thompson *et al.*, 1997).

The phylogenetic relationship to known coronaviruses was analysed using the Phylip 3.6 package (Felsenstein, 1989). The alignments were followed by a bootstrap analysis using the Seqboot programme. The obtained data

sets were used for a maximum parsimony analysis using the DNApars programme and a consensus tree was calculated using Consense. The resulting trees were drawn using the Treeview programme (Page, 1996).

ELISA

ELISA antigen for bovine coronavirus or enteric canine coronavirus (CECV) (the antigens are a preparation from virus infected cell cultures obtained from Churchill Applied Biosciences, Huntingdon, UK) was resuspended in PBS at the concentration recommended by the manufacturer and incubated on 96 well plates (Falcon) overnight at 37°C.

The plates were washed with PBS and blocked with PBS containing 5% skimmed milk powder for 30 min. The sera were diluted 1:100 in blocking buffer and incubated on the plates for 1h. After washing with PBS/ 0.05% Tween 20 (Sigma), a peroxidase labelled rabbit anti-dog IgG conjugate (Sigma) was added (1:5000 in PBS/0.05% Tween 20) for 1 h. The plates were incubated with colour substrate (OPD, Sigma) for 10 min and the reaction was stopped by adding 2M H_2SO4. The adsorption was determined in an ELISA photometer at 492nm.

Virus culture

Virus isolation is performed on canine adult lung fibroblasts (passage 3 to 7), MDCK and A72 cells. The lung fibroblasts are maintained in MEM with 20% fetal calf serum (FCS), MDCK and A72 cells are maintained in MEM with 5% FCS. Tracheal tissue samples (approx. 25mg) are homogenised using a scalpel and mixed vigorously in 1ml MEM containing Penicillin (100U/ml), Streptomycin (0.1mg/ml), Amphotericin B (2.5µg/ml) and Trypsin (1µg/ml). The samples are centrifuged at 13000 rpm for 10 min. and the supernatant is used to inoculate cell cultures. After 30 min. at

37°C the supernatant is removed and maintenance medium added to the cultures. The cultures are passaged three times in the absence of a cytopathic effect. Then, RNA is extracted from the cells and RT-PCR to detect the presence of CRCV is performed.

Statistical analysis

The data were analysed using the chi-square test or Fisher's exact test and p values below 0.05 were considered statistically significant.

Results

PCR using consensus primers for the coronavirus RNA polymerase gene

Using the primers Conscoro5 and Conscoro6, cDNA obtained from 40 tracheal samples was analysed by RT-PCR.

Out of these, seven were found to be positive by PCR and subsequent hybridisation (17.5%).

The PCR products were cloned and sequenced (Figures 1 and 2) and the sequence data were compared to available viral sequences using the FASTA search program (Pearson, 1990).

Comparison of the coronavirus cDNA polymerase sequence obtained from four of the canine tracheal samples to other coronavirus sequences revealed that they were most similar to sequence data from BCV strain Quebec and LY138 (Genbank Accession Nos. AF220295 and AF058942, respectively) and human coronavirus strain OC43 (Genbank Accession No. AF124989). The similarity in the analysed 250 bp sequence was 98.8% for BCV

Quebec, and 98.4% for BCV LY138 and the HCV pol genes, whereas it was only 68.53% for CCV strain 1-71 pol gene (Figures 6 and 7).

An alignment of the novel sequence with the corresponding sequences of 11 coronaviruses and phylogenetic analysis using the maximum parsimony method resulted in the consensus tree shown in Figure 5. The cDNA sequence obtained from a tracheal sample (T101) was found on a common branch with bovine coronavirus, human coronavirus-OC43 and hemagglutinating encephalomyelitis virus.

The virus was called canine respiratory coronavirus (CRCV).

PCR using primers for the spike gene

For further analysis of the RNA sequence of CRCV, an alignment of the RNA for the spike gene of the bovine coronavirus LY 138 strain (AF058942) and the human coronavirus OC43 strain (L14643) was performed using Clustal X (Thompson *et al.,* 1997). Consensus regions were chosen for the selection of the nested primer sets Spike 1-2 and Spike 3-4 (Figure 11). PCR analysis was performed with the cDNA obtained from 119 tracheal and lung samples using these nested primers.

In total 32 tracheal samples (26.9%) and 20 lung samples (16.8%) were found positive by nested PCR. For eight dogs a positive PCR result was obtained for both, trachea and lung.

Sequence analysis of the PCR products obtained from tissues of six different dogs showed identical DNA sequences for these cDNAs (Figures 3 and 4). A comparison to known coronavirus spike sequences using the FASTA program revealed a 98.1% similarity to bovine coronavirus and a 97.8% similarity to human coronavirus OC43 (Figures 9 and 10).

Association of PCR positive samples with respiratory signs

Using primers for the spike gene, tracheal and lung samples from 119 dogs were analysed by RT-PCR for CRCV. Of these 42 were from dogs with no respiratory signs (grade 1), 18 dogs had shown mild respiratory signs (grade 2), 46 had shown moderate (grade 3) and 13 severe respiratory signs (grades 4 and 5). Grades 4 and 5 were merged due to the low case numbers in these groups.

Table 2 shows the PCR results for coronavirus in dogs with different grades of respiratory disease. Specifically, table 2 shows the RT-PCR results from tracheal and lung samples of 119 dogs with different respiratory signs (none to severe) using a nested PCR directed against the coronavirus spike gene as well as the number of positive samples out of total sample number and the percentage of positive samples (in brackets).

Table 2: RT-PCR results for tracheal and lung samples

Respiratory signs	Trachea: Positive samples	Lung Positive samples	Trachea and lung Positive samples
None	11/42 (26.2%)	8/42 (19.1%)	2/42
Mild	10/18 (55.6%)	4/18 (22.2%)	4/18
Moderate	9/46 (19.6%)	8/46 (17.4%)	2/46
Severe	2/13 (15.4%)	0/13	0/13

Establishment of a serological assay for CRCV

Because of the homology of the spike cDNA of CRCV to the spike region of bovine coronavirus, an ELISA antigen for BCV was used for serological analysis of CRCV.

Sera from five dogs with no history of infectious respiratory disease that had not been housed in the investigated kennel were tested. The OD values ranged from –0.013 to 0.39 with an average OD value of 0.154. Furthermore, sera from 30 dogs admitted to a veterinary clinic for various reasons were tested for antibodies to coronavirus. Of these, 20 samples showed an OD of <0.4 (-0.46 to 0.396) and 10 samples showed an OD of >1.0 (1.012 to 1.949). Samples with an OD of 0.6 or above were subsequently considered positive.

Comparison of the immune response to CRCV of dogs with and without respiratory disease

The BCV-antigen ELISA was performed using paired sera of 111 dogs from the study kennel. Of these, 81 dogs had shown symptoms of respiratory disease during a period of 21 days and 30 had remained healthy.

Of the group of dogs with respiratory disease, 17 were positive for antibodies to CRCV on the day of entry into the kennel and 64 were negative.

Of the 64 dogs with no detectable antibodies to BCV on day one, 63 tested positive on day 21. All 46 dogs out of these 63 for which a sample on day 7 was available tested negative on day 7. Therefore 63 dogs showed a

seroconversion during the study-period whereas only one dog remained negative.

Of the 31 dogs that had remained healthy, 17 had antibodies to CRCV on the day of entry. All of the 13 dogs that were negative on day 1 tested negative on day 7 but showed a seroconversion by day 21.

Thus, of 34 dogs that were positive for antibodies to CRCV on arrival in the kennel, 17 developed respiratory disease (50%) whereas of 77 dogs that were negative on arrival, 64 developed respiratory signs during the study-period (83.1%), (Figure 12).

Therefore dogs that had no antibodies to CRCV on entry into the kennel had an increased probability of developing respiratory disease ($p<0.001$).

Only one out of the 77 dogs that were negative on arrival remained negative during the study period of 21 days whereas 76 dogs showed a seroconversion.

Serology using canine enteric coronavirus (CECV) antigen

An ELISA assay using a canine coronavirus antigen was performed to investigate whether CRCV showed a serological cross reaction to canine enteric coronavirus. Sera from 27 dogs, previously tested for antibodies to CRCV using the BCV antigen were selected.

It was found that eight dogs had antibodies to CECV on the day of entry into the kennel, of these four also had antibodies to CRCV. Nineteen dogs were found to be negative for CECV on day 1, 17 of these were also negative for CRCV. Of the 19 negative dogs, five showed a seroconversion to CECV during the 21-day period of the investigation and 17 showed a seroconversion to CRCV.

Analysis of the prevalence of respiratory disease in this group showed that six out of the eight dogs (75%) that were positive for antibodies to CECV on day 1 developed respiratory disease. Out of the group of 19 dogs that had no detectable antibodies to CECV on day 1, 15 showed signs of respiratory disease (78.9%), (p=0.594).

Virus isolation

Tracheal tissue samples from dogs that are identified as positive for CRCV RNA by RT-PCR are inoculated on cell cultures of canine adult lung fibroblasts and MDCK cells. For some samples, virus isolation is also performed on A72 cells. The cultures show no signs of a cytopathic effect during three passages. After several passage, RNA is extracted from the cultures and found to be positive for CRCV by RT-PCR.

Discussion

This study reports the detection of a novel coronavirus, CRCV, in kennelled dogs with respiratory disease.

Coronaviruses have been reported to cause respiratory disease of man, cattle, swine and poultry, but their presence in the respiratory tract of dogs and a possible association with canine infectious respiratory disease (CIRD) has not been determined.

Dogs were investigated from a kennel in which CIRD was endemic and could not be controlled by the use of vaccines recommended against CIRD. Samples taken from the respiratory tract of these dogs were examined using RT-PCR primers directed to the conserved polymerase gene of coronaviruses (Stephensen *et al.*, 1999).

Initially, seven tracheal samples were found to be positive; the sequence of the RT-PCR products was determined and compared to all available coronavirus polymerase gene sequences. This analysis revealed that the cDNA sequence obtained from the canine samples had the highest similarity to the polymerase gene of bovine coronavirus (98.8% and human coronavirus OC43 (98.4%) but only a very low similarity to the polymerase gene of the enteric canine coronavirus (strain 1-71, 68.53% similarity).

A phylogenetic analysis was performed using the polymerase sequences of eleven additional coronaviruses. The coronavirus detected in the respiratory tract of dogs (CRCV) was located on a common branch with three group 2 viruses: BCV, HCV strain OC43 and HEV. However, canine enteric coronavirus, a group 1 coronavirus, was shown to be only distantly related.

Canine respiratory coronavirus therefore is a novel coronavirus of dogs that is most closely related to BCV and HCV-OC43, both of which are known to cause respiratory disease.

To obtain more sequence information and to further determine the relationship to other coronaviruses using a more variable gene, a part of the spike gene was analysed. Since CRCV had been shown to be most similar to BCV and HCV-OC43, an alignment of the sequences of their spike genes was used to design a nested set of primers. Nested primers were chosen to achieve a more sensitive assay.

Sequencing of the products of this RT-PCR confirmed the high similarity of CRCV with BCV and HCV-OC43.

The presence of antibodies to CRCV was analysed using an ELISA based on a BCV antigen because of the high sequence similarity of the two viruses

in the spike cDNA. The ELISA results confirmed the presence of a virus similar to BCV in the study population.

The prevalence of antibodies was 30% at the time of entry into the kennel and 99% after 21 days.

Interestingly and unexpectedly, serological analysis revealed that dogs with antibodies to CRCV on day of entry into the kennel developed respiratory disease less frequently than dogs without antibodies ($p<0.001$). Therefore the presence of antibodies to CRCV had a protective effect against respiratory disease in this population.

Almost all dogs negative on day of entry into the kennel showed a seroconversion to CRCV within three weeks, indicating that the virus is highly contagious. Serology using an antigen for canine enteric coronavirus (CECV) showed a much lower prevalence of antibodies to CECV on day 21. Therefore the BCV-ELISA results did not reflect an infection with canine enteric coronavirus and the cross-reactivity between the two antigens seems to be low.

Serum antibodies to CRCV were present in about 30% of dogs of various origins including dogs entering a re-homing kennel as well as pet dogs. The presence of CRCV is therefore not limited to the investigated kennel and the virus seems to be established in the dog population.

By PCR, CRCV was detected in tracheal tissue and lung tissue and therefore appears to infect the upper and lower respiratory tract of dogs. Within the kennelled population, CRCV-RNA was detected in 27.3% of dogs with all grades of respiratory disease as well as in 26.2% of dogs that were apparently healthy at the time of euthanasia.

CRCV-RNA was most frequently found in the trachea of dogs with mild cough (55%). Studies using the human coronavirus strain 229E have shown, that coronaviruses can cause disruption of the respiratory epithelium and ciliary dyskinesia (Chilvers *et al.,2001*). Without being bound by theory, we believe that an infection with CRCV has a similar effect, and that the virus plays an important role in the early stages of the pathogenesis of CIRD. By damaging the respiratory epithelium and disrupting ciliary clearance CRCV facilitates the entry of other viral or bacterial pathogens. Therefore while CRCV infection on its own may cause only mild respiratory symptoms, in conjunction with other pathogenic agents it could lead to severe respiratory disease.

The pathogenesis of CIRD has not been thoroughly investigated since the 1970s when *Bordetella bronchiseptica*, canine adenovirus type 2 and canine parainfluenza were determined to be the main causes of the disease. However the vaccination of all dogs against CPIV, CAV-2 and distemper virus did not help to control the disease in this kennel despite evidence that the majority of dogs responded to the vaccine within 21 days (data not shown).

This study shows an association of a novel canine respiratory coronavirus with CIRD. The aetiology of CIRD therefore needs to be re-evaluated and the role of novel microorganisms or microorganisms previously not associated with the disease has to be established.

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CLAIMS

1. A coronavirus Spike (S) protein, or fragment thereof, having at least 75% amino acid sequence identity with the CRCV S protein whose amino acid sequence is listed in Figure 4, and comprising at least one of the canine respiratory coronavirus (CRCV)-specific amino acids listed in Table 1.

2. A coronavirus S protein that comprises the amino acid sequence listed in Figure 4, or a variant thereof with at least 97% amino acid sequence identity with the sequence listed in Figure 4.

3. A coronavirus polymerase (pol) protein, or fragment thereof, having at least 90% amino acid sequence identity with BCV pol protein and comprising the amino acid E at the position corresponding to position 4975 in the BCV genome (Accession No. SWALL: Q91A29).

4. A coronavirus pol protein that comprises the amino acid sequence listed in Figure 2.

5. A polynucleotide that encodes the protein according to any one of Claims 1 to 4, or the complement thereof.

6. A polynucleotide according to Claim 5 comprising the nucleotide sequence listed in Figure 3.

7. A polynucleotide according to Claim 5 comprising the nucleotide sequence listed in Figure 1.

8. A vector comprising the polynucleotide of any one of Claims 5 to 7.

9. A host cell comprising the vector of Claim 8.

10. A method of obtaining a protein encoded by the vector of Claim 8, the method comprising culturing the host cell of Claim 9, expressing the protein in the host cell, and purifying the protein.

11. A protein obtainable by the method of Claim 10.

12. A host cell according to Claim 9 wherein the vector is an expression vector comprising a eukaryotic promoter operatively linked to the polynucleotide, and wherein the host cell is a eukaryotic host cell.

13. A method of obtaining a glycosylated S protein encoded by the polynucleotide of Claim 5 or 6, the method comprising culturing the host cell of Claim 12, expressing the protein in the host cell, and purifying the protein.

14. A glycosylated S protein obtainable by the method of Claim 13.

15. A method of making an anti-CRCV antibody comprising (i) raising an immune response to an S protein according to any one of Claims 1, 2, 11 or 14 in an animal and preparing an antibody from the animal or from an immortal cell derived therefrom, or (ii) selecting an antibody from an antibody-display library using an S protein according to any one of Claims 1, 2, 11 or 14.

16 A method according to Claim 15 further comprising determining whether the antibody has greater affinity for the CRCV S protein than for the BCV S protein.

17. An anti-CRCV antibody obtainable by the method of Claim 15 or 16, that has greater affinity for the CRCV S protein than for the BCV S protein.

18. A method of determining whether a dog has been exposed to CRCV, the method comprising:

(a) obtaining a suitable sample from the dog; and

(b) identifying CRCV or an anti-CRCV antibody in the sample.

19. A method according to Claim 18 wherein the anti-CRCV antibody is detected using a CRCV, BCV, human coronavirus (HCV) or hemagglutinating encephalomyelitis virus (HEV) antigen.

20. A method according to Claim 19 wherein the suitable sample is an antibody containing sample such as serum, saliva, tracheal wash and bronchiolar lavage, and wherein identifying an anti-CRCV antibody comprises identifying an antibody that selectively binds to BCV S protein (AF058942) HCV S protein (L14643), or to a coronavirus having an S protein with at least 75% amino acid identity with CRCV S protein (Figure 4) or a fragment thereof.

21. A method according to Claim 18 wherein the suitable sample is an antibody containing sample, and wherein identifying an anti-CRCV antibody comprises identifying an antibody that selectively binds to an S protein according to any one of Claims 1, 2, 11 or 14,

22. A method according to Claim 18 wherein the suitable sample is a lung wash, tracheal wash, tonsilar swab or a biopsy or post-mortem sample from the respiratory tract of the dog.

23. A method according to Claim 22 wherein identifying CRCV comprises identifying a nucleic acid component of CRCV.

24. A method according to Claim 23 wherein identifying a nucleic acid component of CRCV comprises identifying a polynucleotide that hybridises at high stringency to the BCV genome (AF058942).

25. A method according to Claim 23 or 24 wherein identifying CRCV comprises identifying a polynucleotide as defined in any one of Claims 5 to 7.

26. A method according to Claim 22 wherein identifying CRCV comprises identifying a protein component of CRCV.

27. A method according to Claim 26 wherein identifying a protein component of CRCV comprises identifying a protein according to any one of Claims 1, 2, 11 or 14.

28. A method according to Claim 26 wherein identifying a protein component of CRCV comprises using an antibody reactive with CRCV.

29. A method according to Claim 28 wherein the antibody reactive with CRCV is an anti-BCV antibody, an anti-HCV antibody, or an anti-CRCV antibody according to Claim 17.

30. An immunosorbent assay for detecting anti-CRCV S antibodies, the assay comprising:

a solid phase coated with an S protein according to any one of Claims 1, 2, 11 or 14, or an antigenic fragment thereof, wherein anti-CRCV

S antibodies in a sample exposed to the solid phase will bind to the polypeptide; and

a detectable label conjugate which will bind to the anti-CRCV antibodies bound to the solid phase.

31. An immunosorbent assay according to Claim 30, wherein the solid phase is a microtitre well.

32. An immunosorbent assay according to Claim 30, wherein the conjugate comprises anti-dog antibody.

33. An immunosorbent assay according to any one of Claims 30 to 32, wherein the conjugate comprises an enzyme.

34. An immunosorbent assay according to Claim 33, wherein the enzyme is horseradish peroxidase.

35. An immunosorbent assay according to Claim 33 or 34, further comprising a substrate for the enzyme.

36. A solid substrate with an S protein according to any one of Claims 1, 2, 11 or 14, or an antigenic fragment thereof, attached thereto, for capturing anti-CRCV S antibodies from a liquid sample, wherein anti-CRCV S antibodies in a sample exposed to the solid substrate will bind to the S protein.

37. A solid substrate according to Claim 36, wherein the solid substrate is a microtitre well.

38. A vaccine composition for vaccinating dogs comprising a coronavirus having an S protein with at 75% least amino acid identity with BCV S protein, or a coronaviral protein having at least 75% amino acid identity with a BCV protein or an immunogenic fragment thereof, or a nucleic acid encoding said coronaviral protein or immunogenic fraction thereof.

39. A vaccine composition according to Claim 38 wherein the coronaviral protein is a BCV, HCV, HEV or CRCV protein, or a modification thereof.

40. A vaccine composition according to Claim 38 or 39 wherein the coronaviral protein is an S protein.

41. A vaccine composition according to Claim 40 comprising an S protein according to any one of Claims 1, 2, 11 or 14, a BCV S protein, an HCV S protein, an HEV S protein, or an immunogenic fragment thereof.

42. A vaccine composition according to Claim 38 or 39 wherein the coronaviral protein is a hemagglutinin-esterase protein (HE) or an integral membrane protein (M).

43. A vaccine composition according to Claim 38 wherein the virus is selected from BCV, HCV, HEV and CRCV, or a modification thereof.

44. A vaccine composition according to any one of Claims 38 to 43 and also comprising a pharmaceutically acceptable adjuvant.

45. Use of (i) a coronavirus having an S protein with at least 75% amino acid identity with CRCV S protein, or (ii) a coronavirus having an S protein

with at least 75% amino acid identity with BCV S protein, or (iii) a coronavirus protein having at least 75% amino acid identity with a CRCV protein or an immunogenic fragment thereof, or (iv) a coronaviral protein having at least 75% amino acid identity with a BCV protein, or an immunogenic fragment thereof, or (v) a nucleic acid encoding said coronaviral protein or immunogenic fraction thereof, in the preparation of a medicament for stimulating an immune response against CRCV in a dog.

46. Use of (i) a coronavirus having an S protein with at least 75% amino acid identity with CRCV S protein, or (ii) a coronavirus having an S protein with at least 75% amino acid identity with BCV S protein, or (iii) a coronavirus protein having at least 75% amino acid identity with a CRCV protein or an immunogenic fragment thereof, or (iv) a coronaviral protein having at least 75% amino acid identity with a BCV protein, or an immunogenic fragment thereof, or (v) a nucleic acid encoding said coronaviral protein or immunogenic fraction thereof, in the preparation of a medicament for prophylaxis of respiratory disease in a dog.

47. Use according to Claim 45 or 46 wherein the coronaviral protein is a BCV, HCV, HEV or CRCV protein, or a modification thereof.

48. Use according to any one of Claims 45 to 47 wherein the coronaviral protein is an S protein.

49. Use according to Claim 48 wherein the S protein comprises an S protein according to any one of Claims 1, 2, 11 or 14, a BCV S protein, an HCV S protein, or an immunogenic fragment thereof.

50 Use according to any one of Claims 45 to 47 wherein the coronaviral protein is HE or M.

51. Use according to Claim 45 wherein the virus is selected from BCV, HCV, HEV and CRCV, or a modification thereof.

52. An S protein according to any one of Claims 1, 2, 11 or 14 for use in medicine.

53. A method of vaccinating a dog against CRCV, the method comprising administering to the dog a vaccine composition according to any one of Claims 38 to 44.

54. A method for combating the spread of CRCV between dogs comprising determining whether a dog is infected with CRCV according to the method of any one of Claims 18 to 29 and, if the dog is infected with CRCV, quarantining the dog.

55. A method for combating the spread of CRCV between dogs comprising determining whether a dog is infected with CRCV according to the method of any one of Claims 18 to 29 and, if the dog is infected with CRCV, vaccinating other dogs that have been, are, or are likely to be in contact with the dog.

56. A method for identifying a test vaccine capable of preventing canine infectious respiratory disease (CIRD) in dogs, comprising

(a) determining whether a dog has been exposed to CRCV according to the method of any one of Claims 18 to 29,

(b) if the dog has not been exposed to CRCV, administering the test vaccine to the dog,

(c) inoculating the dog with CRCV, and

(d) determining whether the dog develops CIRD,

wherein the absence of CIRD in step (d) indicates that the test vaccine is capable of preventing CIRD.

57. A vaccine identified by the method of Claim 55.

58. The *E. coli* strain Spike D-1 CRCV, containing a plasmid whose insert contains a portion of the CRCV S cDNA, as deposited by the Royal Veterinary College at the NCIMB under Accession number NCIMB 41146 on 25 July 2002.

59. The plasmid contained in *E. coli* strain Spike D-1 CRCV, deposited by the Royal Veterinary College at the NCIMB under Accession number NCIMB 41146 on 25 July 2002.

60. A kit of parts for the immunosorbent assay according to any one of Claims 30-35, comprising a solid phase, a CRCV or CRCV-like S protein for coating the solid phase, and a detectable label conjugate.

61. A kit of parts according to Claim 60 wherein the solid phase comprises a microtitre plate.

62. A kit of parts according to Claim 60 or 61 wherein the detectable label conjugate comprises an anti-dog antibody.

63. A kit of parts according to Claim 60 or 61 wherein the detectable label conjugate comprises an enzyme.

64. A kit of parts according to Claim 63 further comprising a substrate for the enzyme.

65. A kit of parts according to any one of Claims 60 to 64 further comprising a positive control sample that contains an anti-CRCV S protein antibody.

66. A method of passively immunising a dog against CRCV, comprising administering an antibody that reacts with CRCV to the dog.

67. A method according to Claim 66 wherein the antibody that reacts with CRCV comprises an anti-BCV antibody, an anti-HCV antibody, or an anti-CRCV antibody.

68. A method according to Claim 66 or 67 wherein the antibody that reacts with CRCV comprises an anti-S protein antibody.

69. A method according to any one of Claims 66 to 68 wherein the antibody that reacts with CRCV comprises the anti-CRCV antibody according to Claim 17.

70. Use of an antibody that reacts with CRCV in the preparation of a medicament for passively immunising a dog against CRCV.

71. Use according to Claim 70 wherein the antibody that reacts with CRCV is an anti-BCV antibody, an anti-HCV antibody, or an anti-CRCV antibody.

72. Use according to Claim 70 or 71 wherein the antibody that reacts with CRCV is an anti-S protein antibody.

73. Use according to any one of Claims 70 to 72 wherein the antibody that reacts with CRCV comprises the anti-CRCV antibody according to Claim 17.

ABSTRACT

VIRUS

A canine respiratory coronavirus (CRCV) that is present in the respiratory tract of dogs with canine infectious respiratory disease and which has a low level of homology to the enteric canine coronavirus, but which has a high level of homology to the bovine coronavirus strains Quebec and LY138 and human coronavirus strain OC43.

The CRCV spike and polymerase cDNA and protein partial sequences are listed in Figures 1 to 4.

Figure no. 4

FIGURE 1

```
ctcagatgaa tttgaaatat gctattagtg ctaagaatag agcccgcact gttgctggtg      60
tttccatact tagtactatg actggcagaa tgtttcatca aaaatgtttg aaaagtatag     120
cagctacacg tggtgttcct gttgttatag gcaccactaa attttatggc ggctgggatg     180
atatgttacg tcgccttatt aaagatgttg acaatcctgt acttatgggt tgggattatc     240
ctaagtgtga                                                            250
```

FIGURE 2

```
QMNLKYAISA KNRARTVAGV SILSTMTGRM FHQKCLKSIA ATRGVPVVIG TTKFYGGWDD      60
MLRRLIKDVE NPVLMGWDYP KCE                                              84
```

FIGURE 3 (Page 1 of 2)

```
atgtttttga tacttttaat ttccttacca atggctttg ctgttatagg agatttaaag      60
tgtactacgg tttccatcaa tgatgttgac accggtgctc cttctattag cactgatgtt     120
gtcgatgtta ctaatggttt aggtacttat tatgttttag atcgtgtgta tttaaatact     180
acattgttgc ttaatggtta ttatcctact tcaggttcta catatcgtaa tatggcactg     240
aagggaactt tactattgag cacactatgg tttaaaccac catttctttc tgattttatt     300
gatggtgttt ttgctaaggt aaaaaatacc aaggttatta aagatggtgt agtgtatagt     360
gagtttcctg ctataactat aggtagtact tttgtaaata catcctatag tgtggtagta     420
caaccacata ctactaattt agataataaa ttacaaggtc tcttagagat ctctgtttgc     480
cagtatacta tgtgcgatta cccacatacg atgtgtcatc ctaatctggg taataaacgc     540
atagaactat ggcattggga tacaggtgtt gttccctgtt tatataagcg taatttcaca     600
tatgatgtga atgctgatta tttgtattcc catttttatc aagaaggtgg tacttttat      660
gcatatttta cagacactgg tgttgttact aagtttctgt ttcatgttta tttaggcacg     720
gtgctttcac attattatgt catgcccttg acttgtaata gtgctatgac tttagaatac     780
tgggttacac ctctcacttt taaacaatat ttactcgctt tcaatcaaga tggtgttatt     840
tttaatgctg ttgattgtaa gagtgatttt atgagtgaga ttaagtgtaa aacactatct     900
atagcaccat ctactggtgt ttatgaatta aacggttaca ctgttcagcc aattgcagat     960
gtttaccgac gtatacctaa tcttcccgat tgtaatatag aggcttggct taatgataag    1020
tcggtgcctt ctccattaaa ttgggaacgt aagacctttt caaattgtaa ttttaatatg    1080
agcagcctga tgtcttttat ccaggctgac tcgtttactt gtaataatat tgatgctgct    1140
aagatatacg gtatgtgttt tttcagcata actatagata agtttgctat acccaatggt    1200
aggaaggttg acctacaaat gggcaatttg ggctatttgc agtcttttaa ctatagaatt    1260
gatactactg ctacaagttg tcagttgtat tataatttac ctgctagtaa tgtttctatt    1320
agcaggttta atccttctat ttggaatagg agatttggtt ttacagaaca atctgttttt    1380
aagcctcaac ctgtaggtgt ttttactgat catgatgttg tttatgcaca acattgtttt    1440
aaagctccca caaatttctg tccgtgtaaa ttgaatgggt ctttgtgtgt aggtagtggt    1500
tttggtatag atgctggtta taaaaatagt ggtataggca cttgtcctgc aggtactaat    1560
tatttaactt gttataatgc taaccaatgt gattgtttgt gcactccaga ccctatttta    1620
tctaaatcta cagggcctta taagtgcccc caaactaaat acttagttgg cataggtgag    1680
cactgttctg gtcttgctat taaaagtgat tattgtggag gcaatccttg tacttgccaa    1740
ccaaaagcat ttttgggttg gtctgtggac tcttgtttac aaggggatag gtgtaatatt    1800
tttgctaatt ttattttgca tggtgttaat agtggtacta cttgttctac tgatttacaa    1860
aaatcaaaca cagacataat tcttggtgtt tgtgttaatt atgatcttta tggtattaca    1920
ggccaaggta tttttgttga ggttaatgcg acttattata atagttggca gaacctttta    1980
tatgattcta atggtaatct ctatggtttt agggactact taacaaacag aacttttatg    2040
attcgtagtt gctatagcgg tcgtgtttca gcgggctttc actctaactc ttccgaacca    2100
gcattgctat ttcggaatat taaatgcaat tacgttttta ataatactct ttcacgacag    2160
ctgcaaccta ttaactattt tgatagttat cttggttgtg ttgtcaatgc tgataatagt    2220
```

FIGURE 3 (Page 2 of 2)

```
acttctagtt ctgttcaaac atgtgatctc acagtaggta gtggttactg gggggattac   2280
tctacacaaa gacgaagtcg tagaacgatt accactggtt atcggtttac taattttgag   2340
ccatttactg ttaatccagt aaatgatagt ttacaccctg taggtggttt gtatgaaatt   2400
caaatacctt cagagtttac tataggtaat atggaggagt ttattcaaac aagatctcct   2460
aaagttacta ttgattgtcc tgtttttgtc tgtggtgatt atgcagcatg taaatcacag   2520
ttggttgaat atggtagttt ttgtgacaat attaatgcta tactcacaga agtaaatgaa   2580
ctacttgaca ctacacagtt gcaagtagct aatagtttaa tgaatggtgt cactcttagc   2640
actaagctta aagatggctt taatttcaat gtagatgaca tcaatttttc ccctgtatta   2700
ggttgtttag gaagcgaatg taataaagtt tccagtagat ctgctataga ggatttactt   2760
ttttctaaag taaagttatc tgatgttggt tttgttgatg cttataataa ttgtactgga   2820
ggtgccgaaa ttagggacct catttgtgtg caaagttata atggtatcaa agtgttgcct   2880
ccactgctct cagaaaatca gatcagtgga tacactttgg ctgccacctt tgctagtctg   2940
tttcctcctt ggtcagcagc agcaggcgta ccattttatt taaatgttca gtatcgtatt   3000
aatggtattg gtgttaccat ggatgtgcta actcaaaatc aaaagcttat ttctaatgca   3060
tttaacaatg cccttgatgc tattcaggaa gggtttgatg ctaccaattc tgctttagtt   3120
aaaattcaag ctgttgttaa tgcaaatgct gaagctctta ataacttatt gcaacaactc   3180
tctaataaat ttggtgctat aagtgcttct ttacaagaaa ttctatctag acttgatgct   3240
cttgaagcgc aagctcagat agacagactt atcaatgggc gtcttaccgc tcttaatgct   3300
tatgtttctc aacagcttag tgattctaca ctagtaaaat ttagtgcagc acaagctatg   3360
gagaaggtta atgaatgtgt caaaagccaa tcatctagga taaatttttg tggtaatggt   3420
aatcatatta tatcattagt gcagaatgct ccatatggtt tgtattttat ccactttagc   3480
tatgtcccta ctaagtatgt cactgcgaag gttagtcccg gtctgtgcat ygcaggtgat   3540
agaggtatag ctcctaagag tggttatttt gttaatgtaa ataacacttg gatgttcact   3600
ggtagtggtt attactaccc tgaacctata actggaaata atgtggttgt tatgagtacc   3660
tgtgctgtta actatactaa agcaccggat gtaatgctga acatttcaac acccaacctc   3720
cctgatttta aggaagagtt ggatcaatgg tttaaaaacc aaacattaat ggcaccagat   3780
ttgtcacttg attatataaa tgttacattc ttggacctac aagatgaaat gaataggtta   3840
caggaggcaa taaaagtttt aaatcatagc tacatcaatc tcaaggacat tggtacatat   3900
gaatattatg taaaatggcc ttggtatgta tggcttttaa ttggccttgc tggcgtagct   3960
atgcttgttt tactattctt catatgctgt tgtacaggat gtgggactag ttgttttaag   4020
aaatgcggtg gttgttgtga tgattatact ggacatcagg agttagtaat caaaacgtca   4080
catgacgact aa                                                      4092
```

FIGURE 4

```
MFLILLISLP MAFAVIGDLK CTTVSINDVD TGAPSISTDV VDVTNGLGTY YVLDRVYLNT     60
TLLLNGYYPT SGSTYRNMAL KGTLLLSTLW FKPPFLSDFI DGVFAKVKNT KVIKDGVVYS    120
EFPAITIGST FVNTSYSVVV QPHTTNLDNK LQGLLEISVC QYTMCDYPHT MCHPNLGNKR    180
IEWHWDTGV VPCLYKRNFT YDVNADYLYS HFYQEGGTFY AYFTDTGVVT KFLFHVYLGT    240
VLSHYYVMPL TCNSAMTLEY WVTPLTFKQY LLAFNQDGVI FNAVDCKSDF MSEIKCKTLS    300
IAPSTGVYEL NGYTVQPIAD VYRRIPNLPD CNIEAWLNDK SVPSPLNWER KTFSNCNFNM    360
SSLMSFIQAD SFTCNNIDAA KIYGMCFFSI TIDKFAIPNG RKVDLQMGNL GYLQSFNYRI    420
DTTATSCQLY YNLPASNVSI SRFNPSIWNR RFGFTEQSVF KPQPVGVFTD HDVVYAQHCF    480
KAPTNFCPCK LNGSLCVGSG FGIDAGYKNS GIGTCPAGTN YLTCYNANQC DCLCTPDPIL    540
SKSTGPYKCP QTKYLVGIGE HCSGLAIKSD YCGGNPCTCQ PKAFLGWSVD SCLQGDRCNI    600
FANFILHGVN SGTTCSTDLQ KSNTDIILGV CVNYDLYGIT GQGIFVEVNA TYYNSWQNLL    660
YDSNGNLYGF RDYLTNRTFM IRSCYSGRVS AGFHSNSSEP ALLFRNIKCN YVFNNTLSRQ    720
LQPINYFDSY LGCVVNADNS TSSSVQTCDL TVGSGYWGDY STQRRSRRTI TTGYRFTNFE    780
PFTVNPVNDS LHPVGGLYEI QIPSEFTIGN MEEFIQTRSP KVTIDCPVFV CGDYAACKSQ    840
LVEYGSFCDN INAILTEVNE LLDTTQLQVA NSLMNGVTLS TKLKDGFNFN VDDINFSPVL    900
GCLGSECNKV SSRSAIEDLL FSKVKLSDVG FVDAYNNCTG GAEIRDLICV QSYNGIKVLP    960
PLLSENQISG YTLAATFASL FPPWSAAAGV PFYLNVQYRI NGIGVTMDVL TQNQKLISNA   1020
FNNALDAIQE GFDATNSALV KIQAVVNANA EALNNLLQQL SNKFGAISAS LQEILSRLDA   1080
LEAQAQIDRL INGRLTALNA YVSQQLSDST LVKFSAAQAM EKVNECVKSQ SSRINFCGNG   1140
NHIISLVQNA PYGLYFIHFS YVPTKYVTAK VSPGLCIAGD RGIAPKSGYF VNVNNTWMFT   1200
GSGYYYPEPI TGNNVVVMST CAVNYTKAPD VMLNISTPNL PDFKEELDQW FKNQTLMAPD   1260
LSLDYINVTF LDLQDEMNRL QEAIKVLNHS YINLKDIGTY EYYVKWPWYV WLLIGLAGVA   1320
MLVLLFFICC CTGCGTSCFK KCGGCCDDYT GHQELVIKTS HDD                     1363
```

FIGURE 5

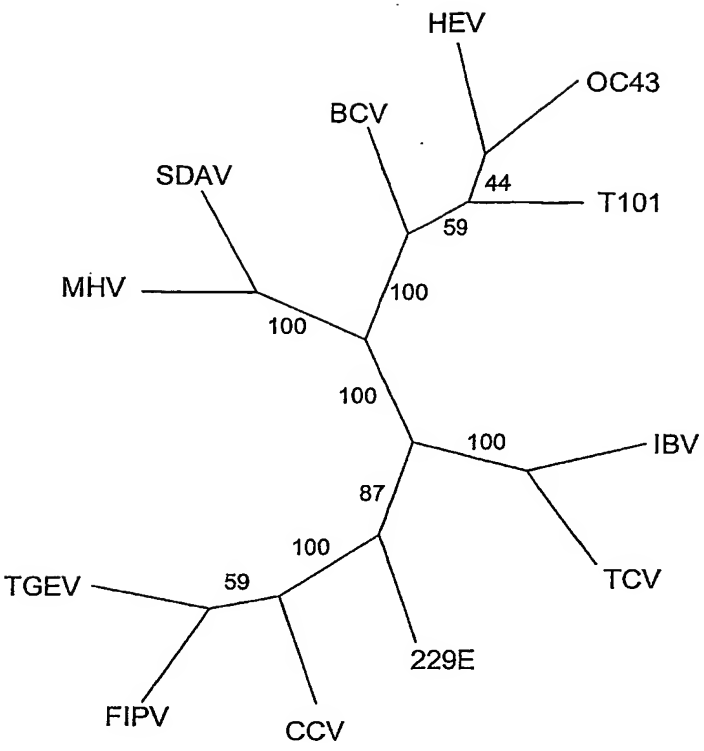

HEV
OC43
BCV
SDAV
44
T101
59
MHV
100
100
100
100
IBV
87
100
TCV
TGEV
59
229E
FIPV
CCV

FIGURE 6

```
T101 CTCAGATGAATTTGAAATATGCTATTAGTGCTAAGAATAGAGCCCGCACTGTTGCTGGTG
BCV  CTCAAATGAATTTGAAATATGCTATTAGTGCTAAGAATAGAGCCCGCACTGTTGCTGGTG
OC43 CTCAAATGAATTTGAAATATGCTATTAGTGCTAAGAATAGAGCCCGCACTGTTGCTGGTG
HEV  CTCAAATGAATTTGAAATATGCTATTAGTGCCAAGAATAGAGCCCGCACTGTTGCTGGTG
CCV  CTCAGATGAATTTGAAATATGCTATTTCTGGAAAGGCTAGAGCTCGTACAGTAGGAGGAG
     **** ********************  **  ***  ****** ** ** ** *  ** *

T101 TTTCCATACTTAGTACTATGACTGGCAGAATGTTTCATCAAAAATGTTTGAAAAGTATAG
BCV  TTTCCATACTCAGTACTATGACTGGCAGAATGTTTCATCAAAAATGTTTGAAAAGTATAG
OC43 TTTCCATACTTAGTACTATGACTGGCAGAATGTTTCATCAAAAATGTTTGAAAAGTATAG
HEV  TTTCCATACTTAGTACTATGACTGGCAGAATGTTTCATCAAAAATGCTTGAAAAGTATAG
CCV  TTTCACTTCTTTCTACCATGACTACGAGACAATACCACCAGAAGCATTTGAAGTCAATTG
     ****  * **   *** ******   ***   *  ** ** **    *****    ** *

T101 CAGCTACACGTGGTGTTCCTGTTGTTATAGGCACCACTAAATTTTATGGCGGCTGGGATG
BCV  CAGCTACACGTGGTGTTCCTGTTGTTATAGGCACCACTAAGTTTTATGGCGGCTGGGATG
OC43 CAGCTACACGTGGTGTTCCTGTAGTTATAGGCACCACTAAATTTTATGGTGGCTGGGATG
HEV  CAGCTACACGTGGCGTTCCTGTGGTTATAGGCACCACTAAATTTTATGGCGGCTGGGATG
CCV  CTGCAACACGCAATGCCACTGTGGTTATTGGCTCAACCAAGTTTTATGGTGGTTGGGATA
     * ** *****    *   **** ***** *** * ** ** ******** ** ******

T101 ATATGTTACGTCGCCTTATTAAAGATGTTGACAATCCTGTACTTATGGGTTGGGATTATC
BCV  ATATGTTACGTCGCCTTATTAAAGATGTTGATAATCCTGTACTTATGGGTTGGGATTATC
OC43 ATATGTTACGCCGCCTTATTAAAGATGTTGACAATCCTGTACTTATGGGTTGGGATTATC
HEV  ATATGTTACGCCGCCTTATTAAAGATGTTGATAATCCTGTACTTATGGGTTGGGATTATC
CCV  ACATGCTTAAAAATTTAATGCGTGATGTTGATAATGGTTGTTTGATGGGATGGGACTATC
     * *** *         * **    ********.***  *    * ***** ***** ****

T101 CTAAGTGTGA
BCV  CTAAGTGTGA
OC43 CTAAGTGTGA
HEV  CAAAGTGTGA
CCV  CTAAGTGTGA
     * ********
```

FIGURE 7

```
protHCVpoly     ---MNLKYAISAKNRARTVAGVSILSTMTGRMFHQKCLKSIAATR
protHEVpoly     ---MNLKYAISAKNRARTVAGVSILSTMTGRMFHQKCLKSIAATR
protBCVpoly     ---MNLKYAISAKNRARTVAGVSILSTMTGRMFHQKCLKSIAATR
protCRCVpol     --QMNLKYAISAKNRARTVAGVSILSTMTGRMFHQKCLKSIAATR
protCECVpol     MTQMNLKYAISGKARARTVGGVSLLSTMTTRQYHQKHLKSIAATR
                   ********.* *****.***:***** * :*** ********

protHCVpoly     GVPVVIGTTKFYGGWDDMLRRLIKDVDNPVLMGWDYPKC
protHEVpoly     GVPVVIGTTKFYGGWDDMLRRLIKDVDNPVLMGWDYPKC
protBCVpoly     GVPVVIGTTKFYGGWDDMLRRLIKDVDNPVLMGWDYPKC
protCRCVpol     GVPVVIGTTKFYGGWDDMLRRLIKDVENPVLMGWDYPKC--
protCECVpol     NATVVIGSTKFYGGWDNMLKNLMRDVDNGCLMGWDYPKC---
                ...****:********:**:.*::**:*  *********
```

FIGURE 8 (Page 1 of 9)

```
CRCVspike       ----------------------ATGTTTTTGATACTTTTA------ATTTCCTTACCAATG
CECVspike       ATGATTGTGCTCGTAACTTGCATTTTATTGTTATGTTCATACCACACTGCTTCGAGTACG
                        ** ** *** **  ** *      * * * *     * *

CRCVspike       GCTTTTGCTG-TTATAGGAGATTTAAAGTGTACTACGGTTTC-CATCAATGATGTTGACA
CECVspike       TCAAATAATGATTGTAGACAAGTTAA--CGTAACACAATTAGATGGCAATGAAAACCTCA
                 *   *  ** ** ***   * ****   ***  **  **      ******      **

CRCVspike       CCGGTG-CTCCTTCTATTAGCACTGATGTTGTCGATGTTACTAATGGTTTAGGTACTTAT
CECVspike       TTAGAGACTTTTTGTTTCAAAACTT-TAAAGAAGAAGGAACTGTAGTTGTTGGTGGTTAC
                   * * **  ** * * *  ***  *   *  ** *  ***   * * * ***  ***

CRCVspike       TATGTTTTAGA----TCGTGTG--TATTTAAATACTACA----TTGTTGCTTAATGGTTA
CECVspike       TACCCTACAGAGGTTTGGTATAACTGTTCTAGAACAGCAACAACTACTGCCTA-TGAGTA
                **   *  ***    * ** *   * **  *  **  **     *  *** ** **  **

CRCVspike       TTATCCTACTTCAGGTTCTACATATCGTAATATGGCA-CTGAAGGGAACTTTACTATTGA
CECVspike       TTTCAGTAATATACACGCATTCTATTTTGATATGGAAGCCATGGAGAATAGTACTGGTAA
                **    ** *  *    *    ***  * ****** * *    * ***   ****  * *

CRCVspike       -GCACACTATGG-TTTAAACCACCATTTCTTTCTGATTTTATTGATGGTGTTTTTGCTAA
CECVspike       TGCACGTGGTAAACCTTTATTATTTCATGTTCATGGTGAGCCTGTTAGTGTCATCATATA
                 ****    *     *  *  *     * **  ** *     ** * ****  *     *

CRCVspike       GGTAAAAAATACCAAGGTTATTAAAGATGGTGTAGTGTATAG---TGAGTTTCCTGCTAT
CECVspike       CATATCTTATAGAGATGATGTGCAACATAGGCCACTTTTAAAACACGGATTAGTGTGCAT
                  **   ***   * * * *  ** ** *   * * *  *     *  **       **

CRCVspike       AACTATAGGTAGTACTTTTG--TA-AATACATCCTATAGTGTGGTAGTACAACCACATAC
CECVspike       AACTGAAAGTCGCAACATTGACTATAACAGTTTCACCAGTA-GCCAGTGGAATTCCATAT
                ****  * ** * *   ***  ** ** * * *   ***  *  ***  **   ****

CRCVspike       -TACTAATTTAGATAATAAATTACAAGGTCTCTTAGAGATCTCTGTTTGCCAGTATACTA
CECVspike       GTACGGGTAATGACAGAAAAATTCCTT-TCTCTGTCATACCCACGGACAATGGAACAAAA
                 ***   *   ** *  *** * *    *****   * * *   *       * * *  *
```

FIGURE 8 (Page 2 of 9)

```
CRCVspike       -TGTGCGATTACCCACATA-CGATGTGTC-ATCCTAATCTGGGT-AATAAACG--CATAG
CECVspike       ATTTATGGTCTTGAGTGGAATGATGAATTTGTTACAGCGTACATTAGTGGTCGTTCTTAT
                 * *  * *        *  ****  *   *  *   *   * * *   **  * **

CRCVspike       AACTATGGCATTGGGATACAGGTGTTGTTCCCTGTT-TATATAAGCGTAATTTCACATAT
CECVspike       AATTGGAACATCAATAATAATTGGTTTAACAATGTCACGCTTCTGTATAGTCGCTCAAGC
                ** *    ***    *   *   ***   *  ***      *  *  ** *  * **

CRCVspike       GATGTGA-ATGCTGATTATTTGTATTCCCATTTTTATCAAGAAGGTGGTACTTT---TTA
CECVspike       ACTGCCACATGGCAACACAGTGC-TGCATACGTTTACCAAGGTGTTTCTAACTTCACTTA
                  **  * ***   *     **  * *  *  **** ****  * *  **  **   ***

CRCVspike       TGCATATTTTACAGACACTGGTGTTGTTACTAAGTTTCTGTTTCATGTTTAT-TTAGGCA
CECVspike       TTACAAGTTAAATAACACCAATGGTCTAA--AAACCTATGAATTATGTGAAGATTATGAA
                *    * ** *   ****   ** * * *  **   * **  * ****  *  *** * *

CRCVspike       CGGTGCTTT---CACATTATTA-TGTCATGCCCTTGACTTGTAATAGTGCTATGACTTTA
CECVspike       TATTGCACTGGCTACGCCACTAACATCTTTGCCCCAACTGTGGGAGGTTACATACCTGAT
                   ***  *    **   * **   ** *  **   ***       **   **  **

CRCVspike       GAATACTGGGTTA-----CACCTCTCACTTTTAAACAATATTTACTCGCTTTCAATCAAG
CECVspike       GGATTTAGTTTTAACAATTGGTTTTTGCTTACAAACAGCTCCACTTTTGTTAGTGGCAGA
                * **   *  ***         * *  ***  *****        *   **     **

CRCVspike       ATGGTGTTATTTTTAATGCTGTTGATTGTAAGAGTGATTTTATGAGTGAGATTAAGTGT-
CECVspike       TTTGTAACAAATCAACCATTATTAGTTAATTGCTTGTGGCCAGTTCCTAGTTTTGGTGTT
                 * **   *  *  *    * **  **    *  **     *      ** **  ****

CRCVspike       --AAAACACTATCTATAGCACCATCTACTGGTGTTTATGAATTAAACGGTTACACTGTTC
CECVspike       GCAGCACAAGAATTTTGTTTTGAAGGTGCACAGTTTAGCCAATGTAATGGTGTGTTTTTA
                  *  ***  *  * *      *         *****   * *  *  * *    * **

CRCVspike       AGCCA-ATTGCAGATGTTTACCGACGTATACCTAATCTTCCCG--ATTGTAATATAGAGG
CECVspike       AATAACACAGTAGATGTCATTAGATTCAACCTTAATTTTACTGCAGATGTACAATCTGGC
                *   * *  * ******     **   *  * **** ** * *    ****  **   *
```

FIGURE 8 (Page 3 of 9)

```
CRCVspike       CTTGGCTTAATGATAAGT-CGGTGCCTTCTCCATTAAATTGGGAACGTAAGACCTTTTCA
CECVspike       ATGGGTGCTACAGTATTTTCACTGAATACAACAGGTGGTTGCATTCTTGAGATTTCTT--
                 * **    *   **  * *  **  * *  **     ***    * * ***  * **

CRCVspike       AATTGTAATTTTAATATGAGCAGCCTGATGTCTTTTATCCAGGCTGACTCGTTTACTTGT
CECVspike       -GTTATAATGATATAGTGAGCGAGTCAAGTTTCTACAGTTATGGTGA---AATTCCCTTC
                  ** ****  **   *****     *  *  *  *   * * ***     ** * *

CRCVspike       AATAATATTGATGCTGCTAAGATATACGGTATGTGTTTTTTCA--GCATAACTATAGATA
CECVspike       GGCGTAACTGATGG-ACCGCGTTAT-TGTTATGTCCTCTATAATGGCACAGCTCTTAAGT
                      * *****   *   * ***  * *****  * * * *  *** * ** *  *

CRCVspike       AGTTTGCTA---TACCCAATGGTAGGAAGGTTGACCTACAAATGGGCAATTTGGGCTATT
CECVspike       ATTTCGGCACATTACCCCCTAGTGTCAAGG--AAATTGCTATTAG-TAAGTGGGGCCAAT
                * ** *  *   *****  * **   ****   *  * * * * *  ** * **** * *

CRCVspike       TGCAGTCTTTTAACTATAGAATTGATACTACTGCTACAAGTTGTCAGTTGTATTATAATT
CECVspike       TTTATATTAATGGTTACAATTTCTTTAGCACTTTTCCTATTGATTGTATATCTTTTAACT
                *  *   *  *   ** *   *   **  ***  * * * *  *    * * ** *** *

CRCVspike       TACCTGCTAGTAATGTTTCTATTAGCAGGTTTAATCCTTCTATTTGGAATA--GGAGATT
CECVspike       TAACCACTGGTGATAGTGGAGCATTTTGGACAATTGCTTACACATCGTACACTGAAGCAT
                ** *  ** ** **  *          **   * * ***  *  * * * *  * **  *

CRCVspike       TGGTTTTA-CAGAACAATCTGTTTTTAAGCCT-CAACCTGTAGGTGTTTTTACTGATCAT
CECVspike       TAGTACAAGTTGAAAACACAGCCATTAAAAAGGTGACGTATTGTAACAGTCAC-ATTAAT
                * **   *   *** *  * *   ****       ** * * *      * **   * **

CRCVspike       GATGTTGTTTATGCACAACATTGTTTTAAAGCTCCCACAAATTTCTGTCCG-----TGTA
CECVspike       AACATCAAATGTTCTCAACTTACTGCTAATTTGCAAAATGGCTTTTATCCTGTTGCTTCA
                 *  *    * * * **** *  *  ***    *  *     ** * ***      *  *

CRCVspike       AATTGAATGGGTCTTTGTGTGTAGGTAGTGGTTTTGGTA--TAGATGCTGGTTATAAA--
CECVspike       AGTGAAGTTGGTCTTGTCAATAAGAGTGTTGTGTTACTACCTAGTTTCTATTCACATACC
                * *  * * ******      **   ** ** **  **  *** * **  * * * *
```

FIGURE 8 (Page 4 of 9)

```
CRCVspike       AATAGTGGTATAGGCACTTGTCCTGCAGGTACTAATTATTTAACTTGTTATAATGCTAAC
CECVspike       AGTGTTAATATAACTATTGATCTTG---GTATGAAGCGTAGTGGTTATGGTCAACCCA--
                * *  *  ****   * *  ** **   ***  **   *     ** *  * *  * *

CRCVspike       CAATGTGATTGTTTGTGCACTCCAGAC--CCTATTTTATCTAAATCTACAGGGCCTTA-T
CECVspike       TAGCCTCAACACTAAGTAACATCACACTACCAATGCAGGATAATAACACCGATGTGTACT
                 *   * *    *     **  ** **  ** **   .  ***    ** *     ** *

CRCVspike       AAGTGCCCCCAAACTAAATACTTAGTTGGCATAGGTGAGCACTGTTCTGGTCTTGCTATT
CECVspike       GTATTCGTTCTAACCAATT-CTCAGTTTATGTTCACTCCACTTGCAAAAGTTCTTTATGG
                   * *   * *** ** * ** ****    *          **     **  *

CRCVspike       AAAAGTGATTATTGTGGAGGCAATCCTTGTACTTGCCAACCAAAAGCATTTTTGGG--TT
CECVspike       GACAACAATTTTAATCAAGATTGCACAGATGTTTTATATGCCACAGCTGTTATAAAAACT
                 * *   *** *  *  **      *   *  **   *  * * ***  ** *      *

CRCVspike       GGTCTGTGGAC--TCTTGTTTACAAGG--GGATAGGTGTAATATTTTTGCTAA-TTTTAT
CECVspike       GGTACTTGCCCCTTCTCATTTGATAAATTGAATAATTACTTAACTTTTAACAAGCTTTGT
                ***   **  *  ***  ***   *    * ***  *     * ****   **  *** *

CRCVspike       TTTGCATGGTGT--TAATAGTG------GTACTACTTGTTCTACTGATT-TACAAAAATC
CECVspike       TTGTCGTTGAATCCTACTGGTGCCAACTGTAAGTTTGATGTTGCTGCCCGTACAAGAACC
                **  * * *  *  ** * ***      ***    *  *  * ***    ***** ** *

CRCVspike       AAACACAGACATAATTCTTGGTGTTTGTGTTAATTATGATCTTTATGGTATTACAGGCCA
CECVspike       AA-TGAGCAGGTTGTTAGAAGTTTATATGTAATATATGAAGAAGGAGACAACATAGTGGG
                **      *  *  **    ** * * *** *  *****       *  *  * **

CRCVspike       AGGTATTTTTGTTGA----GGTTAATGCGACTTATTATAATAGTTGGCAGAACCTTTTAT
CECVspike       TGTACCGTCTGATAATAGTGGTCTTCACGATTTGTCAGTGTTACACTTAGACTCCTGTAC
                 *     * ** * *    ***     *** ** * *   *       ***  * * **

CRCVspike       ATGATTCTAATG---GTAATCTCTATGGTTTTAGGGACTACTTAACAAACAGA-ACTTTT
CECVspike       A-GATTACAATATATATGGTAGAACTGGTGTT-GGTATTATTAGACAAACTAACAGCACA
                * ****  ***     *  *     **** ** ** * ** *  ******  * *
```

FIGURE 8 (Page 5 of 9)

```
CRCVspike       ATGATTCGTAGTTGCTATAGCG-GTCGTGTTTCAGCGGGCTTTCA---CTCTAACTCTTC
CECVspike       ATACTTAGTGGCTTACATTATACATCACTATCAGGTGATTTATTAGGTTTTAAAAATGTT
                **  ** ** * *   **      **    *   * *   * * *    *  **    *

CRCVspike       CGAACCAGCATTG-CTATTTCGGAATATTAAATGCAATTACGTTTTTAATAATACTCTTT
CECVspike       AGTGATGGTGTTGTCTATTCTGTGACACCATGTGATGTAAGCGCACAAGCGGCTGTTATT
                 *     *  *** *****  *  * *  *  **   * *       *       *  **

CRCVspike       CACG-----ACAGCTGCAACCTATTAACTATTTTGATAGTTATCTTGGTTGTGTTGTCAA
CECVspike       GATGGGGCCATAGTTGGAGC-TATGACTTCCATTAATAGTGAACT-GTTAGGTCTAACAC
                 * *     * ** ** * * *** *  *   ** ***** * ** * * *   *  **

CRCVspike       TGCTGATAATAGTAC-----TTCTAGTTCTGTTCAAACATGTGATCTCACAGTAGGTAGT
CECVspike       ATTGGACAACAACACCAAATTTTTATTACTACTCTA-TATATAAT---ACAACAAATGAG
                    ** ** *  **     ** ** * **  ** *  ** * **   ***  *  *

CRCVspike       GGTTACTGGGGGGATTACTCTACACAAAGACGAAGT----CGTAGAACGATTACCACTGG
CECVspike       AGA-ACTCGTGGCACTGCAATCGACAGTAACGATGTAGATTGTGAACCTATCATAACCTA
                 *  *** * ** * * *  *  ***   **** **     **  * * ** *  **

CRCVspike       TT-----ATCGGTTT----ACTAATTTTGAGCCATTTACTGTTAATCCAGTAAATGATAG
CECVspike       TTCTAACATAGGTGTTTGTAAAAATGGTGCGTTGGTTTTTATTAACGTCACACATTCTGA
                **     ** *** *    *  ***  ** *    **  * ****      * **  *

CRCVspike       TTTACACCCTGTAGGTGGTTTGTAT--GAAAT-TCA-AATACCTTCAGAGTTTACTATAG
CECVspike       TGGAGATGTT-CAACCAATTAGCACTGGCAATGTCACGATACCCACAAACTTTACCATAT
                *  * *   *  *     ** * *   * *** ***  *****  ** * ***** ***

CRCVspike       GTAATATGGAGGAGTTTATTCAAACAAGATCTCCTAAAGTTACTATTGATTGTCCTGTTT
CECVspike       CTGTGCAAGTTGAATACATCCAGGTTTACACTACACCGGTGTCAATAGATTGTTCTAGAT
                 *      *  ** *  ** **       ** *    **  * ** ****** **   *

CRCVspike       TTGTCTGTGGTGATTATGCAGCATGTAAATCACAGTTGGTTGAATATGGTAGTTTTTGTG
CECVspike       ACGTTTGTAATGGTAACCCTAGATGTAATAAATTGTTAACACAATATGTTTCTGCATGTC
                  ** ***  ** * *  *   ******   *  ***     ****** *  *   ***
```

FIGURE 8 (Page 6 of 9)

```
CRCVspike       ACAATATTAATGCTATACTCACAG-AAGT----------AAATGAACTACTTGACACTA
CECVspike       AAACTATTGAGCAAGCGCTTGCAATGAGTGCCAGCCTTGAAAACATGGAAGTTGATTCCA
                * * **** *       **  **   ***          ***      * ****  * *

CRCVspike       CACAGTTGCAAGTAGCTAATAGTTTAATGAATGGTGTCACTCTTAGCACTAAGCTTAAAG
CECVspike       TGTTGTTTGTTTCAGAAAATGCCCTTA-AATTGGCATCTGTTGAGGCGTTCAATAGTACA
                   ***      **  ***    * *  * ***  **  *  .  **  * *     *

CRCVspike       ATGGCTTTAATTTCAATGTAGATGACAT----CAATTT---TTCCCCTGTATTAGGTTGT
CECVspike       GAACATTTAGATCCTATTTACAAAGAATGGCCTAACATAGGTGGTTCTTGGCTAGGAGGT
                     ****  * * ** ** *    **     **  *   *    **    ****  **

CRCVspike       TTAGGAAGCGAAT---------GTAATAA-AGTTTCCAGTA--GATCTGCTATAGAGGAT
CECVspike       CTAAAAGACATACTTCCGTCCCATAATAGCAAACGTAAGTATCGTTCTGCTATAGAAGAC
                 **  *  *  *          *****  *      ****  * ********** **

CRCVspike       TTACTTTTTTCTAAAGTAAAGTTATCTGATGTTGGTTTTGTTGATGC---TTATAATAAT
CECVspike       TTGCTTTTTGATAAAGTTGTAACTTCTGGTCTAGGTACAGTTGATGAAGATTATAAACGT
                ** ******  ******       **** * * ***   *******    ******   *

CRCVspike       TGTACTGGAGGTGCCGAAATTAGGGACCTCATTTGTGTGCAAAGTTATAATGGTATCAAA
CECVspike       TGTACAGGTGGTTATGACATAGCTGACTTAGTTTGTGCACAATATTACAATGGCATCATG
                ***** ** ***   ** **    *** *  ******  ***  *** ***** ****

CRCVspike       GTGTTGCCTC-CACTGCTCTCAGAAAATCAGATCAGTGGATACACTTTGGCTGCCACCTT
CECVspike       GTTCTACCTGGTGTTGCTAAT-GATGACAAGATGACTATGTACACAGCCTCTCTTGCAGG
                **  * ***     ****    **  *  **** * *   *****     **    *

CRCVspike       TGCTAGTCTGTTTCCTCC-TTGGTCAGCAGCA--GCAGGCGTACCATTTTATTTAAATGT
CECVspike       TGGTATAGCATTAGGTGCACTAGGTGGTGGCGCCGTGGCTATACCTTTTGCAGTAGCAGT
                ** **     **   * *  * *   *  **   *  *   **** ***    **   **

CRCVspike       TCAGTATCGTATTAATGGTATTGGTGTTACCATGGATGTGCTAACTCAAAATCAAAAGCT
CECVspike       TCAGGCTAGACTTAATTATGTTGCTCTACAAACTGATGTATTGAACAAAAACCAGCAGAT
                ****  * *  *****  * *** * *    *  *****  * *   **** **  ** *
```

FIGURE 8 (Page 7 of 9)

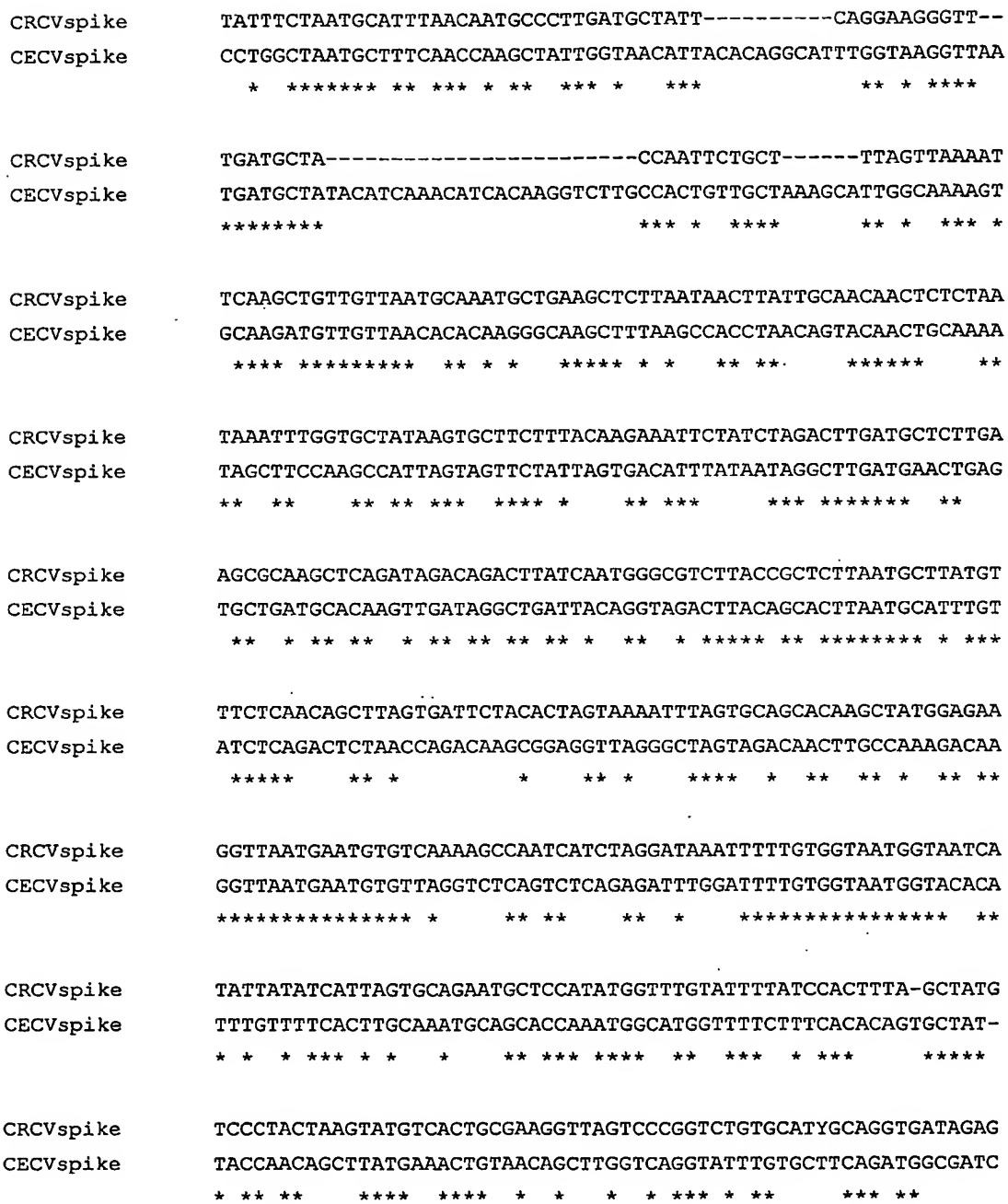


FIGURE 8 (Page 8 of 9)

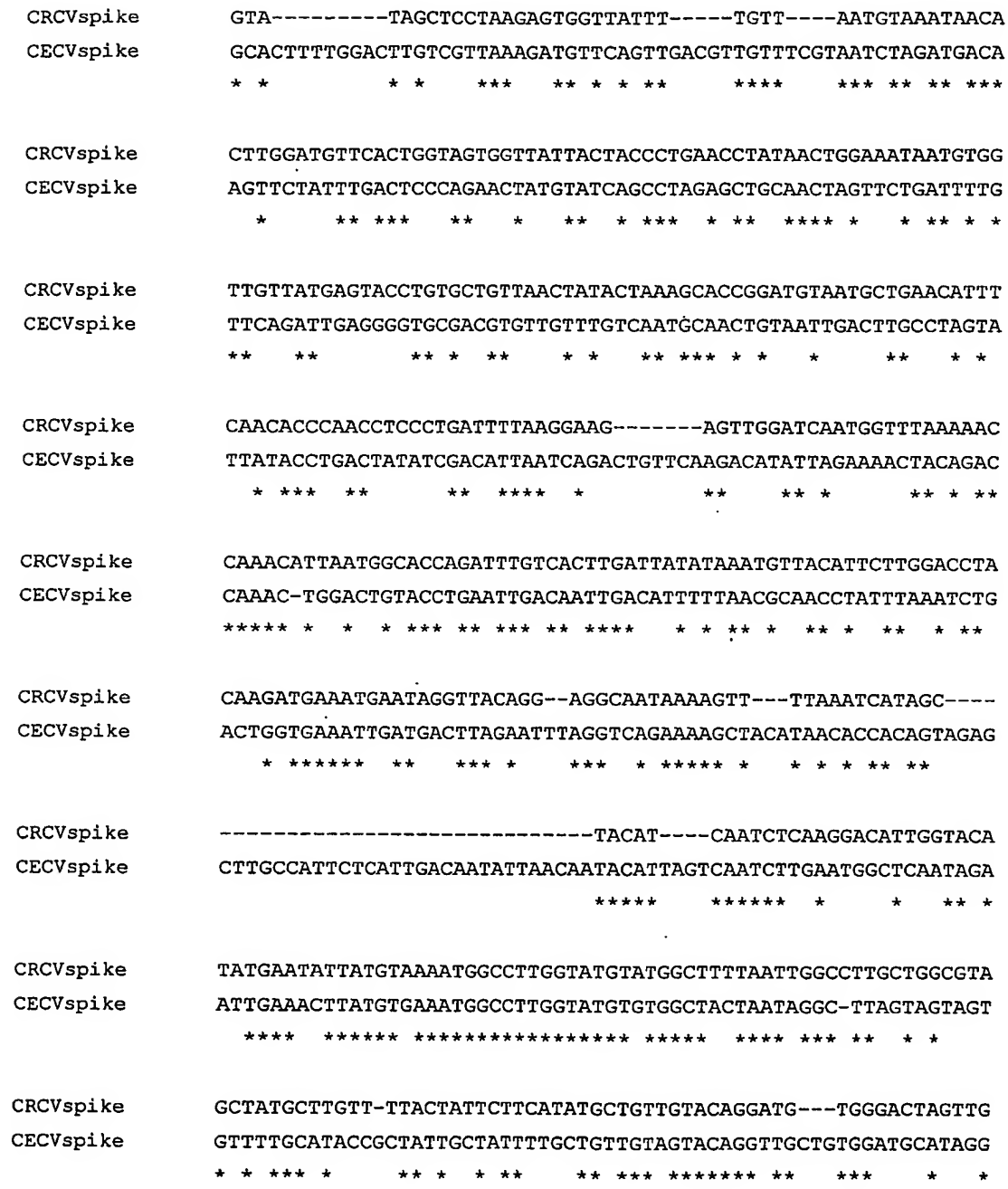


FIGURE 8 (Page 9 of 9)

```
CRCVspike       TTTTAAGAAATGCGGTGGTTGTTGTGATGATTATACTGGACA--TCAGGAGTTAGTAATC
CECVspike       TTGTTTGGGAAGTTGTTGTCATTCTATTTGTAGTAGAAGACAATTTGAAAATTACGAACC
                ** *  *  * *  ** **  ** *  *  *  **   ****  *    * ***  ** *

CRCVspike       AA----AACGTCACATGACGACTAA-----------------------------------
CECVspike       AATTGAAAAAGTGCATGTCCACTAAA-----------------------------------
                **    **     **** * *****
```

FIGURE 9 (Page 1 of 12)

```
BCVspike                ATGTTTTTGATACTTTTAATTTCCTTACCAATGGCTCTTGCTGTTATAG
HCVspike        -----------ATGTTTTTGATACTTTTAATTTCCTTACCAACGGCTTTTGCTGTTATAG
CRCVspike       -----------ATGTTTTTGATACTTTTAATTTCCTTACCAATGGCTTTTGCTGTTATAG
HEVspike        -------     ATGTTTTTTATACTTTTAATCACCCTGCCTTCTGTTTTTGCAGTTATAG
                            ******** ***********  ** * **    * * **** *******

BCVspike        GAGATTTAAAGTGTACTACGGTTTCCATTAATGATGTTGACACCGGTGTTCCTTCTGTTA
HCVspike        GAGATTTAAAGTGTACTACGGTTTCCATTAATGATATTGACACCGGTGCTCCTTCTATTA
CRCVspike       GAGATTTAAAGTGTACTACGGTTTCCATCAATGATGTTGACACCGGTGCTCCTTCTATTA
HEVspike        GGGATTTAAAGTGTAATACTTCATCAATTAATGACGTTGACACTGGTGTGCCATCTATTA
                * ************* ***    ** ** *****  ******* ****  ** *** ***

BCVspike        GCACTGATACTGTCGATGTTACTAATGGTTTAGGTACTTATTATGTTTTAGATCGTGTGT
HCVspike        GCACTGATATTGTCGATGTTACTAATGGTTTAGGTACTTATTATGTTTTAGATCGTGTGT
CRCVspike       GCACTGATGTTGTCGATGTTACTAATGGTTTAGGTACTTATTATGTTTTAGATCGTGTGT
HEVspike        GCTCTGAAGTTGTTGATGTCACTAATGGTTTGGGGACTTTCTATGTTTTAGATCGTGTCT
                ** ****   *** ***** ********** ** ****  ***************** *

BCVspike        ATTTAAATACTACGTTGTTGCTTAATGGTTACTACCCTACTTCAGGTTCTACATATCGTA
HCVspike        ATTTAAATACTACGTTGTTGCTTAATGGTTACTACCCTACTTCAGGTTCTACATATCGTA
CRCVspike       ATTTAAATACTACATTGTTGCTTAATGGTTATTATCCTACTTCAGGTTCTACATATCGTA
HEVspike        ATTTAAATACCACATTGTTGCTCAATGGTTATTACCCAATTTCAGGTGCTACATTTCGTA
                ********** ** ******** ******** ** ** * ******* ****** *****

BCVspike        ATATGGCACTGAAGGGAACTTTACTATTGAGCACACTATGGTTTAAACCACCTTTTCTTT
HCVspike        ATATGGCACTGAAGGGAACTTTACTATTGAGCAGACTATGGTTTAAACCACCTTTTCTTT
CRCVspike       ATATGGCACTGAAGGGAACTTTACTATTGAGCACACTATGGTTTAAACCACCATTTCTTT
HEVspike        ATGTGGCTCTGAAAGGAACTCGATTATTGAGCACCTTGTGGTTTAAGCCGCCTTTTTTAT
                ** **** ***** ******  * *********   * ******** ** ** *** * *

BCVspike        CTGATTTTATTAATGGTATTTTTGCTAAGGTCAAAAATACCAAGGTTATTAAAAATGGTG
HCVspike        CTGATTTTATTAATGGTATTTTTGCTAAGGTCAAAAATACCAAGGTTATTAAAAAGGGTG
CRCVspike       CTGATTTTATTGATGGTGTTTTTGCTAAGGTAAAAAATACCAAGGTTATTAAAGATGGTG
HEVspike        CACCTTTTAATGATGGTATTTTTGCCAAGGTTAAAAACAGCAGATTTTCTAAACATGGTG
                *   ***** * ***** ******* ***** ***** * **   **  **** * ****
```

FIGURE 9 (Page 2 of 12)

```
BCVspike        TAATGTATAGTGAGTTTCCTGCTATAACTATAGGTAGTACTTTTGTAAATACATCCTATA
HCVspike        TAATGTATAGTGAGTTTCCTGCTATAACTATAGGTAGTACTTTTGTAAATACATCCTATA
CRCVspike       TAGTGTATAGTGAGTTTCCTGCTATAACTATAGGTAGTACTTTTGTAAATACATCCTATA
HEVspike        TTATTTATAGTGAGTTTCCTGCTATTACTATAGGTAGTACTTTTGTAAATACTTCCTATA
                *  * ******************** ************************** *******

BCVspike        GTGTGGTAGTACAACCACATACTACCAATTTAGATAATAAATTACAAGGTCTCTTAGAGA
HCVspike        GTGTGGTAGTACAACCACATACTACCAATTTGGATAATAAATTACAAGGTCTCTTAGAGA
CRCVspike       GTGTGGTAGTACAACCACATACTACTAATTTAGATAATAAATTACAAGGTCTCTTAGAGA
HEVspike        GCATAGTAGTAAAGCCTCATACCTCATTTATTAATGGTAATTTACAAGGTTTTTTGCAAA
                *  * ****** * ** *****  *   * *  **  *** ********* * **  * *

BCVspike        TCTCTGTTTGCCAGTATACTATGTGCGAGTACCCACATACGATTTGTCATCCTAATTTGG
HCVspike        TCTCTGTTTGCCAGTATACTATGTGCGAGTACCCACATACGATTTGTCATCCTAATCTGG
CRCVspike       TCTCTGTTTGCCAGTATACTATGTGCGATTACCCACATACGATGTGTCATCCTAATCTGG
HEVspike        TTTCTGTTTGTCAATATACTATGTGTGAATACCCACAGACTATTTGTCATCCTAATTTGG
                * ******** ** *********** ** ******** ** ** ************ ***

BCVspike        GTAATCGGCGCATAGAACTATGGCATTGGGATACAGGTGTTGTTTCCTGTTTATATAAGC
HCVspike        GTAATCGACGCGTAGAACTATGGCATTGGGATACAGGTGTTGTTTCCTGTTTATATAAGC
CRCVspike       GTAATAAACGCATAGAACTATGGCATTGGGATACAGGTGTTGTTCCCTGTTTATATAAGC
HEVspike        GTAATCAACGCATAGAATTATGGCATCATGACACAGATGTTGTTTCTTGTTTATACAGGC
                *****   *** ***** ********   ** **** ******* * ******** * **

BCVspike        GTAATTTCACATATGATGTGAATGCTGATTATTTGTATTTCCATTTTTATCAAGAAGGTG
HCVspike        GTAATTTCACATATGATGTGAATGCTGATTACTTGTATTTCCATTTTTATCAAGAAGGTG
CRCVspike       GTAATTTCACATATGATGTGAATGCTGATTATTTGTATTCCCATTTTTATCAAGAAGGTG
HEVspike        GTAATTTCACATATGATGTGAATGCTGATTATTTATATTTTCACTTTTATCAGGAAGGTG
                ******************************** ** ****  ** ******** *******

BCVspike        GTACTTTTTATGCATATTTTACAGACACTGGTGTTGTTACTAAGTTTCTGTTTAATGTTT
HCVspike        GTACTTTTTATGCATATTTTACAGACACTGGTGTTGTTACTAAGTTTCTGTTTAATGTTT
CRCVspike       GTACTTTTTATGCATATTTTACAGACACTGGTGTTGTTACTAAGTTTCTGTTTCATGTTT
HEVspike        GCACTTTTTATGCATACTTTACAGATACTGGTTTTGTGACCAAGTTTCTGTTTAAGTTGT
                * ************** ******** ****** **** ** ************ *  * *
```

FIGURE 9 (Page 3 of 12)

```
BCVspike        ATTTAGGCACGGTGCTTTCACATTATTATGTCATGCCTTTGACTTGTAATAGTGCTATGA
HCVspike        ATTTAGGCACGGTGCTTTCACATTATTATGTCCTGCCTTTGACTTGTAATAGTGCTATGA
CRCVspike       ATTTAGGCACGGTGCTTTCACATTATTATGTCATGCCCTTGACTTGTAATAGTGCTATGA
HEVspike        ATTTAGGCACTGTGCTGTCACACTATTATGTTATGCCATTGACTTGTGATAGCGCTTTAT
                ********** ***** ***** ********  **** ********* **** *** *

BCVspike        CTTTAGAATATTGGGTTACACCTCTCACTTCTAAACAATATTTACTCGCTTTCAATCAAG
HCVspike        CTTTAGAATATTGGGTTACACCTCTCACTTCTAAACAATATTTACTAGCTTTCAATCAAG
CRCVspike       CTTTAGAATACTGGGTTACACCTCTCACTTTTAAACAATATTTACTCGCTTTCAATCAAG
HEVspike        CTTTAGAATATTGGGTTACACCTCTCACTACTAGACAATTTCTTCTAGCCTTTGACCAGG
                ********** ******************  ** ***** * * ** ** **  * ** *

BCVspike        ATGGTGTTATTTTTAATGCTGTTGATTGTAAGAGTGATTTTATGAGTGAGATTAAGTGTA
HCVspike        ATGGTGTTATTTTTAATGCTGTTGATTGTAAGAGTGATTTTATGAGTGAGATTAAGTGTA
CRCVspike       ATGGTGTTATTTTTAATGCTGTTGATTGTAAGAGTGATTTTATGAGTGAGATTAAGTGTA
HEVspike        ATGGTGTTTTATACCATGCTGTTGATTGTGCTAGTGATTTTATGAGTGAGATTATGTGTA
                ******** * *   **************   ********************** *****

BCVspike        AAACACTATCTATAGCACCATCTACTGGTGTTTATGAATTAAACGGTTACACTGTTCAGC
HCVspike        AAACACTATCTATAGCACCATCTACTGGTGTTTATGAATTAAACGGTTACACTGTTCAGC
CRCVspike       AAACACTATCTATAGCACCATCTACTGGTGTTTATGAATTAAACGGTTACACTGTTCAGC
HEVspike        AAACTTCTTCAATTACACCACCTACTGGTGTTTATGAACTAAACGGTTACACAGTTCAAC
                ****    ** **  ***** ***************** ************* ***** *

BCVspike        CAATTGCAGATGTTTACCGACGTATACCTAATCTTCCCGATTGTAATATAGAGGCTTGGC
HCVspike        CAATTGCAGATGTTTACCGACGTATACCTAATCTTCCCGATTGTAATATAGAGGCTTGGC
CRCVspike       CAATTGCAGATGTTTACCGACGTATACCTAATCTTCCCGATTGTAATATAGAGGCTTGGC
HEVspike        CTGTTGCCACTGTGTATCGTAGAATACCTGACTTACCCAATTGCGATATCGAAGCTTGGC
                *  ****   *** ** **  * ****** *  * *** ****  **** ** *******

BCVspike        TTAATGATAAGTCTGTGCCCTCTCCATTAAATTGGGAACGTAAGACCTTTTCAAATTGTA
HCVspike        TTAATGATAAGTCGGTGCCCTCTCCATTAAATTGGGAACGTAAGACCTTTTCAAATTGTA
CRCVspike       TTAATGATAAGTCGGTGCCTTCTCCATTAAATTGGGAACGTAAGACCTTTTCAAATTGTA
HEVspike        TTAATTCTAAGACCGTTTCTTCGCCTCTTAATTGGGAACGTAAAATTTTTTCTAATTGTA
                ***** **** * **  * ** **  * ************** *  ***** *******
```

FIGURE 9 (Page 4 of 12)

```
BCVspike        ATTTTAATATGAGCAGCCTGATGTCTTTTATTCAGGCAGACTCATTTACTTGTAATAATA
HCVspike        ATTTTAATATGAGCAGCCTGATGTCTTTTATTCAGGCAGACTCATTTACTTGTAATAATA
CRCVspike       ATTTTAATATGAGCAGCCTGATGTCTTTTATCCAGGCTGACTCGTTTACTTGTAATAATA
HEVspike        ATTTTAACATGGGCAGGCTGATGTCTTTTATTCAGGCTGACTCTTTTGGTTGTAACAATA
                ******* *** **** ************** ***** ***** ***  ****** ****

BCVspike        TTGATGCAGCTAAGATATATGGTATGTGTTTTTCCAGCATAACTATAGATAAGTTTGCTA
HCVspike        TTGATGCTGCTAAGATATATGGTATGTGTTTTTCCAGCATAACTATAGATAAGTTTGCTA
CRCVspike       TTGATGCTGCTAAGATATACGGTATGTGTTTTTTCAGCATAACTATAGATAAGTTTGCTA
HEVspike        TTGATGCTTCTCGCTTATATGGTATGTGTTTTGGTAGCATTACTATTGACAAGTTTGCTA
                *******  **    **** ************   ***** ***** ** **********

BCVspike        TACCCAATGGTAGGAAGGTTGACCTACAATTGGGCAATTTGGGCTATTTGCAGTCTTTTA
HCVspike        TACCCAATGGTAGGAAGGTTGACCTACAATTGGGCAATTTGGGCTATTTGCAGTCTTTTA
CRCVspike       TACCCAATGGTAGGAAGGTTGACCTACAAATGGGCAATTTGGGCTATTTGCAGTCTTTTA
HEVspike        TACCCAATAGTAGAAAGGTTGATCTGCAAGTGGGTAAATCTGGTTATTTACAATCTTTTA
                ******** **** ******** ** *** **** ** *  ** ***** ** *******

BCVspike        ACTATAGAATTGATACTACTGCTACAAGTTGTCAGTTGTATTATAATTTACCTGCTGCTA
HCVspike        ACTATAGAATTGATACTACTGCTACAAGTTGTCAGTTGTATTATAATTTACCTGCTGCTA
CRCVspike       ACTATAGAATTGATACTACTGCTACAAGTTGTCAGTTGTATTATAATTTACCTGCTAGTA
HEVspike        ATTATAAGATTGACACTGCTGTTAGCAGTTGTCAACTCTATTATAGTTTGCCTGCAGCAA
                * ****  ***** *** *** **  ********  * ******* *** *****    *

BCVspike        ATGTTTCTGTTAGCAGGTTTAATCCTTCTACTTGGAATAGGAGATTTGGTTTTACAGAAC
HCVspike        ATGTTTCTGTTAGCAGGTTTAATCCTTCTACTTGGAATAGGAGATTTGGTTTTACAGAAC
CRCVspike       ATGTTTCTATTAGCAGGTTTAATCCTTCTATTTGGAATAGGAGATTTGGTTTTACAGAAC
HEVspike        ACGTATCTGTCACTCATTATAATCCTTCATCTTGGAACAGAAGGTATGGGTTTAT----T
                * ** *** * *     * *********   ****** ** ** * *** ****

BCVspike        AATCTGTTTTTAAGCCTCAACCTGTAGGTGTTTTTACTGATCATGATGTTGTTTATGCAC
HCVspike        AATCTGTTTTTAAGCCTCAACCTGTAGGTGTTTTTACTCATCATGATGTTGTTTATGCAC
CRCVspike       AATCTGTTTTTAAGCCTCAACCTGTAGGTGTTTTTACTGATCATGATGTTGTTTATGCAC
HEVspike        AATCAGAGTTTTGGTTCCAG-----AGGC-CTT--------CATGATGCTGTATATTCAC
                **** *  ***  *   **      ***   **        ******* *** *** ***
```

FIGURE 9 (Page 5 of 12)

```
BCVspike        AACATTGTTTTAAAGCTCCCACAAATTTCTGTCCGTGTAAATTGGATGGGTCTTTGTGTG
HCVspike        AACATTGTTTTAAAGCTCCCACAAATTTCTGTCCGTGTAAATTGGATGGGTCTTTGTGTG
CRCVspike       AACATTGTTTTAAAGCTCCCACAAATTTCTGTCCGTGTAAATTGAATGGGTCTTTGTGTG
HEVspike        AGCAATGTTTTAATACACCTAATACATATTGTCCTTGTA----GAACAAGTC--AATGCA
                * ** ********  * ** *  *  *  ***** ****    * *   ***    **

BCVspike        TAGGTAGTGGTTCTGGTATAGATGCTGGTTATAAAAATAGTGGTATAGGCACTTGTCCTG
HCVspike        TAGGTAATGGTCCTGGTATAGATGCTGGTTATAAAAATAGTGGTATAGGCACTTGTCCTG
CRCVspike       TAGGTAGTGGTTTTGGTATAGATGCTGGTTATAAAAATAGTGGTATAGGCACTTGTCCTG
HEVspike        TAGGTGGTG---CTGGCACAGGAACTTGTCCTGTAGGCACCACTGTGCGCAAGTGTTTTG
                *****  **    *** * **   ** **  *  *   *    * *  ***  ***  **

BCVspike        CAGGTACTAATTATTTAACTTGTCATAATGCTGCCCAATGTAATTGTTTGTGCACTCCAG
HCVspike        CAGGTACTAATTATTTAACTTGCCATAATGCTGCCCAATGTGATTGTTTGTGCACTCCCG
CRCVspike       CAGGTACTAATTATTTAACTTGTTATAATGCTAACCAATGTGATTGTTTGTGCACTCCAG
HEVspike        CTG---C-AGTTAC--A---------AACGCTACTAAGTGTACTTGCTGGTGTCAACCAG
                * *   * * ***   *         ** ***    * ***  *** * ***    ** *

BCVspike        ACCCCATTACATCTAAATCTACAGGGCCTTATAAGTGCCCCCAAACTAAATATTTAGTTG
HCVspike        ACCCCATTACATCTAAATCTACAGGGCCTTACAAGTGCCCCCAAACTAAATACTTAGTTG
CRCVspike       ACCCTATTTTATCTAAATCTACAGGGCCTTATAAGTGCCCCCAAACTAAATACTTAGTTG
HEVspike        ATCCTTCCACATATAAAGGTGTAAATGCCTGGACTTGTCCGCAATCTAAAGTTTCTATAC
                * **      ** ****  *  *    * *  *  ** ** *** *****   *   *

BCVspike        GCATAGGTGAGCACTGTTCGGGTCTTGCTATTAAAAGTGATTATTGTGGAGGTAATCCTT
HCVspike        GCATAGGTGAGCACTGTTCGGGTCTTGCTATTAAAAGTGATTATTGTGGAGGTAATCCTT
CRCVspike       GCATAGGTGAGCACTGTTCTGGTCTTGCTATTAAAAGTGATTATTGTGGAGGCAATCCTT
HEVspike        AACCAGGTCAGCATTGCCCTGGCTTGGGTCTTGTGGAGGATGATTGCTCTGGTAATCCTT
                    **** **** **  * **  * * * **     *** ****    ** *******

BCVspike        GTACTTGCCAACCACAAGCATTTTTGGGTTGGTCTGTTGATTCTTGTTTACAAGGGGATA
HCVspike        GTACTTGCCAACCACAAGCATTTTTGGGTTGGTCTGTTGACTCTTGTTTACAAGGGGATA
CRCVspike       GTACTTGCCAACCAAAAGCATTTTTGGGTTGGTCTGTGGACTCTTGTTTACAAGGGGATA
HEVspike        GCACTTGTAAACCACAGGCTTTCATAGGCTGGAGTTCAGAAACTTGTTTGCAAAATGGTA
                * *****  ***** * ** **  * ** ***  *   **  ******* ***   * **
```

FIGURE 9 (Page 6 of 12)

```
BCVspike        GGTGTAATATCTTTGCTAATTTTATTTTGCATGATGTTAATAGTGGTACTACTTGTTCTA
HCVspike        GGTGTAATATTTTTGCTAATTTTATTTTGCATGATGTTAATAGTGGTACTACTTGTTCTA
CRCVspike       GGTGTAATATTTTTGCTAATTTTATTTTGCATGGTGTTAATAGTGGTACTACTTGTTCTA
HEVspike        GGTGTAATATTTTTGCTAATTTTATTTTGAATGATGTTAATAGCGGTACTACCTGTTCTA
                ********** ****************** *** ********* ******** *******

BCVspike        CTGATTTACAAAAATCAAACACAGACATAATTCTTGGTGTTTGTGTTAATTATGATCTTT
HCVspike        CTGATTTACAAAAATCAAACACAGACATAATTCTTGGTGTTTGTGTTAATTATGATCTTT
CRCVspike       CTGATTTACAAAAATCAAACACAGACATAATTCTTGGTGTTTGTGTTAATTATGATCTTT
HEVspike        CTGATTTACAACAGGGTAATACTAATATTACTACTGATGTTTGTGTTAATTATGACCTAT
                *********** *    ** **  * ** * *  ** ****************** ** *

BCVspike        ATGGTATTACAGGCCAAGGTATTTTTGTTGAGGTTAATGCGACTTATTATAATAGTTGGC
HCVspike        ATGGTATTACAGGCCAAGGTATTTTTGTTGAGGTTAATGCGCCTTATTATAATAGTTGGC
CRCVspike       ATGGTATTACAGGCCAAGGTATTTTTGTTGAGGTTAATGCGACTTATTATAATAGTTGGC
HEVspike        ATGGCATTACAGGCCAGGGCATACTTATAGAAGTTAATGCCACGTATTATAATAGTTGGC
                **** ********** ** **  ** * ** ********  * ****************

BCVspike        AGAACCTTTTATATGATTCTAATGGTAATCTCTATGGTTTTAGAGACTACTTAACAAACA
HCVspike        AGAACCTTTTATATGATTCTAATGGTAATCTCTATGGTTTTAGAGACTACTTAACAAACA
CRCVspike       AGAACCTTTTATATGATTCTAATGGTAATCTCTATGGTTTTAGGGACTACTTAACAAACA
HEVspike        AGAATCTTCTTTATGATTCTAGTGGTAATCTCTATGGCTTTAGAGATTATTTATCAAATA
                **** *** * ********** *************** ***** ** ** *** **** *

BCVspike        GAACTTTTATGATTCGTAGTTGCTATAGCGGTCGTGTTTCAGCGGCCTTTCATGCTAATT
HCVspike        GAACTTTTATGATTCGTAGTTGCTATAGCGGTCGTGTTTCAGCGGCCTTTCATGCTAACT
CRCVspike       GAACTTTTATGATTCGTAGTTGCTATAGCGGTCGTGTTTCAGCGGGCTTTCACTCTAACT
HEVspike        GAACCTTTCTTATTCGTAGCTGCTATAGTGGAAGAGTTTCAGCAGTCTTTCATGCTAACT
                **** *** * ******** ******** **  * ******** * ******  **** *

BCVspike        CTTCCGAACCAGCATTGCTATTTCGGAATATTAAATGCAATTACGTTTTTAATAATACTC
HCVspike        CTTCCGAACCAGCATTGCTATTTCGGAATATTAAATGCAGTTACGTTTTTAATAATACTC
CRCVspike       CTTCCGAACCAGCATTGCTATTTCGGAATATTAAATGCAATTACGTTTTTAATAATACTC
HEVspike        CTTCTGAACCAGCTTTGATGTTTCGTAATCTTAAATGCAGCCACGTTTTTAATTATACCA
                **** ******** *** * ***** *** *********   ********** ****
```

FIGURE 9 (Page 7 of 12)

```
BCVspike      TTTCACGACAGCTGCAACCTATTAACTATTTTGATAGTTATCTTGGTTGTGTTGTCAATG
HCVspike      TTTCACGACAGCTGCAACCTATTAACTATTTTGATAGTTATCTTGGTTGTGTTGTCAATG
CRCVspike     TTTCACGACAGCTGCAACCTATTAACTATTTTGATAGTTATCTTGGTTGTGTTGTCAATG
HEVspike      TTTTAAGACAAATACAGCTTGTTAATTATTTTGATAGTTACCTTGGTTGTGTTGTTAATG
              *** * ****  * ** * * **** ************** ************** ****

BCVspike      CTGATAATAGTACTTCTAGTGCTGTTCAAACATGTGATCTCACAGTAGGTAGTGGTTACT
HCVspike      CTGATAATAGTACTTCTAGTGTTGTTCAAACATGTGATCTCACAGTAGGTAGTGGTTACT
CRCVspike     CTGATAATAGTACTTCTAGTTCTGTTCAAACATGTGATCTCACAGTAGGTAGTGGTTACT
HEVspike      CTTATAATAATACAGCTAGTGCTGTAAGTACTTGTGATTTAACCGTTGGTAGCGGCTATT
              ** ****** ***  *****  ***    ** ****** * ** ** ***** ** ** *

BCVspike      GTGTGGATTACTCTACAAAAAGACGAAGTCGTAGAGCGATTACCACTGGTTATCGGTTTA
HCVspike      GTGTGGATTACTCTACAAAAAGACGAAGTCGTAGAGCGATTACCACTGGTTATCGGTTTA
CRCVspike     GGGGGGATTACTCTACACAAAGACGAAGTCGTAGAACGATTACCACTGGTTATCGGTTTA
HEVspike      GTGTTGATTATGTTACAGCACTTAGATCACGTAGATCTTTTACTACAGGTTATCGCTTTA
              * *  *****   ****  *    **   ****** *  **** ** ******** ****

BCVspike      CTAATTTTGAGCCATTTACTGTTAATTCAGTAAATGATAGTTTAGAACCTGTAGGTGGTT
HCVspike      CTAATTTTGAGCCATTTACTGTTAATTCAGTAAATGATAGTTTAGAACCTGTAGGTGGTT
CRCVspike     CTAATTTTGAGCCATTTACTGTTAATCCAGTAAATGATAGTTTACACCCTGTAGGTGGTT
HEVspike      CTAATTTTGAACCATTTGCCGCTAATTTGGTAAATGATAGTATAGAACCTGTTGGTGGTT
              ********** ****** * * ***   ************ ** * ***** *******

BCVspike      TGTATGAAATTCAAATACCTTCAGAGTTTACTATAGGTAATATGGAGGAGTTTATTCAAA
HCVspike      TGTATGAAATTCAAATACCTTCAGAGTTTACTATAGGTAATATGGAGGAGTTTATTCAAA
CRCVspike     TGTATGAAATTCAAATACCTTCAGAGTTTACTATAGGTAATATGGAGGAGTTTATTCAAA
HEVspike      TGTATGAAATACAGATACCTTCAGAGTTTACCATTGGTAATTTAGAAGAATTCATTCAAA
              ********** ** ***************** ** ****** * ** ** ** *******

BCVspike      TAAGCTCTCCTAAAGTTACTATTGATTGTTCTGCTTTTGTCTGTGGTGATTATGCAGCAT
HCVspike      CAAGCTCTCCTAAAGTTACTATTGATTGTTCTGCTTTTGTCTGTGGTGATTATGCAGCAT
CRCVspike     CAAGATCTCCTAAAGTTACTATTGATTGTCCTGTTTTTGTCTGTGGTGATTATGCAGCAT
HEVspike      CGAGTTCCCCTAAGGTTACTATAGATTGTGCTACATTTGTTTGTGGTGACTATGCTGCAT
                ** ** ***** ******** ****** **   ***** ******** ***** ****
```

FIGURE 9 (Page 8 of 12)

```
BCVspike      GTAAATCACAGTTGGTTGAATATGGTAGTTTCTGTGACAATATTAATGCTATACTCACAG
HCVspike      GTAAATCACAGTTGGTTGAATATGGTAGCTTCTGTGACAATATTAATGCTATACTCACAG
CRCVspike     GTAAATCACAGTTGGTTGAATATGGTAGTTTTTGTGACAATATTAATGCTATACTCACAG
HEVspike      GTAGACAACAGTTAGCTGAGTATGGTAGTTTTTGTGAGAACATTAATGCTATACTCATAG
              *** *  ****** * *** ******** ** ***** ** ********** **

BCVspike      AAGTAAATGAACTACTTGACACTACACAGTTGCAAGTAGCTAATAGTTTAATGAATGGTG
HCVspike      AAGTAAATGAACTACTTGACACTACACAGTTGCAAGTAGCTAATAGTTTAATGAATGGTG
CRCVspike     AAGTAAATGAACTACTTGACACTACACAGTTGCAAGTAGCTAATAGTTTAATGAATGGTG
HEVspike      AAGTAAATGAACTACTTGACACTACACAGTTGCAAGTAGCTAATAGTTTAATGAATGGAG
              ********************************************************* *

BCVspike      TCACTCTTAGCACTAAGCTTAAAGATGGCGTTAATTTCAATGTAGACGACATCAATTTTT
HCVspike      TCACTCTTAGCACTAAGCTTAAAGATGGCGTTAATTTCAATGTAGACGACATCAATTTTT
CRCVspike     TCACTCTTAGCACTAAGCTTAAAGATGGCTTTAATTTCAATGTAGATGACATCAATTTTT
HEVspike      TCACCCTTAGTACTAAGATTAAGGATGGGATTAATTTCAATGTTGACGATATCAACTTCT
              **** ***** ****** **** *****  ********** ** ** ***** ** *

BCVspike      CCCCTGTATTAGGTTGTTTAGGAAGCGATTGTAATAAAGTTTCCAGTAGATCTGCTATAG
HCVspike      CCCCTGTATTAGGTTGTTTAGGAAGCGCTTGTAATAAAGTTTCCAGCAGATCTGCTATAG
CRCVspike     CCCCTGTATTAGGTTGTTTAGGAAGCGAATGTAATAAAGTTTCCAGTAGATCTGCTATAG
HEVspike      CCTCTGTATTAGGTTGTTTAGGAAGCGAATGTAACAGAGCTTCCACTAGATCTGCTATAG
              ** ************************  ***** * ** *****  **********

BCVspike      AGGATTTACTTTTTTCTAAAGTAAAGTTATCTGATGTCGGTTTTGTTGAGGCTTATAATA
HCVspike      AGGATTTACTTTTTTCTAAAGTAAAGTTATCTGATGTCGGTTTCGTTGAGGCTTATAATA
CRCVspike     AGGATTTACTTTTTTCTAAAGTAAAGTTATCTGATGTTGGTTTTGTTGATGCTTATAATA
HEVspike      AGGATTTACTTTTTGATAAAGTAAAATTGTCTGATGTCGGTTTTGTACAGGCCTATAATA
              ***************  ********* ** ******** ***** **  * ** *******

BCVspike      ATTGTACTGGAGGTGCCGAAATTAGGGACCTCATTTGTGTGCAAAGTTATAATGGTATCA
HCVspike      ATTGTACTGGAGGTGCCGAAATTAGGGACCTCATTTGTGTGCAAAGTTATAATGGTATCA
CRCVspike     ATTGTACTGGAGGTGCCGAAATTAGGGACCTCATTTGTGTGCAAAGTTATAATGGTATCA
HEVspike      ACTGCACTGGAGGAGCCGAAATTAGGGATCTCATTTGTGTGCAAAGTTATAATGGTATCA
              * ** ******** ********** ********************************
```

FIGURE 9 (Page 9 of 12)

```
BCVspike      AAGTGTTGCCTCCACTACTCTCAGAAAATCAGATCAGTGGATACACTTTGGCTGCTACCT
HCVspike      AAGTGTTGCCTCCACTGCTCTCAGTAAATCAGATCAGTGGATACACTTTGGCTGCCACCT
CRCVspike     AAGTGTTGCCTCCACTGCTCTCAGAAAATCAGATCAGTGGATACACTTTGGCTGCCACCT
HEVspike      AAGTGTTGCCTCCATTGTTATCTGAAAATCAGATTAGTGGTTACACTTCGGCAGCCACCG
              ************** *  * ** * ********* ***** ******* *** ** ***

BCVspike      CTGCTAGTCTGTTTCCTCCTTGGTCAGCAGCAGCAGGCGTACCATTTTATTTAAATGTTC
HCVspike      CTGCTAGTCTGTTTCCTCCTTGGTCAGCAGCAGCAGGTGTACCATTTTATTTAAATGTTC
CRCVspike     TTGCTAGTCTGTTTCCTCCTTGGTCAGCAGCAGCAGGCGTACCATTTTATTTAAATGTTC
HEVspike      CTGCTAGCCTATTTCCTCCCTGGACAGCTGCAGCAGGTGTACCATTTTATTTAAATGTTC
               ****** ** ******** *** **** ******** **********************

BCVspike      AGTATCGTATTAATGGGATTGGTGTTACCATGGATGTTCTAAGTCAAAATCAAAAGCTTA
HCVspike      AGTATCGTATTAATGGGATTGGTGTTACCATGGATGTGTTAAGTCAAAATCAAAAGCTTA
CRCVspike     AGTATCGTATTAATGGTATTGGTGTTACCATGGATGTGCTAACTCAAAATCAAAAGCTTA
HEVspike      AGTATCGTATAAATGGGCTTGGCGTCACCATGGATGTGCTAAGCCAAAACCAAAAGCTTA
              ********** *****  **** ** **********  ***  ***** **********

BCVspike      TTGCTAATGCATTTAACAATGCCCTTGATGCTATTCAGGAAGGGTTTGATGCTACCAATT
HCVspike      TTGCTAATGCATTTAGCAATGCTCTTGATGCTATTCAGGAAGGGTTTGATGCTACCAATT
CRCVspike     TTTCTAATGCATTTAACAATGCCCTTGATGCTATTCAGGAAGGGTTTGATGCTACCAATT
HEVspike      TTGCTAGTGCATTTAACAACGCTCTTGATTCTATCCAGGAAGGGTTCGACGCAACCAATT
              ** *** ******** *** ** ****** **** ********** ** ** *******

BCVspike      CTGCTTTAGTTAAAATTCAAGCTGTTGTTAATGCAAATGCTGAAGCTCTTAATAACTTAT
HCVspike      CTGCTTTAGTTAAAATTCAAGCTGTTGTTAATGCAAATGCTGAAGCTCTTAATAACTTAT
CRCVspike     CTGCTTTAGTTAAAATTCAAGCTGTTGTTAATGCAAATGCTGAAGCTCTTAATAACTTAT
HEVspike      CTGCTTTAGTTAAAATTCAGGCTGTTGTTAATGCAAATGCTGAAGCACTTAATAACTTAT
              ****************** ************************* *************

BCVspike      TGCAACAACTCTCTAATAGATTTGGTGCTATAAGTTCTTCTTTACAAGAAATTCTATCTA
HCVspike      TGCAACAACTCTCTAATAGATTTGGTGCTATAGGTTCTTCTTTACAAGAAATTCTATCTA
CRCVspike     TGCRACAACTCTCTAATAAATTTGGTGCTATAAGTGCTTCTTTACAAGAAATTCTATCTA
HEVspike      TGCAGCAACTCTCTAACAGATTTGGTGCCATAAGTGCCTCTTTACAAGAAATTTTATCCA
              ***  *********** * ********* *** ** * *************** **** *
```

FIGURE 9 (Page 10 of 12)

```
BCVspike      GACTTGATGCTCTTGAAGCGCAAGCTCAGATAGACAGACTTATTAATGGGCGTCTTACCG
HCVspike      GACTGGATGCTCTTGAAGCGCAAGCTCAGATAGACAGACTTATTAATGGGCGTCTTACCG
CRCVspike     GACTTGATGCTCTTGAAGCGCAAGCTCAGATAGACAGACTTATCAATGGGCGTCTTACCG
HEVspike      GGCTCGATGCTCTTGAAGCTAAAGCTCAGATAGACAGACTTATTAATGGGCGTCTCACCG
              * ** **************  ********************** *********** ****

BCVspike      CTCTTAATGCTTATGTTTCTCAACAGCTTAGTGATTCTACACTAGTAAAATTTAGTGCAG
HCVspike      CTCTTAATGCTTATGTTTCTCAACAGCTTAGTGATTCTACACTAGTAAAATTTAGTGCAG
CRCVspike     CTCTTAATGCTTATGTTTCTCAACAGCTTAGTGATTCTACACTAGTAAAATTTAGTGCAG
HEVspike      CTCTTAATGCTTATGTTTCTCAGCAGCTTAGTGATTCTACACTAGTAAAATTTAGTGCAG
              ********************** ************************************

BCVspike      CACAAGCTATGGAGAAGGTTAATGAATGTGTCAAAAGCCAATCATCTAGGATAAATTTTT
HCVspike      CACAAGCTATGGAGAAGGTTAATGAATGTGTCAAAAGCCAATCATCTAGGATAAATTTTT
CRCVspike     CACAAGCTATGGAGAAGGTTAATGAATGTGTCAAAAGCCAATCATCTAGGATAAATTTTT
HEVspike      CACAAGCTATTGAGAAAGTTAATGAATGTGTTAAAAGCCAATCATCTAGGATAAATTTCT
              ********** ***** ************** ************************** *

BCVspike      GTGGTAATGGTAATCATATTATATCATTAGTGCAGAATGCTCCATATGGTTTGTATTTTA
HCVspike      GTGGTAATGGTAATCATATTATATCATTAGTGCAGAATGCTCCATATGGTTTGTATTTTA
CRCVspike     GTGGTAATGGTAATCATATTATATCATTAGTGCAGAATGCTCCATATGGTTTGTATTTTA
HEVspike      GTGGTAATGGTAATCATATTATATCATTAGTACAGAATGCTCCATATGGTTTGTATTTTA
              ****************************** ****************************

BCVspike      TCCACTTTAGCTATGTCCCTACTAAGTATGTCACTGCGAAGGTTAGTCCCGGTCTGTGCA
HCVspike      TCCACTTTAGCTATGTCCCTACTAAGTATGTCACTGCGAAGGTTAGTCCCGGTCTGTGCA
CRCVspike     TCCACTTTAGCTATGTCCCTACTAAGTATGTCACTGCGAAGGTTAGTCCCGGTCTGTGCA
HEVspike      TCCATTTTAGCTATGTCCCCACCAAGTATGTTACAGCAAAGGTTAGTCCTGGTTTGTGCA
              **** ************** ** ******** ** ** *********** *** ******

BCVspike      TTGCTGGTGATAGAGGTATAGCCCCTAAGAGTGGTTATTTTGTTAATGTAAATAACACTT
HCVspike      TTGCTGGTGATAGAGGTATAGCCCCTAAGAGTGGTTATTTTGTTAATGTAAATAATACTT
CRCVspike     TYGCAGGTGATAGAGGTATAGCTCCTAAGAGTGGTTATTTTGTTAATGTAAATAACACTT
HEVspike      TTGCTGGCGATATAGGAATATCGCCTAAGAGTGGTTATTTTATTAATGTAAATAACTCTT
              * ** ** **** *** *** * ****************** *************  ***
```

FIGURE 9 (Page 11 of 12)

```
BCVspike      GGATGTTCACTGGTAGTGGTTATTACTACCCTGAACCTATAACTGGAAATAATGTTGTTG
HCVspike      GGATGTTCACTGGTAGTGGTTATTACTACCCTGAACCCATAACTGGAAATAATGTTGTTG
CRCVspike     GGATGTTCACTGGTAGTGGTTATTACTACCCTGAACCTATAACTGGAAATAATGTGGTTG
HEVspike      GGATGTTCACTGGTAGTGGCTATTACTACCCTGAACCTATAACCCAAAATAATGTTGTTG
              ******************* ***************** *****   ********* ****

BCVspike      TTATGAGTACCTGTGCTGTTAATTACACTAAAGCACCGGATGTAATGCTGAACATTTCAA
HCVspike      TTATGAGTACCTGTGCTGTTAACTATACTAAAGCGCCGGATGTAATGCTGAACATTTCAA
CRCVspike     TTATGAGTACCTGTGCTGTTAACTATACTAAAGCACCGGATGTAATGCTGAACATTTCAA
HEVspike      TGATGAGTACGTGTGCTGTTAATTATACTAAAGCACCGGATCTAATGCTGAACACATCGA
              * ******** *********** ** ******** ****** ************  ** *

BCVspike      CACCCAACCTCCCTGATTTTAAGGAAGAGTTGGATCAATGGTTTAAAAACCAAACATCAG
HCVspike      CACCCAACCTCCATGATTTTAAGGAAGAGTTGGATCAATGGTTTAAAAACCAAACATCAG
CRCVspike     CACCCAACCTCCCTGATTTTAAGGAAGAGTTGGATCAATGGTTTAAAAACCAAACATTAA
HEVspike      CACCCAACCTTCCTGATTTCAAGGAAGAATTGTATCAATGGTTTAAAAACCAATCTTCAT
              ********** * ****** ******** *** ******************** * * *

BCVspike      TGGCACCAGATTTGTCACTTGATTATATAAATGTTACATTCTTGGACCTACAAGATGAAA
HCVspike      TGGCACCAGATTTGTCACTTGATTATATAAATGTTACATTCTTGGACCTACAAGATGAAA
CRCVspike     TGGCACCAGATTTGTCACTTGATTATATAAATGTTACATTCTTGGACCTACAAGATGAAA
HEVspike      TGGCACCAGATTTGTCATTTGATTATATTAATGTTACGTTCTTGGACCTACAAGATGAAA
              ***************** ********** ******** **********************

BCVspike      TGAATAGGTTACAGGAGGCAATAAAAGTTTTAAATCAGAGCTACATCAATCTCAAGGACA
HCVspike      TGAATAGGTTACAGGAGGCAATAAAAGTTTTAAATCAGAGCTACATCAATCTCAAGGACA
CRCVspike     TGAATAGGTTACAGGAGGCAATAAAAGTTTTAAATCATAGCTACATCAATCTCAAGGACA
HEVspike      TGAATAGGTTACAAGAAGCTATAAAAGTTCTAAATCATAGCTACATCAATCTCAAGGACA
              ************* ** ** ********* ******* **********************

BCVspike      TTGGTACATATGAGTATTATGTAAAATGGCCTTGGTATGTATGGCTTTTAATTGGCCTTG
HCVspike      TTGGTACATATGAGTATTATGTAAAATGGCCTTGGTATGTATGGCTTTTAATTGGCTTTG
CRCVspike     TTGGTACATATGAATATTATGTAAAATGGCCTTGGTATGTATGGCTTTTAATTGGCCTTG
HEVspike      TTGGTACATATGAGTATTATGTGAAATGGCCTTGGTATGTATGGCTTTTAATTTGCCTTG
              ************* ******** ****************************** ** ***
```

FIGURE 9 (Page 12 of 12)

```
BCVspike        CTGGTGTAGCTATGCTTGTTTTACTATTCTTCATATGCTGTTGTACAGGATGTGGGACTA
HCVspike        CTGGTGTAGCTATGCTTGTTTTACTATTCTTCATATGCTGTTGTACAGGATGTGGGACTA
CRCVspike       CTGGCGTAGCTATGCTTGTTTTACTATTCTTCATATGCTGTTGTACAGGATGTGGGACTA
HEVspike        CTGGTGTAGTTATGCTTGTTTTACTATTCTTCATATGCTGCTGTACAGGATGTGGGACTA
                **** **** ******************************** ******************

BCVspike        GTTGTTTTAAGAAATGTGGTGGTTGTTGTGATGATTATAC--------------------
HCVspike        GTTGTTTTAAGATATGTGGTGGTTGTTGTGATGATTATACTGGACACCAGG---------
CRCVspike       GTTGTTTTAAGAAATGCGGTGGTTGTTGTGATGATTATACTGGACATCAGG---------
HEVspike        GTTGTTTTAAGAAATGTGGCGGTTGTTTTGATGATTATACTGGACACCAGGAGTTTGTAA
                ************ *** ** ******* ************

BCVspike        ------------------------------
HCVspike        ------------------------------
CRCVspike       ------------------------------
HEVspike        TCAAAACTTCACATGACGATTAATTTCGT
```

FIGURE 10 (Page 1 of 5)

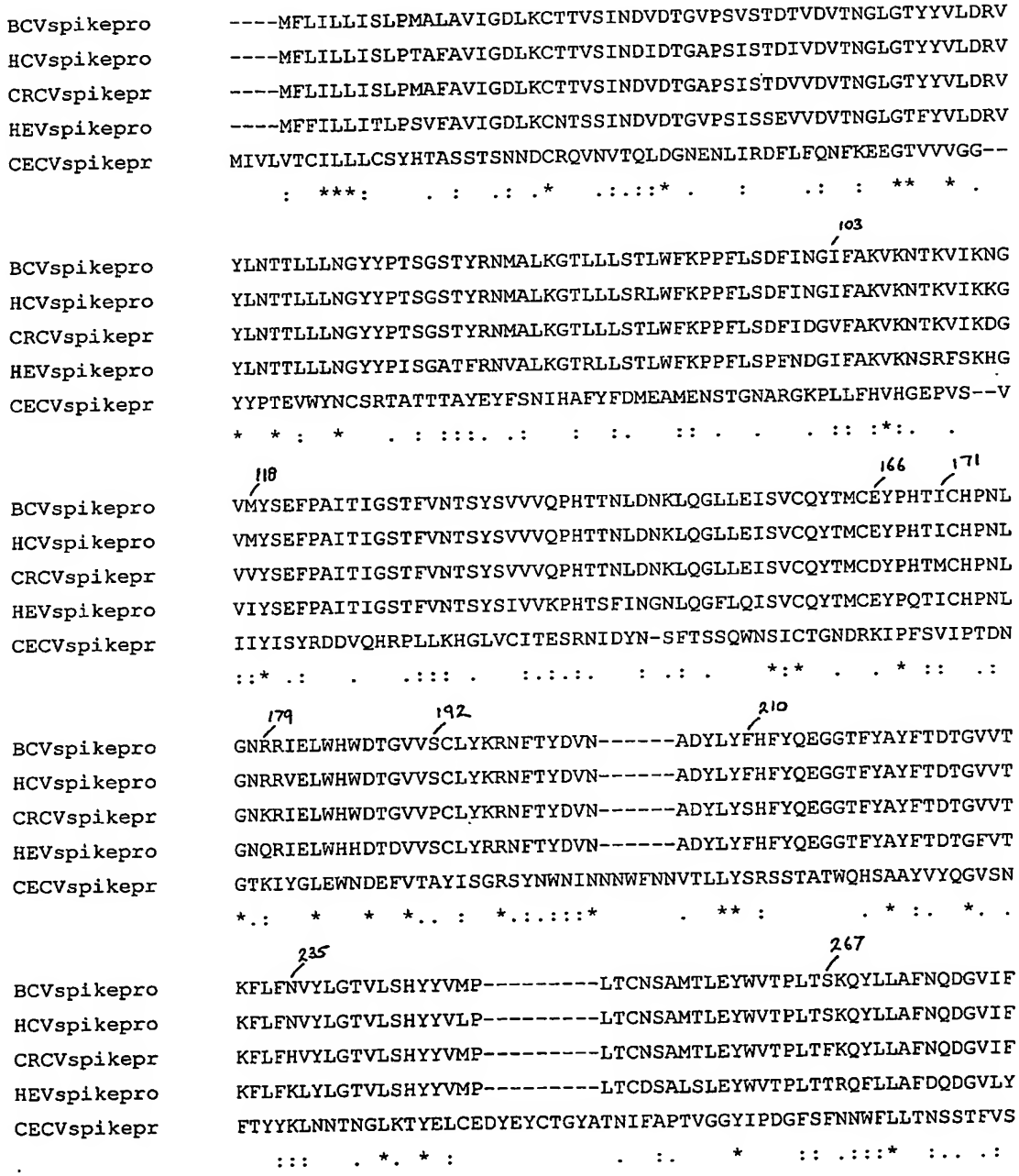


FIGURE 10 (Page 2 of 5)

```
BCVspikepro     NAVDCKSDFMSEIKCKTLSIAPSTGVYELNGYTVQPIADVYRR-IPNLPDCNIEAWLNDK
HCVspikepro     NAVDCKSDFMSEIKCKTLSIAPSTGVYELNGYTVQPIADVYRR-IPNLPDCNIEAWLNDK
CRCVspikepr     NAVDCKSDFMSEIKCKTLSIAPSTGVYELNGYTVQPIADVYRR-IPNLPDCNIEAWLNDK
HEVspikepro     HAVDCASDFMSEIMCKTSSITPPTGVYELNGYTVQPVATVYRR-IPDLPNCDIEAWLNSK
CECVspikepr     GRFVTNQPLLVNCLWPVPSFGVAAQEFCFEGAQFSQCNGVFLNNTVDVIRFNLNFTADVQ
                  :   . :: :    . *:  .:  : ::*  ..    *: .   ::   :::.  : :
                                                                    388
BCVspikepro     SVPSPLNWERKTFSNCNFNMSSLMSFIQADSFTCN------NIDAAKIYGMCFSSITIDK
HCVspikepro     SVPSPLNWERKTFSNCNFNMSSLMSFIQADSFTCN------NIDAAKIYGMCFSSITIDK
CRCVspikepr     SVPSPLNWERKTFSNCNFNMSSLMSFIQADSFTCN------NIDAAKIYGMCFFSITIDK
HEVspikepro     TVSSPLNWERKIFSNCNFNMGRLMSFIQADSFGCN------NIDASRLYGMCFGSITIDK
CECVspikepr     SGMGATVFSLNTTGGCILEISCYNDIVSESSFYSYGEIPFGVTDGPRYCYVLYNGTALKY
                :  ..  :. :  ..* :::.   .::. .** .         *..:   : : . ::.
                          407                                436  440    447
BCVspikepro     FAIPNGRKVDLQLGNLGYLQSFNYRIDTTATSCQLYYNLPAAN-VSVSRFNPSTWNRRFG
HCVspikepro     FAIPNGRKVDLQLGNLGYLQSFNYRIDTTATSCQLYYNLPAAN-VSVSRFNPSTWNRRFG
CRCVspikepr     FAIPNGRKVDLQMGNLGYLQSFNYRIDTTATSCQLYYNLPASN-VSISRFNPSIWNRRFG
HEVspikepro     FAIPNSRKVDLQVGKSGYLQSFNYKIDTAVSSCQLYYSLPAAN-VSVTHYNPSSWNRRYG
CECVspikepr     FGTLPPSVKEIAISKWGQFYINGYNFFSTFPIDCISFNLTTGDSGAFWTIAYTSYTEALV
                *.       :: :.: * :   .*.: :: .   : :.*.:.:  :.     : :..
                                                                   501
BCVspikepro     FTEQSVFKPQPVGVFTDHDVVYAQHCFKAPTNFCPCKLDGSLCVGSGSGIDAGYKNSGIG
HCVspikepro     FTEQSVFKPQPVGVFTHHDVVYAQHCFKAPTNFCPCKLDGSLCVGNGPGIDAGYKNSGIG
CRCVspikepr     FTEQSVFKPQPVGVFTDHDVVYAQHCFKAPTNFCPCKLNGSLCVGSGFGIDAGYKNSGIG
HEVspikepro     FINQS------FGSRGLHDAVYSQQCFNTPNTYCPCRT--SQCIGG----------AGTG
CECVspikepr     QVENTAIKKVTYCNSHINNIKCSQLTANLQNGFYPVASSEVGLVNKSVVLLPSFYSHTSV
                  :::            ::   :*   :  . : *        :.
                          525    528        540
BCVspikepro     TCPAGTNYLTCH----NAAQCNCLCTPDPITSKSTGPYKCPQTKYLVGIGEHCSGLAIKS
HCVspikepro     TCPAGTNYLTCH----NAAQCDCLCTPDPITSKSTGPYKCPQTKYLVGIGEHCSGLAIKS
CRCVspikepr     TCPAGTNYLTCY----NANQCDCLCTPDPILSKSTGPYKCPQTKYLVGIGEHCSGLAIKS
HEVspikepro     TCPVGTTVRKCFAAVTNATKCTCWCQPDPSTYKGVNAWTCPQSKVSIQPGQHCPGLGLVE
CECVspikepr     NITIDLGMKRSGYG--QPIASTLSNITLPMQDNNTDVYCIRSNQFSVYVHSTCKSSLWDN
                . . .    .    :.  .    . *   :... :   ..:  :   . * .    .
```

FIGURE 10 (Page 3 of 5)

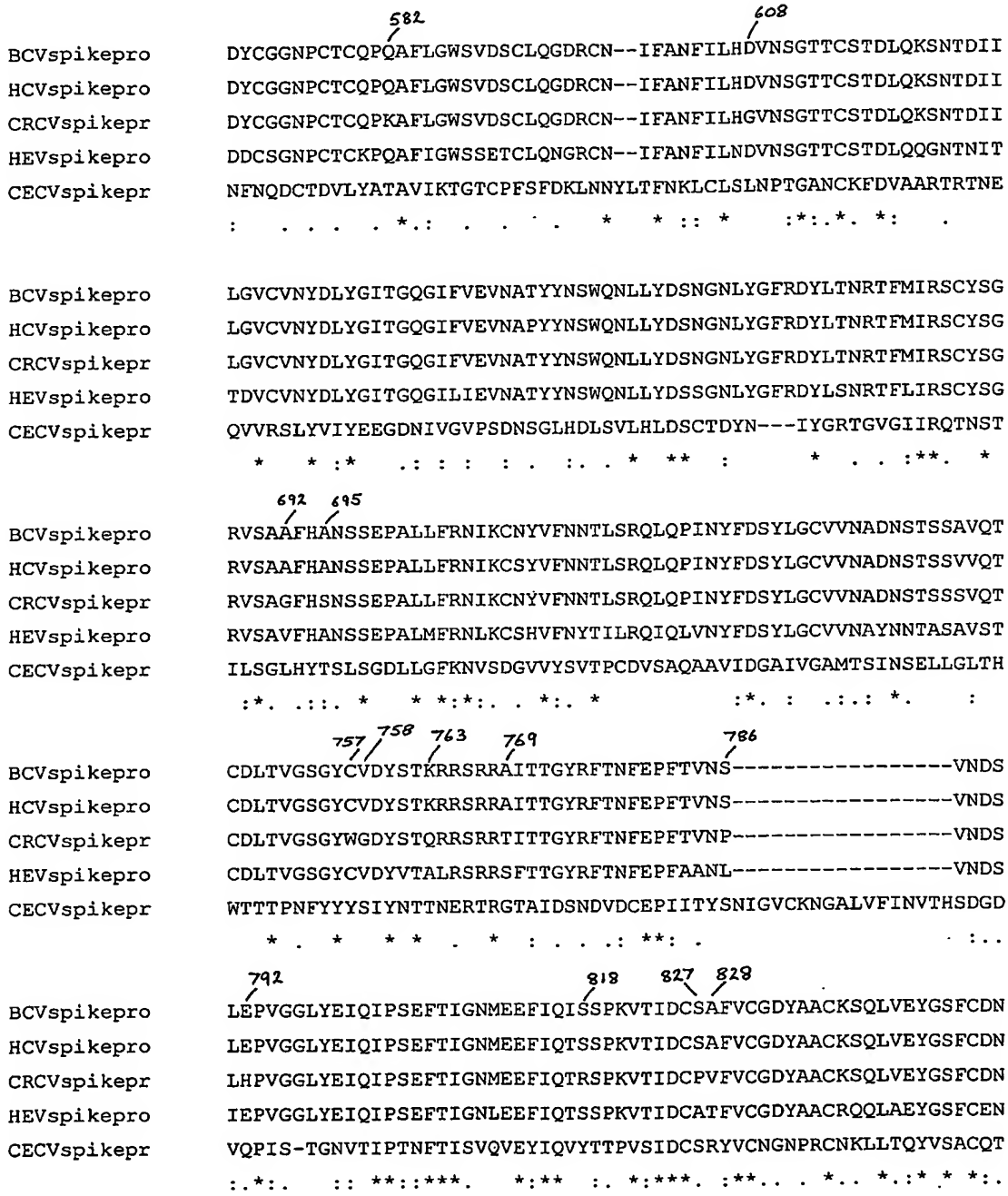


FIGURE 10 (Page 4 of 5)

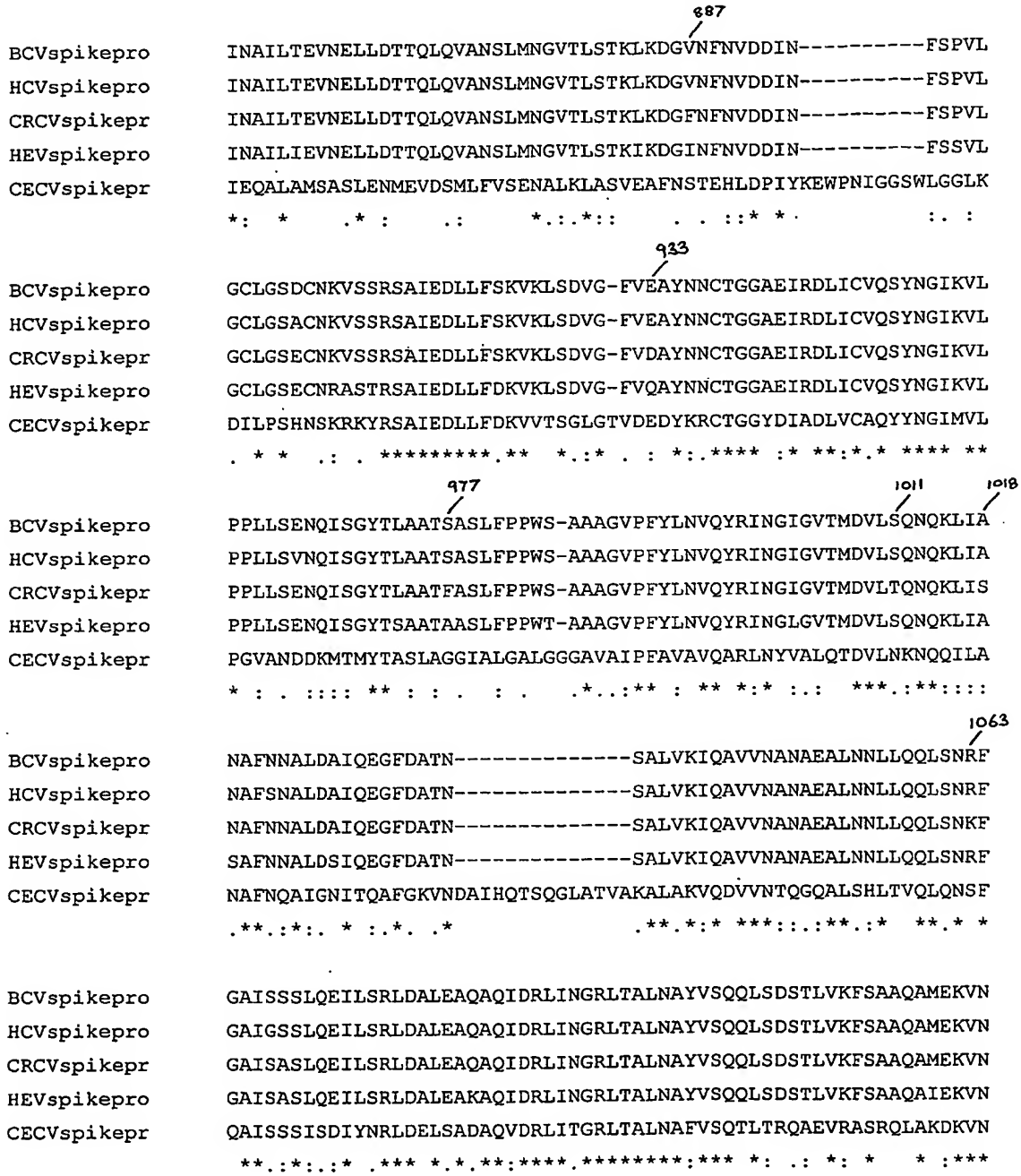


FIGURE 10 (Page 5 of 5)

```
BCVspikepro     ECVKSQSSRINFCGNGNHIISLVQNAPYGLYFIHFSYVPTKYVTAKVSPGLCIAGDRGIA
HCVspikepro     ECVKSQSSRINFCGNGNHIISLVQNAPYGLYFIHFSYVPTKYVTAKVSPGLCIAGDRGIA
CRCVspikepr     ECVKSQSSRINFCGNGNHIISLVQNAPYGLYFIHFSYVPTKYVTAKVSPGLCIAGDRGIA
HEVspikepro     ECVKSQSSRINFCGNGNHIISLVQNAPYGLYFIHFSYVPTKYVTAKVSPGLCIAGDIGIS
CECVspikepr     ECVRSQSQRFGFCGNGTHLFSLANAAPNGMVFFHTVLLPTAYETVTAWSGICASDGDRTF
                ***:***.*:.*****.*::**.: ** *: *:*   :** * *..  .*:* :..

BCVspikepro     ------PKSGYFVNVNNTWMFTGSGYYYPEPITGNNVVVMSTCAVNYTKAPDVMLNISTP
HCVspikepro     ------PKSGYFVNVNNTWMFTGSGYYYPEPITGNNVVVMSTCAVNYTKAPDVMLNISTP
CRCVspikepr     ------PKSGYFVNVNNTWMFTGSGYYYPEPITGNNVVVMSTCAVNYTKAPDVMLNISTP
HEVspikepro     ------PKSGYFINVNNSWMFTGSGYYYPEPITQNNVVVMSTCAVNYTKAPDLMLNTSTP
CECVspikepr     GLVVKDVQLTLFRNLDDKFYLTPRTMYQPRAATSSDFVQIEGCDVLFVNATVIDLPSIIP
                      :   * *::::.: :*    * *.. * .:.* :. * * :.:*. : *    *

                                  1256  1257
BCVspikepro     NLPDFKEELDQWFKNQTS--VAPDLSLDYINVTFLDLQDEMN--------------RLQE
HCVspikepro     NLHDFKEELDQWFKNQTS--VAPDLSLDYINVTFLDLQDEMN--------------RLQE
CRCVspikepr     NLPDFKEELDQWFKNQTL--MAPDLSLDYINVTFLDLQDEMN--------------RLQE
HEVspikepro     NLPDFKEELYQWFKNQSS--LAPDLSFDYINVTFLDLQDEMN--------------RLQE
CECVspikepr     DYIDINQTVQDILENYRPNWTVPELTIDIFNATYLNLTGEIDDLEFRSEKLHNTTVELAI
                :  *::: : : ::*      .*:*::* :*.*:*:* .*::              .*

BCVspikepro     AIKVLNQSYINLKDIGTYEYYVKWPWYVWLLIGLAGVAMLVLLFFICCCTGCGTSCFKKC
HCVspikepro     AIKVLNQSYINLKDIGTYEYYVKWPWYVWLLIGFAGVAMLVLLFFICCCTGCGTSCFKIC
CRCVspikepr     AIKVLNHSYINLKDIGTYEYYVKWPWYVWLLIGLAGVAMLVLLFFICCCTGCGTSCFKKC
HEVspikepro     AIKVLNHSYINLKDIGTYEYYVKWPWYVWLLICLAGVVMLVLLFFICCCTGCGTSCFKKC
CECVspikepr     LIDNINNTLVNLEWLNRIETYVKWPWYVWLLIGLVVVFCIPLLLFCCCSTGCCG-CIGCL
                 *. :*:: :**: :.  * ************ :. *  : **:* **.***   *:

BCVspikepro     GGCCDDYTGHQELVIKTSHDD----------------------------------------
HCVspikepro     GGCCDDYTGHQELVIKTSHDD----------------------------------------
CRCVspikepr     GGCCDDYTGHQELVIKTSHDD----------------------------------------
HEVspikepro     GGCFDDYTGHQEFVIKTSHDD----------------------------------------
CECVspikepr     GSCCHSICSRRQFENYEPIEKVHVH------------------------------------
                *.* ..  .:::::    . :.
```

Figure 11

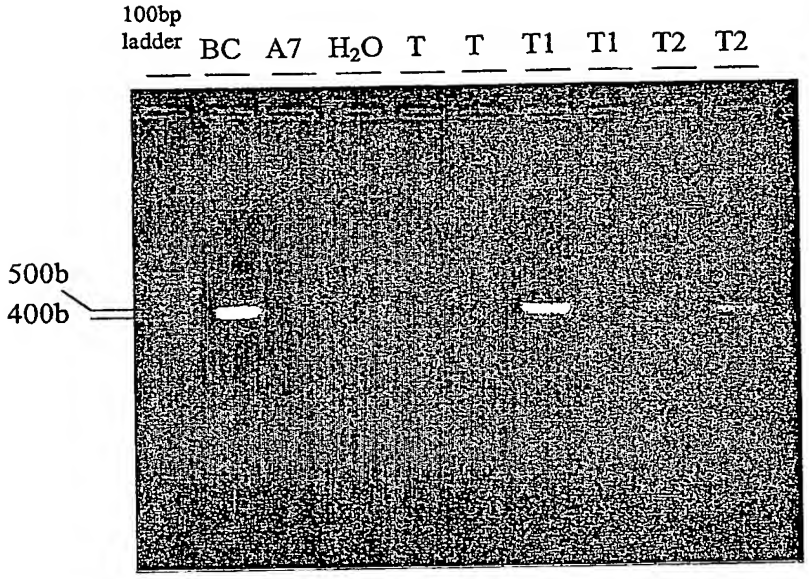
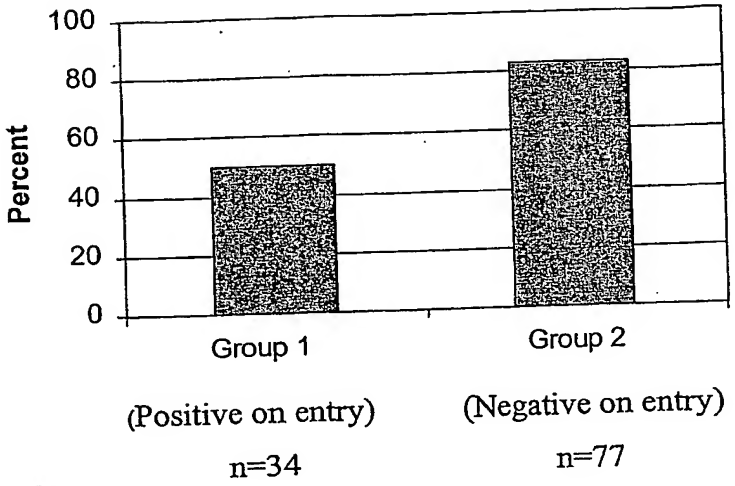


FIGURE 12



(Positive on entry)
n=34

(Negative on entry)
n=77